



RESEARCH ARTICLE

LIPID BASED DRUG DELIVERY SYSTEMS: A STRATEGY FOR ENHANCING THE ORAL BIOAVAILABILITY OF POORLY WATER-SOLUBLE DRUGS

^{1,a,*}Irfan A. Mohammed and ²BhavinY. Gajera

¹Department of Pharmaceutical Sciences, University of the Sciences, Philadelphia, PA 19104, USA

²Impel NeuroPharma, Seattle, Washington, USA

^aNevakar, Inc. Bridgewater, New Jersey, USA

ARTICLE INFO

Article History:

Received 08th April, 2018
Received in revised form
27th May, 2018
Accepted 24th June, 2018
Published online 30th July, 2018

Key Words:

Oral bioavailability,
Liposomes, Niosomes,
Lipid Nanoparticles, Nanosuspension,
Poorly Water Soluble Drugs.

ABSTRACT

The poor oral bioavailability of many drugs is mainly due to the poor aqueous solubility, chemical instability and preabsorptive metabolism. Numerous approaches have been developed for enhancement of oral bioavailability and are currently in the clinical application. Even though, some drugs do not meet the required clinical application due to the patient compliance and ineffective therapeutic levels. Vesicular delivery systems are considered as alternative delivery for the enhancement of the bioavailability of this category of drugs. The enhanced bioavailability of the lipophilic drugs from the vesicular systems mainly due to the increased effective surface area of the drug in the presence of lipids, surfactants and co surfactants, enhanced lymphatic uptake, altered gastric motility and by virtue of their small particle size. Extensive literature is available for the properties, applications, and preparation and evaluation methods. This review mainly dealt with the reported drug loaded various vesicular systems such as liposomes, niosomes, lipid nanoparticles, self-emulsifying delivery system, nanosuspensions.

Copyright © 2018, Irfan A. Mohammed and BhavinY. Gajera. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Citation: Irfan A. Mohammed and BhavinY. Gajera, 2018. "Lipid based drug delivery systems: A strategy for enhancing the oral bioavailability of poorly water-soluble drugs", *International Journal of Current Research*, 10, (07), 71000-71006.

INTRODUCTION

Oral route of administration of drugs is the preferred choice of drug delivery, principally due to better patient compliance, ease of administration, and cheaper in terms of cost of production. The oral bioavailability and thus the efficacy, is an indication of the solubility of drug in the gastrointestinal fluids and its intestinal membrane permeability (Lipinski, 2002). Earlier, lower bioavailability of oral dosage forms was only considered to be the aspect of physicochemical properties of the drug. In later phase due to advancement in technologies of drug delivery systems, numerous biochemical, biological, and receptors level interactions came into light as causes for such experimental results (Palmer, 2003). The prevalent application of combinatorial chemistry and high-throughput screening in the process of drug discovery during the period of past two decades has made new molecular entities (NME) highly insoluble in aqueous media.

Orally administered drugs are obliged to dissolve in gastrointestinal (GI) fluids preceding their absorption into the body (Prajapati and Patel, 2007). In view of the fact that in many instances the drug dissolution step is proved to be the rate limiting step, suitable formulation design can be a functional approach to improve the dissolution and thus the oral bioavailability of such molecules. The drug dissolution in the gastrointestinal tract (GIT) will be influenced by the characteristics of the GI components like volume of GI fluids, pH, surfactant concentration, and also the physico-chemical properties like pKa and log P of the drug. Numerous factors like molecular size, lipophilicity of the drug, in addition to its affinity to influx or efflux transporter proteins. Modification physicochemical properties like particle size reduction, salt formation may always do not work due to the limitations. In case of salt forms problems like feasibility of salt formation of neutral compounds, form conversion from salt to original acid or base form may lead to aggregation in GIT. Particle size reduction is not advantageous in case of very fine powders with poor wetting properties. Various strategies like solid dispersions (Narendar *et al.*, 2016, Carmen *et al.*, 2015), cyclodextrins (Ettireddy *et al.*, 2017), buccal delivery (Palem *et al.*, 2011; Chinna *et al.*, 2016), gastro retentive delivery (Desai and Bolton, 1993; Reddy *et al.*, 2012), floating tablets

*Corresponding author: Irfan A. Mohammed,

¹Department of Pharmaceutical Sciences, University of the Sciences, Philadelphia, PA 19104, USA.

^aNevakar, Inc. Bridgewater, New Jersey, USA.

DOI: <https://doi.org/10.24941/ijcr.31364.07.2018>

(Dudipala *et al.*, 2011) and nanoparticulate delivery systems were employed to overcome these issues concerned to poor bioavailability (Pitta *et al.*, 2018). In this review, attempts were made to discuss about the strategies to enhance the oral bioavailability of drugs using various liposomal drug delivery systems such as liposomes, niosomes, lipid nanoparticles (solid lipid nanoparticles and nanostructures lipid carriers), and nanosuspensions and self-emulsifying drug delivery systems. Previously, some reports are published for the development of various nanodelivery systems for independent drugs such as Candesratancilexetil (Kishan, 2017), Zaleplon (Doodipala *et al.*, 2016)

Liposomes: Liposomes are spherical shape bilayers vesicles obtained from the combination of cholesterol and phospholipids (Allen, 1997). They are the most promising drug delivery systems owing to their physicochemical and biocompatibility properties. The physical and stability properties of liposomes differ considerably and depends on lipid composition, surface charge, size, and the method of preparation used for making (Abolfazl *et al.*, 2013). Furthermore, the rigidity and the charge of the bilayer membrane are mainly influenced by the composition of the bilayer (Karthik *et al.*, 2012). It has been also reported that, saturation and source of the phosphatidylcholine (PC) species could impact the vesicle rigidity. Unsaturated PC from natural source, like egg or soybean phosphatidylcholine, resulted in much more permeable and less stable bilayers, while the saturated phospholipids with long acyl chains (dipalmitoylphosphatidylcholine etc..) form a rigid and impermeable vesicular construct (Shehata *et al.*, 2008). Liposomes and proliposomes are used to improve the oral bioavailability by increasing the residence time in the gastro intestinal tract (GIT), surface modification promote uptake and avoiding first-pass metabolism. Various reported liposome and proliposome formulations are presented in Table 1.

Niosomes: Niosomes are multi-lamellar structures similar to liposomes and are composed of non-ionic surfactant (Cosco *et al.*, 2009; Rampal *et al.*, 2011). Niosomes are now widely studied as an alternative approach to liposome. There are different types of surfactants that have the ability to form vesicles, entrap and retain the hydrophilic and hydrophobic solute particles. They primarily consist of two types of components, i.e., nonionic surfactant and the additives. The additives used in the preparation of niosome are cholesterol and the charged molecules (Toshimitsu *et al.*, 1994). Cholesterol is a critical component of the cell membrane and its presence improves the rigidity of the bilayer. The presence of cholesterol in the membrane also plays an important role in imparting fluidity and permeability to the cell membrane. As a carrier system for both small and large molecule, niosomes protect the drug molecules from premature degradation and inactivation due to untoward immunological and pharmacological effects (Aranya *et al.*, 2003). Niosomes have also been used to overcome challenges with drug solubility, stability and degradation. Table 2 describes the various reported niosomal formulations.

Lipid nanoparticles: Solid lipid nanoparticles and nanostructured lipid carriers are the extensively used lipid based nanoparticles. Solid lipid nanoparticles (SLNs) are sub-micron colloidal carrier nanoparticles with particle size range from 50-1000 nm (Muller, 2000). SLNs are mainly composed of solid lipids, which are stable as solid at room temperature.

Stability and aggregation of particles are reduced by the incorporation of surfactant and co-surfactant, respectively. The commonly used solid lipids include triglycerides (Dynasan-112, Dynasan-114, Dynasan-116 and Dynasan-118) mixed glycerides (Compritrol ATO 888, glyceryl monobenhenate) and monoglycerides (stearic acid, glyceryl monostearate) (Mukherjee *et al.*, 2009). SLNs have the advantages of biocompatibility, reduced toxicity, used for incorporation of both hydrophilic and lipophilic drugs and enhanced oral bioavailability and also pharmacodynamic activity (Gorre *et al.*, 2017). The enhanced oral bioavailability of drugs might be due to enhanced surface area by the addition of surfactants, by virtue of small particle size, presence of lipids promote the gastric motility and also promote the lymphatic transport by reducing the first-pass metabolism and also tumor targeting (Narendar and Goverdhan, 2018). Drug loaded SLNs for enhancement of oral BA are showed in Table 3.

Nanostructured lipid carriers: Nanostructured lipid carriers (NLCs) are considered as modified form of solid lipid nanoparticles. The difference between the SLNs and NLCs is replace one part of the solid lipid with liquid lipid and observed for improved properties (Muller *et al.*, 2002). In general, the NLCs made of 1:3 to 1:4 ratio of liquid lipid to solid lipid and along with surfactants and cosurfactants. NLCs minimize drug expulsion, increase the drug encapsulation and stability of loaded drug compared with SLNs (Radtke *et al.*, 2005). Preparation of NLCs mainly involves in the selection of liquid lipid and solid lipid and their respective ratios, selection of surfactants and cosurfactants, in some instances miscellaneous agents such as viscosity modifiers, antioxidants and preservatives (Ana *et al.*, 2016). NLCs like SLNs also help in controlled and sustained drug delivery (Westesen *et al.*, 1997), increased gastric residence time (Garcia-Fuentes *et al.*, 2003). Various drugs loaded NLCs are depicted in Table 4.

Nanosuspension: Nanosuspensions are submicron colloidal dispersions of nano-sized drug particles stabilized by the surfactants (Muller *et al.*, 2000). Nanosuspensions consist of poorly water-soluble drug without any matrix material suspended in dispersion (Rabinow, 2004). Nanosuspensions are useful for molecules with poor solubility and/or poor permeability, which poses a significant challenge for formulators. The reduction in particle size gives nanosuspensions a possibility to administer drugs intravenously. These suspensions can also be lyophilized to obtain a solid matrix. It also has the advantages of liquid formulations over others (Liversidge and Cundy, 1995). They are suitable for hydrophilic drugs, higher drug loading is possible, the dose of the active can be reduced, aid in enhancing the physical and chemical stability of drugs (Karri *et al.*, 2015) and also provides passive drug targeting (Grau *et al.*, 2000; Keck and Müller, 2006). List of drugs developed as nanosuspensions are reported in Table 5.

Self-emulsifying drug delivery systems: Self-emulsifying drug delivery systems (SEDDS) are one of the approaches to improve the oral bioavailability of poorly soluble drugs by presenting the drug in the form of small droplets of oil and maintaining it in a dissolved state throughout its transit time in gastrointestinal tract (GIT) (Pouton, 1985). The composition of SEDDS consists of oil and surfactant. They can form oil-in-water emulsions upon the natural agitation provided within the gastrointestinal movements.

Table 1. List of reported liposomes for improved oral bioavailability

Drug	Type of Formulation	Components	Pre-Clinical Study/ Subject	Outcome	Reference
Vincristine	Liposome	Dihydrospingomyelin	In vivo/ Rats	Increase in half-life and residence time	Johnston <i>et al.</i> , 2007
Dehydrosilymarin	Proliposome	Soybean phospholipids, cholesterol	In vivo/ Rabbits	2.28-folds increase in bioavailability	Chang <i>et al.</i> , 2011
Zaleplone	Proliposome	Soyphosphatidylcholine (HSPC) and cholesterol	In vivo/ Rats	2 to 5 fold increase in bioavailability	Karthik <i>et al.</i> , 2012
RaloxifeneHCl	Proliposome	Soyphosphatidylcholine and cholesterol	In vivo/ Rats	3-fold increase in bioavailability	Ashok <i>et al.</i> , 2013
Heparin	Liposome	Soyphosphatidylcholine and cholesterol	In vivo/ Rats	Bioavailability increased by 3-times	Lavanya <i>et al.</i> , 2016
Valsartan	Proliposome	Dimyristoylphosphatidylglycerol sodium (DMPG) and cholesterol	In vivo/ Rats	2-fold increase in bioavailability	Nekkanti <i>et al.</i> , 2016

Table 2. List of reported niosomes for improved oral bioavailability

Drug	Components	Pre-Clinical Subjects	Study/ Outcome	Reference
Aciclovir	Cholesterol, span 60, Dicetylphosphare	In vivo/ Rabbits	2-fold increase in bioavailability	Ismail <i>et al.</i> , 2007
Griseofulvin	Span 20, 40, 60, cholesterol, DCP	In vivo/ Rats	Increase in MRT	Jadon <i>et al.</i> , 2009
Clarithromycin	Span 20, 40, 60, and 80 and cholesterol	In vivo/ Rats	1.5-fold increase in bioavailability	Gyati <i>et al.</i> , 2016
Diltiazem	Span 60 or Brij-52 wif cholesterol	In vivo/ Rats	Increased AUC	Ammar <i>et al.</i> , 2017
Gliclazide	Span 60 and cholesterol	In vivo/ Rats	Reduction in blood glucose level	Tamizharasiet <i>et al.</i> , 2009

Table 3. Various drug loaded solid lipid nanoparticles

Drug	Lipid	Outcome	Reference
Baicalin	Stearic acid	Enhanced bioavailability	Hao <i>et al.</i> , 2012
Carvedilol	Poloxamer	Improved BA	Vinay Kumar <i>et al.</i> , 2012
Raloxifene hydrochloride	Compritol 888 ATO	Bioavailability enhanced	Burra <i>et al.</i> , 2013
Nisoldipine	Tripalmitate	Improved BA	Narendar and Kishan, 2015
Rosuvastatin calcium	Dynasan 114, Dynasan116, Dynasan 118	Improved BA	Suvarna <i>et al.</i> , 2015
Felodipine	Dynasan 114, 116 and 118	Improved BA	Usha <i>et al.</i> , 2015
Lacidipine	Dynasan 114, 116 and 118	Improved oral BA	Sandeep <i>et al.</i> , 2016
Candesartan cilexetil	Dynasan 114, Dynasan116, Dynasan 118	Improved BA	Reddy and Veerabrahma, 2016
Rosuvastatin calcium	Dynasan 112	Improved BA	Dudhipala and Veerabrahma, 2017
Zaleplon	Dynasan 114	Improved BA	Reddy and Janga, 2017
Olmesartanmedoxomil	GMS and SA	Improved BA	Arun <i>et al.</i> , 2018

Table 4. Various drug loaded solid lipid nanoparticles

Drug	Lipid	Outcome	Ref
Chlorambucil	Stearic acid and oleic acid	Improved drug action	Sharma <i>et al.</i> , 2009
Curcumin	Soylecithin and Poloxamer 188	11.93-foldbioavailability enhancement	Min Fang <i>et al.</i> , 2012
Carvedilol	Stearic acid and Oleic acid	3.95-foldbioavailability enhancement	Mishra <i>et al.</i> , 2016
Raloxifene hydrochloride	Glyceryl monostearate and Capmul MCM C8	3.57-fold bioavailability enhancement	Shah <i>et al.</i> , 2016
Vincristine sulfate	Hyaluronic acid	1.8-foldbioavailability enhancement	Xuan <i>et al.</i> , 2017
Atorvastatin	Capryol, lecithin	3.6-foldbioavailability enhancement	Mohammed <i>et al.</i> , 2017
Nisoldipine	Dynasan 114	Improved drug action	Narendar <i>et al.</i> , 2018

Table 5. Nanosuspensions of drugs as oral delivery vehicle

Drug	Components	Pre-Clinical Study/ Subject	Outcome	Reference
Curcumin	SLS and PVP	In vivo/ Rats	6.8-foldincrease in bioavailability	Gao <i>et al.</i> , 2016
Efavirin	Sodium lauryl sulfate and PVP K30	In vivo/ Rats	2.19-fold increase in bioavailability	Patel <i>et al.</i> , 2014
Furosemide	PVP	In vivo/ Rats	1.38-fold increase in bioavailability	Bhanu and Malay, 2014
Felodipine	PVA and HPMC	In vivo/ Rats	Enhanced AUC	Sahu and Dasu, 2014
Cefdinir	SLS and PVP	In vivo/ Rats	1.75-fold increase in bioavailability	Thota <i>et al.</i> , 2014
Cefdinir	Zirconium oxide	In vivo/ Rats	3-foldincrease in bioavailability	Sawant <i>et al.</i> , 2016
Curcumin	SLS and PVP	In vivo/ Rats	4.2-foldincrease in bioavailability	Li <i>et al.</i> , 2016
Olmesartnmedoxo mil	SLS	In vivo/ Rats	2.45-foldincrease in bioavailability	Nagaraj <i>et al.</i> , 2017

Table 6. List of reported SEDDS formulations

Drug	Components	Size (nm)	Outcome	Reference
Cinnarizine SNEDDS	Sesame oil/ Cremophor RH 40 Oleic acid Brij 97 (Co-surfactant) Ethanol	28.1 ± 0.96	Approximately increased by 25% compared to conventional tablets	Larsen <i>et al.</i> , 2012
Silymarin SMEDDS	Ethyl linoleate/Tween 80/ethyl alcohol	10-20	48.82-fold compared to drug suspension	Iosioet <i>et al.</i> , 2011
Rosuvastatin calcium SNEDDS	Cinnamon oil /Iabrasol; CapmulMCMC8	120-170	2.45-fold compared to drug suspension	Balakumaret <i>et al.</i> , 2013
Curcumin SMEDDS	Ethyl oleate/ emulsifier OP + Cremophor EL (1:1), co-surfactant (PEG 400)	21.4 ± 1.5	3.86-fold compared to drug suspension	Jing <i>et al.</i> , 2009
Amiodarone and talinolol SNEDDS	Triglyceride (trilaurin for amiodarone and tricaprin (for talinolol) / polyoxyl 40-hydroxy castor oil, Tween 20, and Span 80 and lecithin	10±0.03 (for Amiodarone) 45±0.07 (for tricaprin)	2 and 3 fold increase for Amiodarone and Talinolol respectively	Anna <i>et al.</i> , 2013
Lercanidipine Zaleplon SNEDDS powder	Capmul and Tween 80 Neusilin US2	147 ± 3 138 ± 6	Enhanced drug release 3.5-fold	Kalakuntlaet <i>et al.</i> , 2012 Yadav <i>et al.</i> , 2013

In such a system, the lipophilic drug is incorporated in solution, in small droplets of oil in solution from. The large interfacial area generated by these smaller size of droplets, facilitates drug diffusion into intestinal fluids (O'Driscoll 2002). Additionally increased fraction of absorption by lymphatic transport, avoids hepatic first-pass metabolism of drugs which are prone to extensive metabolism (Cuiné *et al.*, 2007). The SEDDS are the delivery systems which were engineered through a specific combination of selected lipids and emulsifiers in a specific ratio.

In addition, for a specific drug a particular SEDDS should be developed using different excipients with different physicochemical properties to improve overall hydrophilicity of the drug (Kossena *et al.*, 2005). On digestion in GIT lipid excipients form different colloidal species (vesicles, micelles and liquid crystalline phases) in the intestinal lumen which further had an impact on dissolution and absorption of drug co-administered (Porter *et al.*, 2008). Many of the excipients were reported to aid in lymphatic bypass and also considerably reducing the access to pre-systemic transporter mediated drug efflux like P-glycoprotein (P-gp) (Zhang *et al.*, 2003). Table 6 presents the list of different drug loaded SEDDS formulations.

Conclusion

The enhancement of oral bioavailability of drugs mainly depends on aqueous solubility and permeability properties. Various oral drug delivery forms, such as, buccal delivery, floating delivery, etc. approaches have been developed for the enhancement of bioavailability. Some of the drug delivery technologies enhancing oral bioavailability have been successfully commercialized and many other promising technologies are currently under investigation. Nevertheless, in the coming years, the technologies to enhance oral drug bioavailability will see tremendous growth to help scientists develop new and improved drug products for better health outcomes. Furthermore, these technologies can also be used to improve the drug formulation and/or delivery of existing and approved active pharmaceutical ingredients for their respective indications or even a new indication which is certainly a possibility with advancement in drug delivery and development technologies.

REFERENCES

- Abolfazl Akbarzadeh,  Rogaie Rezaei-Sadabady,^{1,2} Soodabeh Davaran,¹ Sang Woo Joo,  Nosratollah Zarghami,  ¹Younes Hanifehpour,⁵ Mohammad Samiei,³ Mohammad Kouhi,⁴ and Kazem Nejati-Koshki. Liposome: classification, preparation, and applications. *Nanoscale Res Lett.* 2013; 8(1): 102.
- Allen TM. 1997. Liposomes. Opportunities in drug delivery. *Drugs.*, 54(4):8–14.
- Ammar, H. O., Haider, M., Ibrahim, M. and El HOFFY. N. M. 2017. *In vitro* and *in vivo* investigation for optimization of niosomal ability for sustainment and bioavailability enhancement of diltiazem after nasal administration. *Drug delivery*, 24(1):414-421.
- Ana Beloqui, María Ángeles Solinís, Alicia Rodríguez-Gascón, António J. Almeida, Véronique Préat. 2016. Nanostructured lipid carriers: Promising drug delivery systems for future clinics. *Nanomed, nanotech, bio and med*, 2(1):143-161.
- Anna Elgart, Irina Cherniakov, Yanir Aldouby, Abraham J. Domb, Amnon Hoffman. 2013. Improved Oral Bioavailability of BCS Class 2 Compounds by Self Nano-Emulsifying Drug Delivery Systems (SNEDDS): *The Underlying Mechanisms for Amiodarone and Talinolol.* *Pharm Res.*, 30:3029–3044.
- Aranya Manosroi, Paveena Wongtrakul, Jiradej Manosroi, Hideki Sakaie, 2003. Fumio Sugawara, Makoto Yuasa, Masahiko Abe. Characterization of vesicles prepared with various non-ionic surfactants mixed with cholesterol. *Colloids Surf., Biointerfaces*, 30, 129–138.
- Arun, B., Reddy ND, and Kishan, V. 2018. Development of olmesartan medoxomil lipid based nanoparticles and nanosuspension: preparation, characterization and comparative pharmacokinetic evaluation. *Artificial cells, nanomed and biotech*, 46(1), 126-137.
- Ashok Velpula, Raju Jukanti, Karthik Yadav Janga, 2013. Sharath Sunkavalli, Suresh Bandari, Prabhakar Kandadi. Pro-liposome powders for enhanced intestinal absorption and bioavailability of raloxifene hydrochloride: effect of surface charge. *J microenc.*, 39(12):1895-1906.
- Balakumar K, Raghavan CV, Selvan NT, Prasad RH, Abdu S. 2013. Self nanoemulsifying drug delivery system (SNEDDS) of rosuvastatin calcium: design, formulation,

- bioavailability and pharmacokinetic evaluation. *Colloids Surf B Biointerfaces*. 112:337-43.
- Bhanu P. Sahu, Malay K. 2014. Das Formulation, optimization, and in vitro/in vivo evaluation of furosemide nanosuspension for enhancement of its oral bioavailability. *J Nanopart Res.*, 16:2360.
- Burra M, RajuJukanti, Karthik Yadav Janga, Sharath Sunkavalli, Ashok Velpula, SrinivasAmpati, K.N. Jayaveera. 2013. Enhanced intestinal absorption and bioavailability of raloxifene hydrochloride via lyophilized solid lipid nanoparticles. *Advanced Powder tech*, 24(1):393-402.
- Carmen Popescu, Prashanth Manda, Abhishek Juluri, Karthik Yadav Janga, ManasaCidda, SN Murthy. 2015. Enhanced dissolution efficiency of zaleplon solid dispersions via modified β -cyclodextrin molecular inclusion complexes. *J. Pharm Pharm Sci.*, 1(1), 12-21.
- Chang Chu, Shan-shan Tong, Ying Xu, Li Wang, Min Fu, Yan-ru Ge, Jiang-nan Yu and Xi-ming Xu. 2011. Proliposomes for oral delivery of dehydrosilymarin: preparation and evaluation *in vitro* and *in vivo*. *Acta Pharmacologica Sinica* volume32, pages973–980.
- Chinna PR, ReddyND., Sunil, B., Repka, M.A., andYamsani MR. 2016. Development, Optimization and in vivo Characterization of Domperidone Controlled Release Hot Melt Extruded Films for Buccal Delivery. *Drug Dev Ind Pharm*, 42(3), 473-484.
- Cosco D¹, Paolino D, Muzzalupo R, Celia C, Citraro R, Caponio D, Picci N, Fresta, M. 2009. Novel PEG-coated niosomes based on bola-surfactant as drug carriers for 5-fluorouracil. *Biomed Microdevices.*, Oct;11(5):1115-25.
- Cuiné, J.F., Charman,W.N., Pouton, C.W., Edwards, G.A., Porter, C.J.H., 2007. Increasing the proportional content of surfactant (Cremophor EL) relative to lipid in self-emulsifying lipid-based formulations of danazol reduces oral bioavailability in beagle dogs. *Pharm. Res.* 24, 748–757.
- Desai, S. and Bolton, S. 1993. A Floating Controlled-Release Drug Delivery System: *In Vitro-in Vivo Evaluation*. *Pharm Res.*, 10(9):1321-1325.
- Doodipala R. 2016. A review of novel formulation strategies to enhance oral delivery of zaleplon. *J Bioequi avail.* 8(5): 211-213.
- Driscoll, D.F., Nehne, J., Peterss, H., Franke, R., Bistran, B.R., Niemann, W., 2002. The influence of medium-chain triglycerides on the stability of all-in-one formulations. *Int. J. Pharm.* 240, 1–10.
- Dudhipala N, and Veerabrahma K. 2017. Improved anti-hyperlipidemic activity of Rosuvastatin Calcium via lipid nanoparticles: pharmacokinetic and pharmacodynamic evaluation. *Euro J Pharm Biopharm.* 110 (1), 47-57.
- Dudipala R, Palem, C.R., Reddy, S., and Rao, Y.M. 2011. Pharmaceutical development and clinical pharmacokinetic evaluation of gastroretentive floating matrix tablets of levofloxacin. *Int J Pharma Sci and Nanotech*,4(3), 1461-1467.
- Ettireddy S, and Reddy ND. 2017. Influence of β -cyclodextrin and hydroxypropyl- β -cyclodextrin on enhancement of solubility and dissolution of isradipine. *Int J Pharma Sci and Nanotech*, 10(3), 3752-3757.
- Gao, Yan and Wang, Chao and Sun, Min and Wang, Xin and Yu, Aihuaand Li, AiguoandZhai, Guangxi. 2012. In Vivo Evaluation of Curcumin Loaded Nanosuspensions by Oral Administration. *Journal of biomedical nanotechnology.* 8. 659-68.
- Garcia-Fuentes M, Torres D, Alonso MJ. 2003. Design of lipid nanoparticles for the oral delivery of hydrophilic macromolecules. *Colloids Surf B*27:159–68.
- Gorre T, Swetha E and Reddy D. 2017. Role of isradipine loaded solid lipid nanoparticles in the pharmacodynamic effect of isradipine in rats. *Drug res*, 67(03): 163-169.
- Grau MJ, Kayser O, Müller RH. 2000. Nanosuspensions of poorly soluble drugs--reproducibility of small scale production. *Int J Pharm. Mar 10; 196(2):155-9.*
- Gyati ShilakariAs TEMPthana, * Parveen Kumar Sharma, and AbhayAs TEM Pthana. *In Vitro and In Vivo Evaluation of Niosomal Formulation for Controlled Delivery of Clarithromycin*. Scientifica (Cairo). 2016; 2016: 6492953.
- Hao J¹, Wang F, Wang X, Zhang D, Bi Y, Gao Y, Zhao X, Zhang Q. 2012. Development and optimization of baicalin-loaded solid lipid nanoparticles prepared by coacervation method using central composite design. *Eur J Pharm Sci.*, Sep 29;47(2):497-505.
- Iosio, T., Voinovich, D., Perissutti, B., Serdoz, F., Hasa, D., Grabnar, I., Dall' Acquac, S., Zarad, G.P., Muntonid, E., Pintoe. J.F. 2011. Oral bioavailability of silymarinphytocomplex formulated as self-emulsifying pellets. *Phytomedicine* 18 505–512.
- Ismail A. Attia, Sanaa A. El-Gizawy, Medhat A. Fouda, and Ahmed M. Donia. 2007. Influence of a niosomal formulation on the oral bioavailability of acyclovir in rabbits. *AAPS Pharm Sci Tech.*, Oct; 8(4): 206–212.
- Jadon P.S., Gajbhiye V., Rajesh S.J., Kavita R., Narayanan G. 2009. A Controlled and Novel Drug Delivery System. *AAPS Pharm.Sci.Tech.* 10: 1187-1192.
- Jing Cuia,b, Bo Yuc,d, Yu Zhaoe, WeiweiZhua, Houli Li a, Hongxiang Louf, Guangxi Zhaia. 2009. Enhancement of oral absorption of curcumin by self-microemulsifying drug delivery systems. *International Journal of Pharmaceutics*, 371 148–155.
- Johnston MJ, Semple SC, Klimuk SK, Ansell S, Maurer N, Cullis PR. 2007. Characterization of the drug retention and pharmacokinetic properties of liposomal nanoparticles containing dihydrosphingomyelin. *Biochim Biophys Acta.*, 1768:1121–1127.
- Kallakunta VR, Bandari S, Jukanti R, Veerareddy PR. 2012. Oral self emulsifying powder of lercanidipine hydrochloride: Formulation and evaluation. *Adv Powder Tech*, 221, 3750382.
- Karthik JY, Jukanti, R., Velpula, A., Sunkavalli, S., Bandari, S., and Kandadi, P. 2012. Bioavailability enhancement of zaleplon via proliposomes: Role of surface charge. *Eur J Pharma and Biopharma*, 80 (2), 347-357.
- Karri V, Butreddy A, Narender R. 2015. Fabrication of Efavirenz Freeze Dried Nanocrystals: Formulation, Physicochemical Characterization, In Vitro and Ex Vivo Evaluation. *Advanced Science, Engineering and Medicine.* 7(5): 385-392.
- Keck CM, Müller RH. Drug nanocrystals of poorly soluble drugs produced by high pressure homogenisation. *Eur J Pharm Biopharm.* 2006 Jan; 62(1):3-16.
- Kossena, G.A., Charman, W.N., Boyd, B.J. Porter, C.J.H.2005. Influence of the intermediate digestion phases of common formulation lipids on the absorption of a poorly water-soluble drug. *J. Pharm. Sci.* 94 481–492.
- Larsen, A.T., Ogbonna, A., Abu-Rmaileh, R., Abrahamsson, B., Østergaard, J., Müllertz, A., 2012. SNEDDS containing poorly water soluble cinnarizine; development and In vitro characterization of dispersion, digestion and solubilization. *Pharmaceutics* 4, 641–665.

- Li X¹, Yua H¹, Zhang C¹, Chen W¹, Cheng W¹, Chen X¹, Ye X¹. Preparation and in-vitro/in-vivo evaluation of curcumin nanosuspension wif solubility enhancement. *J Pharm Pharmacol*. 2016 Aug;68(8):980-8.
- Lipinski. C. 2002. Poor aqueous solubility—an industry wide problem in drug discovery. *American Pharmaceutical Review*, 5, pp. 82-85.
- Liversidge GG, Cundy KC. 1995. Particle size reduction for improvement of oral bioavailability of hydrophobic drugs: Absolute oral bioavailability of nanocrystalline danazol in beagle dogs. *Int J Pharm*. 125:91-7.
- Min Fang, Yilin Jin, Wei Bao, Hui Gao, Mengjin Xu, Di Wang, Xia Wang, Ping Yao, and Liegang Liu. 2012. In vitro characterization and in vivo evaluation of nanostructured lipid curcumin carriers for intragastric administration. *Int J Nanomedicine*. 7: 5395-5404.
- Mishra A¹, Imam SS², Aqil M¹, Ahad A³, Sultana Y¹, Ameenuzzafar¹, Ali A. 2016. Carvedilol nano lipid carriers: formulation, characterization and in-vivo evaluation. *Drug Deliv*. May;23(4):1486-94.
- Mohammed Elmowafy, Hany M. Ibrahim, Mohammed A. Ahmed, Khaled Shalaby, Ayman Salama and Hossam Hefesha. 2017. Atorvastatin-loaded nanostructured lipid carriers (NLCs): strategy to overcome oral delivery drawbacks. *Drug deli*, 24(1):932-941.
- Mukherjee, S., Ray, S. and Thakur. R. S. Solid Lipid Nanoparticles: A Modern Formulation Approach in Drug
- Muller RH, Gohla S, Dinger A, Schneppe T. Wise D. 2000. Handbook of pharmaceutical controlled release technology. New York: Marcel Dekker; Large-scale production of solid-lipid nanoparticles (SLN) and nanosuspension (Dissocubes) pp. 359-375.
- Muller RH, Radtke M, Wissing SA. 2002. Nanostructured lipid matrices for improved microencapsulation of drugs. *Int J Pharm*., 242(1-2):121-8.
- Müller, R.H., Mäder, K., and Gohla, S. 2000. Solid lipid nanoparticles (SLN) for controlled drug delivery – a review of the state of the art. *Eur J Pharm Biopharm*, 50(1), 161-177.
- Nagaraj, K., Narendar D. and Kishan. V. 2017. Development of olmesartan medoxomil optimized nanosuspension using the Box-Behnken design to improve oral bioavailability. *Drug Dev Ind Pharm*., 43(7):1186-1196.
- Nallaguntla Lavanya,¹ Yallamalli Indira Muzib, JiTEMPthan Aukunuru,² and Umamahesh Balekari. 2016. Preparation and evaluation of a novel oral delivery system for low molecular weight heparin. *Int J Pharm Investig*. ul-Sep; 6(3): 148-157.
- Narendar D, Arjun, N., Dinesh, S., and Karthik, J. 2016. Biopharmaceutical and Preclinical Studies of Efficient Oral Delivery of Zaleplon as Semisolid Dispersions with Self-emulsifying Lipid Surfactants. *Int J Pharma Sci and Nanotech*, 9(1),1-8.
- Narendar D, Govardhan K. 2018. Capecitabine lipid nanoparticles for anti-colon cancer activity in 1, 2-dimethylhydrazine induced colon cancer: Preparation, cytotoxic, pharmacokinetic and pathological evaluation. *Drug dev Ind pharm*, Early online, March. doi: 10.1080/03639045.2018.1445264.
- Narendar, D., and Kishan, V. 2015. Pharmacokinetic and pharmacodynamic studies of nisoldipine-loaded solid lipid nanoparticles developed by central composite design. *Drug Dev Ind Pharm*, 41(12), 1968-77.
- Narendar, D., Karthik, Yadav, J., and Thirupathi, G. 2018. Comparative study of nisoldipine-loaded nanostructured lipid carriers and solid lipid nanoparticles for oral delivery: preparation, characterization, permeation and pharmacokinetic evaluation. *Artificial cells, nanomed and biotech*, Early online 11 April, doi.org/ 10.1080/21691401.2018.1465068.
- Nirmal V. Shah *, Avinash K. Seth, R. Balaraman, Chintan J. Aundhia, Rajesh A. Maheshwari, Ghanshyam R. Parmar. 2016. Nanostructured lipid carriers for oral bioavailability enhancement of raloxifene: Design and in vivo study. *Journal of Advanced Research*., 7, 423-434.
- Palem CR, Ramesh, G., Doodipala N, Vamshi, V, Y., and Madhusudan R, Y. 2011. Transmucosal delivery of domperidone from bilayered buccal patches: in vitro, ex vivo and in vivo characterization. *Arch Pharm Res*, 34(10), 1701-1710.
- Palmer. A.M. 2003. New horizons in drug metabolism, pharmacokinetics and drug discovery. *Drug News and Perspectives*, 16, pp. 57-62.
- Patel GV¹, Patel VB, Pathak A, Rajput SJ. 2014. Nanosuspension of efavirenz for improved oral bioavailability: formulation optimization, in vitro, in situ and in vivo evaluation. *Drug Dev Ind Pharm*. Jan;40(1):80-91.
- Pitta S, Dudhipala N, Narala A and Veerabrahma K. 2017. Development and evaluation of zolmitriptan transfersomes by Box-Behnken design for improved bioavailability by nasal delivery. *Drug Dev Ind Pharm*, 2018, 44(3):484-492..
- Porter, C.J.H., Pouton, C.W., Cuine, J.F., Charman, W.N., 2008. Enhancing intestinal drug solubilisation using lipid-based delivery systems. *Adv. Drug Deliv. Rev.* 60, 673-691.
- Pouton, C.W. 1985. Self-emulsifying drug delivery systems: assessment of the efficiency of emulsification, *Int. J. Pharm.* 27 (2-3) 335-348.
- Prajapati, B.G., Patel. M. 2007. Conventional and alternative methods to improve oral bioavailability of lipophilic drugs *Asian Journal of Pharmaceutical*, 1, pp. 1-8.
- RabinowBE .Nanosuspensions in drug delivery. *Nat Rev Drug Discov*. 2004 Sep; 3(9):785-96.
- Radtke M, Souto EB, Muller RH. 2005. Nanostructured lipid carriers: a novel generation of solid lipid drug carriers. *Pharm Technol Eur*.,17(4):45-50.
- Rampal RAJERA, Kalpana NAGPAL, Shailendra Kumar SINGH,* and Dina Nath MISHRA. 2011. Niosomes: A Controlled and Novel Drug Delivery System. *Biol. Pharm. Bull.* 34(7) 945-953.
- Reddy N and Janga KY. 2017. Lipid nanoparticles of zaleplon for improved oral delivery by Box-Behnken design: Optimization, in vitro and in vivo evaluation. *Drug Dev Ind Pharm*, 43(7):1205-1214.
- Reddy ND and Kishan V. Candesartan cilexetil nanoparticles for improved oral bioavailability. *Ther deli*, 8(2):79-88.
- Reddy, N.D., Chinna R. P., Sunil, R., Madhusudan, R. Y. 2012. Development of floating matrix tablets of Ofloxacin and Ornidazole in combined dosage form: in vitro and in vivo evaluation in healthy human volunteers. *Int J Drug Deli*, 4, 462-469.
- Sahu BP¹, Das MK. 2014. Preparation and in vitro/in vivo evaluation of felodipine nanosuspension. *Eur J Drug Metab Pharmacokin*. Sep;39(3):183-93. doi: 10.1007/s13318-013-0158-5. Epub 2013 Nov 7.
- Sandeep, V., Reddy ND, Arjun, N., and Kishan, V. 2016. Lacidipine loaded solid lipid nanoparticles for oral delivery: Preparation, characterization and In vivo evaluation. *Int J Pharma Sci Nanotech*, 9(6), 3524-30.

- Sawant KK¹, Patel MH¹, Patel K¹. 2016. Cefdinirnanosuspension for improved oral bioavailability by media milling technique: formulation, characterization and in vitro-in vivo evaluations. *Drug Dev Ind Pharm.*, 42(5):758-68.
- Sharma P, Ganta S, Denny AW, Garg S. 2009. Formulation and pharmacokinetics of lipid nanoparticles of a chemically sensitive nitrogen mustard derivative. *Chlorambucil. Int J Pharm.*, 67:187-94.
- Shehata T, Ogawara K, Higaki K, Kimura T. 2008. Prolongation of residence time of liposome by surface-modification with mixture of hydrophilic polymers. *Int J Pharm.*, 359:272-279.
- Suvarna, G., Reddy D., and Kishan, V. 2015. Preparation, characterization and in vivo evaluation of rosuvastatin calcium loaded solid lipid nanoparticles. *Int J Pharma Sci and Nanotech*, 8(1), 2779-2785.
- Tamizharasi S, Dubey A, Rathi V1, Rathi JC. 2009. Development and Characterization of Niosomal Drug Delivery of Gliclazide. *J Young Pharm.*, 1(3):205-209.
- Thota S, Afzal MS, Bomma R and Veerabrahma K. 2014. Development and in vivo evaluation of cefdinirnanosuspensions. *Int J Pharm Sci nanotech*, 7(3):2553-2560.
- Toshimitsu Yoshioka, Brigitte Sternberg, Alexander T. Florence. 1994. Preparation and properties of vesicles (niosomes) of sorbitan monoesters (Span 20, 40, 60 and 80) and a sorbitantriester (Span 85). *International Journal of Pharmaceutics*. 105 (1). Pages 1-6.
- Usha, G.,Dudhipala N., andVeerabrahma K. 2015. Preparation, characterization and in vivo evaluation of felodipine solid lipid nanoparticles to improve the oral bioavailability. *Int J Pharma Sci Nanotech*. 8 (4), 2995-3002.
- Venishetty VK¹, Chede R, Komuravelli R, Adepu L, Sistla R, Diwan PV. 2012. Design and evaluation of polymer coated carvedilol loaded solid lipid nanoparticles to improve the oral bioavailability: a novel strategy to avoid intraduodenal administration. *Colloids Surf B Biointerfaces*. 2012 Jun 15;95:1-9.
- Vijaykumar Nekkanti, Z Wang, Guru V. Betagiri. 2016. Pharmacokinetic Evaluation of Improved Oral Bioavailability of Valsartan: Proliposomes Versus Self-Nanoemulsifying Drug Delivery System. *AAPS Pharm Sci Tech.*, 17(4); 851-862.
- Westesen K, Bunjes H, Koch MHJ. 1997. Physicochemical characterization of lipid nanoparticles and evaluation of their drug loading capacity and sustained release potential. *J Controlled Release.*, 48(2-3):223-36.
- Xuan Gao, Jun Zhang, Qiang Xu, Zun Huang, Yiyue Wang and Qi Shen. 2017. Hyaluronic acid-coated cationic nanostructured lipid carriers for oral vincristine sulfate delivery. *DDIP*, 47(3):661-667.
- Yadav KJ,Raju Jukanti, Sharath Sunkavalli, Ashok Velpula, Suresh Bandari, Prabhakar Kandadi and Prabhakar Reddy Veerareddy. 2013. *In situ* absorption and relative bioavailability studies of zaleplon loaded self-nanoemulsifying powders. *J microencapsu*, 30(2); 161-172.
- Zhang H, Yao M, Morrison RA, Chong S. 2003. Commonly used surfactant, Tween 80, improves absorption of P-glycoprotein substrate, digoxin, in rats. *Arch Pharm Res*. 26(9):768-72.
