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REVIEW ARTICLE

ON THE STUDY OF SOIL MICROORGANISM DIVERSITY IN INDIAN CONTEXT

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ABSTRACT

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Key words:

Soil Microorganisms, Diversity, Soil Ecosystem, Morphological and Biochemical Properties. Microorganisms are responsible for different soil ecosystem and biodiversity. Diversity of microorganisms of different area varies both in quality and quantity of microbs. Microorganisms available in biogeochemical cycles, mineralization, and solubilization of organic and inorganic matter in soil vary widely. Symbiotic and non-symbiotic bacteria make colony in plant root, rhizosphere and mycorhiza. Diversity and interaction of microorganisms in plant root and microbial activity in soil has been studied in detail in India and abroad. Present paper deals with such studies and reviews the available literature which may be useful for the researchers and planners equally.

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INTRODUCTION

A global concerns focus on environmental protection and natural resources management. Microbial diversity is responsible for variable reaction in different soil orders. Understanding of soil ecosystem function at the process level is necessary. Ecological approaches have primarily focused on the function of natural ecosystem where sustainability is viewed in terms of the maintenance and stability of ecosystem productivity has not been fully explored because of great microbial biodiversity in soil ecosystem.

Soil ecosystem

Around 1.5billion years ago; motile microorganisms migrated to different environments under the influence of different factors. Soil formation on the earth was started through biomediation but they were existence recently in history, about 300 years with Leeuwenhoek's invention of the microscope. The microbes execute most of the chemical transformation in the soil ecosystem. The thar desert of North Western India is distributed over 2.34 million km with about 91% area endemic to the Rajasthan.

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The low fertile sandy soil of this desert is poor in organic matter, having rapid water infiltration rates and rapid oxidation (Chowdhury, Michael, Anton, and Tripathi, 2007). Desert soil is an excellent ecosystem colonized by various groups of microorganisms with extremities of environment conditions soil microbial communities are among the most complex. diverse, and important assemblages in the biosphere (Zhou et al., 2003). Soil contains billions microorganisms, so many different types that accurate numbers remain unknown. The understanding of the composition of the microbial communities in soilis complex. In 1931, Waksman believed that a large body of information has accumulated that enables us to construct a clear picture of microscopic population of the soil and in 1932 Bergey's manual clarified that no organisms could be classified devoid of being cultured. Soil microorganisms are mostly small and inconspicuous and therefore rarely enjoy public attention in contrast to birds, flowers or other large and eye catching organisms and there is nothing similar for soil microfouna because these seem to be less facitating (Barrios, 2007).

Rhizosphere

Soil microorganisms are included in a wide variety of metabolic and physiological activities that influence the microhabitat. The plant root influcing soil volume where there is a high concentration of the carbon with zone of intense microbial metabolic activity occurring is called the rhizosphere. Higher population of bacteria including Azotobactorchrococcum, Beijerinekia and Actinomycetes from the rhizosphere soil of Mycorrhizal plants (Bagayraj and Mengi, 1978). is observed because of root exudation (Barea et al., 1975) in rhizosphere affects rhizospheric microflora. In general mycorrhizal fungi through modifications to the plant root system, interact with beneficial soil organisms such as N2fixing bacteria, Phosphate-solubilising Bacteria, Fungi and inhabiting nematodes. These interactions are important in the natural ecosystem for nutrient cycling (Gryndler, 2000). A large number of portion of soluble inorganic phosphate applied to agriculture soil as chemical fertilizer is rapidly immobilized soon after application and becomes unavailable to plants (Dadarwal et al., 1997). PSMs (Phosphate solubilizing microorganisms) include different group of microorganisms which not only assimilate phosphorus from insoluble forms of phosphates but they also cause a large portion of soluble phosphate to be released in quantities in excess of their requirements species of Aspergillus and Penicilliumare among fungal isolates identified to have phosphate solubilizing capabilities. Among bacterial genera with this capability are Pseudomona, Azospirillum, Bacillus, Rhizobium, Arthrobacter, Burkholderia, Alcaligenes, Serratia, Enterobacter, Acinetobacter, Flavobacterium and Erwinia (Rodriguez et al., 1996).

Gram positive and gram negative bacteria Arthobactersp. and Pseudomonassp. was greatest in the lowPhosphorus soil but the bacterial population of fertilized P and AM soil was generally not significantly different. They also found that G. mosseae had the lowest gram negative bacterial population while G. etunication soil had the highest population of both gram positive and gram-negative bacteria (Schreiner et al., 1997). Soil microorganisms isolated were identified on the basis of morphological and biochemical properties morphological method consisted of microscopic and microscopic method. The microscopic characterization method (Kawato and Sinobu, 1959). Isolated bacteria Bacillus subtilis, B. Cerus, B. Polymyxa. Agriculture soil Microorganisms isolated were Escharichiacoli, Pseudomonas aeruginosa, Klebsiella pneumoniae, Enterobacter aerogenes, Shigella sp., Proteus Mirabilis, Bacillus anthracis, Bacillus Subtilis, Staphylocococcus aureus and Staphylococcus Epidermidis species, Fungi suchas Aspergillus niger, A. Flavus, Fusarium oxysporum, Trichoderma sp. and Rhizopus sp. (Amir and Pineau, 1998). Microorganisms play an important role in biodegradation of soil agriculture waste and increasing the soil Fertility and improvement of crop production (Okoh et al., 1999).

Agriculture soil is a dynamic medium in which a large number of pathogenic and non- pathogenic bacterial and fungal flora lives in close association. Microbes in the soil are the key to carbon and nitrogen recycling. Microorganisms produce some useful compound that are beneficial to soil health, plant growth and play an important role in nutritional chains that are important part of the biological balance in the life in our planet (Paul and Clerk, 1966; Kummerer, 2004). Microbial activity in desert soil is highly dependent on characteristics such as temperature, moisture and the availability of organic carbon (Buyanovsky *et al.*, 1982, Parker *et al.*, 1984). Of these, moisture availability is the major constraint affecting microbial diversity, community structure and activity. Gram-positive spore formers and dominant and the population do not decline significantly even during summers (Rao *et al.*, 1983). Actinomycetes may constitute around 50% of the total microbial population in desert soils. However, Hethener reported only 1-2% representation of Actinomycetes in the sandy soil of Tassili N, Ajjer (Hethener, 1967). Dominant microflora desert soil is made up of Coryneforms i.e. Cystobacter, Myxococcus, Polvangium, Archangium, Sorangium and Stigmatella; Sub-dominant forms comprise Acinetobacter. Bacillus, Micrococcus. Proteus and Pseudomonas, Cvanobacteria also contribute significantly to the biota of hot arid regions in terms of primary productivity and nitrogen fixation (Bhatnagar et al., 2003). The dominant Cynobacterial forms of Thar Desert are chroococcusminutes, Oscillatoria Pseudogeminata and Phormidiumtenue, Nostoc dominants amongst heterocystous forms. Fungal sp. populations as viable propagate range from nil to 6.3x103 in Uzboi desert Takyr (Lobova et al., 1967; Cameron, 1969).

The Dominant genera include Aspergillus, Curvularia, Fusarium, Mucor, Paecilomyces, Penicillium, Phoma and Stemphylium, XericMushrooms such as Coprinus, Fomes, Terfezia and Termania have also been reported from desert (Trappe, 1981). The removal of zinc from soil polluted by effluents from textile industries was studies using two fungi Strands Aspergillus Fumigatus RH05 and Aspergillus Flavus RH07 (Goldstein et al., 1997). Evidence of the involvement of microorganisms in solubilization of inorganic phosphates was reported as early as 1903 (Kucey et al., 1989, Khan et al., 2007). Phosphate solubilizing microorganisms (PSM) are ubiguitous and their numbers vary from soil to soil. Population of (PSM) and organic matter content of some selected arid soils of Rajasthan, India was reported by (Venkateswarul et al., 1984). Phosphorous solubilization is carried out by a large number of Saprophytic bacteria and fungi acting on sparingly soluble soil phosphate, mainly by chelating-mediated mechanism (Bajpai and Rao, 1971, Moghimi et al., 1978, Whitelaw 2000).

In general among the whole microbial population in soil phosphatsolubilizing fungi 0.1to0.5% of the total respective population (Chem *et al.*, 2006). Phosphate solubilizing bacteria generally out number phosphat- solubilizing fungi by 2-150 flads (Banik and Dey 1982, Kucey,1983, Kucey *et al.*, 1989, Alam *et al.*, 2002). The efficiency of different phosphatase and phytase secreating fungi isolated from arid soils of Rajasthan, India to hydrolyse different organic phosphatecompounds were reported (Tarafdar *et al.*, 2003. Aseri *et al.*, 2009).

Mineralization of organic Phosphate

Mineralization of soil organic phosphate plays an imperative role in Phosphorus cycling of a farming system. Mineralization of organic to inorganic phosphate involves processes Catalyzed by phosphatases use organic phosphate as a substrate to convert it into inorganic form (Beech et. al. 2001). Some heterotropic microorganisms are also capable of solubilizing phosphates combined with calcium or magnesium (Atlas and Bartha 1998). These soluble forms can now be readily taken up by plants, algae, cynobacteria and autotrophic bacteria and assimilated into organic cellular components such as DNA, RNA, and ATP. Phosphatase enzyme is present in all organisms but only bacteria, fungi, and some algae are able to secrete the outside of their cell (Jones, 2002). Soil microorganisms such as Bacteria, yeast, Fungi, could produce pectinases (Yadav et al., 2009). Extracellular enzymes are produced by different varieties of microorganisms i.e. fungi,

bacteria, yeast and Actinomycetes (Devi et al., 2008). Bacteria are on the top of the list of alkaline protease produces. The genus Bacillus is the most common source of proteases (Gupta et al., 2002). Some example of potent alkaline protease producing Bacilli strain are B. amyloliquifuciens, B. subtills and B. Lichiniformis. Some other bacterial sp. which is also known for their protease production potential are Staphylococcus. Flababacterium, Serratia, Alcaligenes. Vibrio, Brevibacterium, Pseudomonas, and Halobacterium, (Gupta et al., 2005). Among Actinomycetes strain of Nocardiopsis, Streptomyces and Nocardia are potential ones. In fungi, Aspergili is the most predominant group for protease production and the strain of Ophiostoma, Myxococcus, Penicillium, Rhizopus, and Neurospora, etc are common produces of proteases (Gupta et al., 2005).

Conclusion

Microorganisms are playing important role in soil ecosystem and different industrial use. Soil provides wide habitat for different microbial activity is highly dependent on its characteristics. This is the major constraint affecting microbial diversity. On the basis of above account it can be concluded that in spite of much research done in this field, there are certain gaps in terms of ecosystems (including microecosystems), species diversity and microbe-plant interactions. This field provides ample scope to investigate deeply in this area.

REFERENCES

- Alam S. et al. 2002. In vitro solubilization of inorganic phosphate by phosphate solubilizing microorganisms (PSMs) for Mize Rhizosphere. International J. of Agriculture and Biology, Vol. 4, No. 4 (December 2002), pp. 454-458. ISSN1560-8530.
- Amir H., Pineau R. 1998. Influence of plant and cropping on microbiological characteristics of some new Caledonian Ultramafic soils. *Aust. J. soil Res.*, 36: 457-470.
- Aseri G. K. *et al.* 2009. Hydrolysis of organic phosphate forms by phosphatase and some phytase producing fungi of arid and semi arid soils of India. *American – Eurasian journal* of Agriculture and Environment science, vol. 5, No.4, (December 2009), pp. 564-570, ISSN 1818-6768.
- Atlas R. M. and Bartha R. 1988.Microbial Ecology. Fundamentals and Application, *Benjamin Cummings Publishing Company Ine,JSBN* 10:0805306563, USA.
- Bagyaraj D.J. and Menge J.A. 1978. Interaction between VA-Mycorrhiza and Azatobacter and Plant growth. New phytologist 80:561-573.
- Bajpai P.D. and Sundra Rao, W.V.B. 1971. Phosphate solubilizing bacteria part 2. Extracellular production of organic acid by selected bacteria solubilizing insoluble phosphate. Soil science and nutrition, vol. 17, No. 2, (April 1971), pp. 544-45, ISSN 1747-0765.
- Banik S. and Day B. K. 1982. Available phosphate content of an Alluvial soil as influenced by inoculation, of some isolated phosphate solubilizing microorganisms. *Plant and soil*, Vol.69 No.3, (September 1982), pp. 353-364, ISSN 1573-5036.
- Barea J. M. et al. 1975 Endo mycorrhizas, Academic press London.
- Beech I. B. et. al.2001. Enzymatic activity and within biofilms of sulphate-Reducing bacteria in biofilm community interactions: change or necessity? P.G. Gilbert D. Allison

M. Brading., J. Verran and J. Walker (Eds) 231-239 Boiline, ISBN 9780521813405 Cardiff, U.K.

- Bhatnagar A. *et al.* 2003. Satellite center for micro algal Biodiversity in arid zone of Rajasthan Project completion report, *Funded by Department of Biotechnology*, New delhi.2003.
- Buyanovsky G. et al. 1982. Soil Environment and activity of soil microflora in the Negev desert J. Arid Environment 5, 13-28.
- Cameron R.E. 1969. Cold desert characteristics and problems related toother arid lands. In arid lands in perspective (eds Mc Ginnies, W.G. and Goldman. B. J.) University of Arizona press, Tucson, pp. 167-205.
- Chen Y.P. *et al.* 2006. Phosphate solubilizing bacteria from subtropical soil and their Tricalcium phosphate solubilizing abilities. Applied soil ecology. Vol 34, No.1, (November 2006), pp. 33-4, ISSN 0929-1393.
- Choudhary A. K. *et al.* 1984. Present status of the geochronology of the Precambrian rocks of Rajasthan. Tectonophysics. 105:131-140.
- Chowdhary S.P., Michael S., Anton H. and Tripathi A.K. 2007. Identification of diazotrophs in the cultureable bacterial community associated with roots Lasiurus sindicus, a perennial grass of the desert, India. Microbial Ecology, 54, 82-90, doi: 10, 1007/ s0038-006, 9174-1.
- Dadarwal, Yadav K.S. and K.r.1997 Phosphate solubilization and mobilization through soil microorganisms. In: biotechnology approaches in soil microorganisms for sustainable crop production (Dadarwal, K.R., Ed.) pp.293-308. Scientific publisher, Jodhpur.
- Doco T, Williams P. *et al.* 1997. Carbohydrate Research, Vol.297 No.2, p 181.
- Goldstein N. D, Block, "Nation Wide Inventry of Food Residuals Composting" *Biocycle Journal of Composting and Recycling*, August, 1997, pp.46-47.
- Gryndler, M. 2000. Interactions of arbuscular mycorrhizas. Physiology and function. Edited by Y.
- Gupta R, *et al.* 2002. An overview on jermentation, downstream processing and properties of microbial alkaline protease. *Appl. of Microbiology, Biotechnology.* 60: 381-395.
- Gupta R. 2005. Bacterial alkaline proteases: Resent trends and industrial application. In microbial diversity current perspectives and potential application, Satyanarrayana T. and johri BN, eds., IK International Pvt.Ltd.
- Hethener P.1967. Active microbiologique des sols a Cupressus dupresiana. A Camus an Tasal. N' ajjer (Sahara central) Bull. Soc. Hist. Nat. Afr. Nord 58, 39-100.
- Khan E.A. and Sogni pc. Stratigraphy and sedimentation of Trans-Aravalli Vindhyans of Western Rajasthan.
- Khan M.S. *et al.* 2007. Role of phosphate solubilizing microorganisms in sustainable agriculture a review. Agronomy for sustainable development, vol.27, No.1 (March 2007), pp. 29-43, ISSN 1774-0746.
- Khathuria N.1998. Rhizosphere microbiology of desert, M.Sc. Dissertation, Department Microbiology, Maharshi Dayanand Saraswati University, Ajmer (Rajasthan), P-52.
- Kucey R.M.N. *et al.* 1989. Microbial Mediated increases in plant available phosphatas. Advances in Agronomy vol.42, pp.199-288, ISSN 0065-2113.
- Lobova E.V. 1967. Pochvy pustynnolzony SSSR Akademiya Nauk Moscow (translation Lobova E.V. Soils of the desert zone of USSR. US Department of Commerce, Springfield, Virginia.

- Moghimi A. 1987. Charactersterization of rhizospheric products Espically 2-ketogluconic acid soil biology and biochemistry, vol.10, No.4 (April 1978), pp.283-287, ISSN0038-0717.
- Okoh L.A. *et al.* 1999. Studies on the bacteria, fungi, and springtails (Collembola) of an agro forestly arboretum in Nigeria. Pedobia. 43: 18-27.
- Parker L.W. *et al.* 1984. Effects of simulated rain fall and litter quantities on desert soil biota: soil respiration, microflora and protozoa, pedobiology 27, 185-195.
- Paul EA *et al.* 1996. Soil microbiology and biochemistry. 2nd Edition New York. Academic press.pp.225-229.
- Rao A.V. et al 1983. Microbial ecology of the soil of India's desert agric. *Ecosystem environment* 10, 361-365.
- Schreiner R.P. *et al.* 1997. Mycorrhizal fungi influence, plant and soil functions and interactions. 188: 199-209.
- Tarafdar J.C. *et al* 2003. Efficiency of some phosphatase producing soil fungi Indian journal of microbiology, Vol. 43, No.1,(March 2003), pp. 27-32 ISSN 0046-8991.

- Trappe J.M. 1981. Mycorrhizae and productivity of arid and semi arid range lands, In adrances in food production system for Arid and semi Arid lands (eds Manassah J.J. and Briskey E.J.), Academic press, New York, pp. 581-599.
- Venkateswarlu B. *et al.* 1984. Evaluation of phosphorous solubilization by microorganisms isolated from arid soils. *Journal of the Indian society of soil science*, Vol. 32, No. 3, (September 1984), pp.273-277, ISSN 0019-638X.
- Whitelow M.A. 2000. Growth promotion of plants inoculated with phosphate solubilizing fungi. Advance in Agronomy, Vol.69, (December 1999), pp. 99-151, ISSN 0065-2113.
- Yadav S. et al. 2009. Processes Biochemistry, Vol.44, No. 1. P-1.
- Zhou J. *et al.* 2003. Bacterial phylogenetic diversity and a novel candidate division of two humid regions. Sandy surface soil. *Soil Biology and Biochemistry*, 35, 915-924, doi: 10.1016/s0038-0717(03) 00124-X.
