



ISSN: 0975-833X

Available online at <http://www.journalcra.com>

INTERNATIONAL JOURNAL  
OF CURRENT RESEARCH

International Journal of Current Research  
Vol. 10, Issue, 11, pp.75403-75408, November, 2018  
DOI: <https://doi.org/10.24941/ijcr.33459.11.2018>

## RESEARCH ARTICLE

### FASN EXPRESSION CORRELATION ANALYSIS TO ANTI-CANCER DRUGS

<sup>1,\*</sup>Asima Tayyeb, <sup>1</sup>Uzma Qaisar, <sup>1</sup>Muhammad Shahjahan, <sup>1</sup>Hanfa Ashfaq and <sup>2</sup>Zafar Abbas Shah

<sup>1</sup>School of Biological Sciences, University of the Punjab, Lahore, Pakistan

<sup>2</sup>Department of Bioinformatics, Hazara University Mansehra, Pakistan

#### ARTICLE INFO

##### Article History:

Received 09<sup>th</sup> August, 2018  
Received in revised form  
20<sup>th</sup> September, 2018  
Accepted 19<sup>th</sup> October, 2018  
Published online 30<sup>th</sup> November, 2018

##### Key Words:

Photography camera, Books, Images etc.

#### ABSTRACT

The FASN is essential enzyme in de novo fatty acid synthesis that converted into phospholipids which provides resistance to drug uptake in malignancies. The FASN over expression and intrinsic/acquired both types of drug resistance reported in carcinomas. To assess the FASN up-regulation overall therapeutic response in drug screened cancer cell line data from Cancer Cell Line Encyclopedia (CCLE) by a rational CCLE GDSC gene expression - drug sensitivity correlations tool. We identify differential response of FASN in different drug treated tissue in which few cancer studies the drug performance increases in presence of FASN over expression but several cancer studies showed drug resistance by FASN over expression. The FASN increased expression drug resistance mainly linked with MAPK, EGFR, AKT, BCR/ABL, MDM2, HDAC and IGFR pathways that are responsible for angiogenesis, growth, survival, migration, differentiation and proliferation. Our study signifies the FASN elevated expression resistance to anti-proliferatory drugs for multiple oncogenes which indicate the FASN inter/intra-pathway interactions with oncogenes for their effective survival. We diagnose the FASN over expression as predictive marker in drug resistance genomes to design molecular medicines that consider it secondary target in generally accepted therapy.

Copyright © 2018, Asima Tayyeb et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Citation: Asima Tayyeb, Muhammad Shahjahan, Hanfa Ashfaq and Zafar Abbas Shah. 2018. "Fasn expression correlation analysis to anti-cancer drugs", International Journal of Current Research, 10, (11), 75403-75408.

## INTRODUCTION

The well-known homodimeric protein Fatty acid synthase (FASN) consist on six enzymatic domains that transform the acetyl-CoA and melonyl-CoA into palmitate fatty acid (Jensen-Urstad *et al.*, 1859). The initial studies reported their role in embryonic development by triggering the proliferation of several tissues (Chirala *et al.*, 2003). FASN perform significant contribution in molecular activity under aberrant metabolic states, inter-cellular integrity, cardiac stress related homeostasis response and regeneration of neural stem cells (Knobloch *et al.*, 2012). The FASN differential expression monitored in normal and tumor tissues including colorectal, pancreatic and breast cancer has overexpression of FASN (Cai *et al.*, 2014; Bhatt *et al.*, 2012). The FASN elevated expression regulated through various factors such as SREBP-1c, ChREBP, mTOR, AMPK, LXR $\alpha$ , NAC1, miRNAs and acetyltransferase p300 in malignancies (Ishii *et al.*, 2004; Hansmann *et al.*, 2006; Wang *et al.*, 2016). The USP2a expression stimulates the FASN up-regulation in autocrine manner in prostate cancer (Graner *et al.*, 2004).

The population genetic studies explore the FASN increased level with poor prognosis in 424 obese men in prostate carcinoma (Nguyen *et al.*, 2010). The FASN overexpression develops tumorigenesis by increasing the lipogenesis, Her2 activity, growth and proliferation of breast cells that induce factors-independent growth in culture (Vazquez-Martin *et al.*, 2008). The breast malignancies transforms into non-malignant states by FASN expression inhibition (Gonzalez-Guerrico *et al.*, 2016). The prostate cells gain 90% risk to develop invasive tumors with FASN overexpression and 30% risk with FASN down regulation (Fiorentino *et al.*, 2008). The FASN overexpression leads to excessive palmitoylation of Wnt-1 and AR receptors which are responsible for adenocarcinomas establishment that induce the regulation of FASN transcription factor SREBP (Migita *et al.*, 2009). The tumor cells utilize the palmitate for the synthesis of phospholipids to develop cell membrane inducing proliferation and 16-18C fatty acids makes alterations in plasma membrane fluidity (Rysman *et al.*, 2010; Li, 2014). Various proliferating agents like KRAS-A, NRAS, HRAS, tubulin and Wnt factors need palmitoylation for functional heterogeneity (Heuer, 2016). The Wnt/ $\beta$ -catenin, mTOR and PI3K/AkT signaling pathways showed loss of functionality by the inhibition of FASN expression (Röhrig, 2016). The decline of FASN expression leads to decrease the quantity of diacyl glycerols that induce cell death by reduction of Kinase C signaling pathway (Benjamin *et al.*, 2015). The

\*Corresponding author: Asima Tayyeb,

School of Biological Sciences, University of the Punjab, Lahore, Pakistan.

FASN over expression provides the nourishment to tumor microenvironment through vascularization, inflammatory response and lipogenesis in colorectal cancer (Zaytseva *et al.*, 2014). The FASN mediated drug resistance reported in several studies that produce high amount of fatty acids which induced cell membrane reduced permeability of chemotherapeutic drugs such as doxorubicin (Rysman *et al.*, 2010). There is urgent need to evaluate the multi-dimensional FASN differential expression behavior towards group of anti-cancer drugs to identify it as a secondary drug target and primary cause of drug resistance. We use a novel CCLE GDSC gene expression - drug sensitivity correlations tool ([www.public.tableau.com](http://www.public.tableau.com)) that operate on drug treated cell lines datasets to determine the effects of FASN expression on anti-cancer drug activity and efficiency.

## RESULTS

### FASN expression correlation towards anti-cancer drugs:

We obtain the FASN expression correlation in 24 types of cancers with 21 anti-cancer drugs. The Crizotinib or PF-2341066 is anti-cancer drug targeting ALK, ROS1, c-Met/HGFR which are significant promoter of oncogenesis in various malignancies. The crizotinib competitively interact with ATP-binding pocket of above kinases to prevent the risk of aberrant cell proliferation (Kwak *et al.*, 2010; Cui *et al.*, 2011; Wong *et al.*, 2009; Rikova *et al.*, 2007). Here FASN showed negative correlation with PF-2341066 in lung, ovary, soft tissue and urinary tract carcinomas that indicates the FASN supportive role in drug targeting activity and efficiency to inhibit tumorigenesis. The PH-665752 or small molecule c-MET inhibitor reported in prevention of cell migration, motility and proliferation (Christensen *et al.*, 2003). Here FASN showed negative correlation with PH-665752 in multiple myeloma to enhance drug activity. Erlotinib is EGFR inhibitor by interacting to its ATP-binding site and prevent the development of homodimer that initiate signaling cascade of cellular proliferation (Herbst, 2005b).

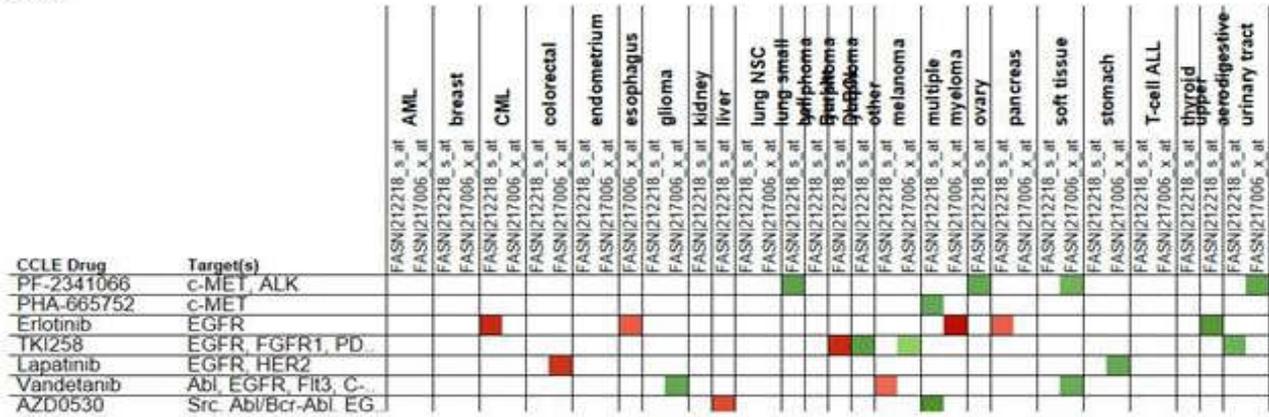
FASN showed positive correlation with Erlotinib in CML, esophagus, multiple myeloma and pancreatic cancers that is the sign of drug resistance which reduce the efficiency of drug. It also showed negative correlation in upper aerodigestive malignancies that invites to determine their supporting role in Erlotinib activity. Dovitinib or TK1258 inhibits VEGFR, PDGFR, FGFR1/3, FMS-3, c-KIT and colony stimulating factor receptor 1 that promote cancer cell proliferation, survival, angiogenesis and differentiation (Engebraaten *et al.*, 1993). FASN showed positive correlation with Dovitinib in lymphoma to resist the effects of drug on cell functions. In urinary tract, melanoma and few lymphomas FASN showed negative correlation to TK1258 for enhancement of its effects. Lapatinib inhibits EGFR and HER2 kinases in breast cancer via interacting to ATP-binding pocket to block self-phosphorylation and signal activation for cell growth (Moy, 2007). FASN has positive correlation in colorectal and negative correlation in stomach cancer that showed both resistance and supportive behavior towards Lapatinib. Vandetanib inhibits VEGFR2, EGFR and RET kinases in various forms of thyroid gland tumors (Fallahi *et al.*, 2015; Chougnat *et al.*, 2015; Ammer *et al.*, 2009). FASN has positive correlation in melanoma to reduce the activity of drug and negative correlation in soft tissue and glioma to enhance the activity of drug. Saracatinib or AZD0530 inhibits Src family of kinases, Abl-kinases and LCK kinases in treatments

of Alzheimer, schizophrenia and T-cell leukemia (Ammer *et al.*, 2009; Dong, 2010). FASN has positive correlation in liver cancer and negative correlation in myeloma which has tissue specific resistance. PLX4720 inhibit B-RAF pathway to prevent the cellular proliferation in melanomas (Li *et al.*, 2006). FASN has negative correlation in soft tissue cancer that indicates drug better activity in FASN expression. RAF265 inhibit RAF kinases and VEGFR2 to limit the risk of tumor cell proliferation and angiogenesis (Kumar *et al.*, 2009). FASN has positive correlation in breast and soft tissue cancer to decrease the effects of RAF265 and negatively correlation in CML, colorectal and breast cancer to provide support in drug efficiency. Selumetinib or AZD6244 is antineoplastic drug inhibit MEK1/2 which is the major driver of proliferative cellular pathways (Yeh *et al.*, 2007). FASN showed strong positive correlation in CML and lymphoma that uncover its resistance to AZD6244. It has negative correlation in upper aerodigestive and stomach cancers to support the drug performance. Topotecin inhibit topoisomerase I to induce cell death by preventing DNA replication. It is specific S-Phase anti-cancer drug (Léger *et al.*, 2004). FASN has healthy positive correlation in lymphoma and AML that indicates the resistance and gains intentions to determine their interaction as a secondary target. FASN has negative correlation in kidney cancer as a tissue specific scenario. Tanespimycin or 17AAG is inhibitor of HSP90 which is the controller of protein confirmations, growth and survival signaling pathways (Dimopoulos *et al.*, 2011).

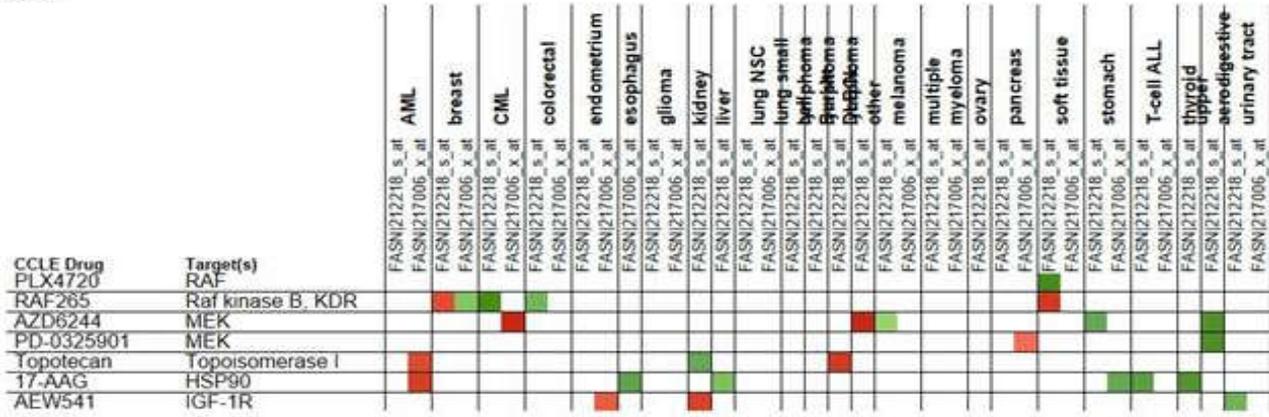
FASN showed negative correlation esophagus, liver, thyroid, stomach and T-cell ALL which indicates drug ideal activity in FASN expression. It has positive correlation in AML which showed its drug resistance property. AEW541 inhibit IGF-1R receptor kinase which is responsible for angiogenesis, transformations, survival, proliferation and metastasis. The drug inhibits the autophosphorylation of IGF-1R (García-Echeverría *et al.*, 2004). FASN has strong positive correlation in endometrium and kidney cancers specify its resistance to drug effects. L-685458 is  $\gamma$ -secretases inhibitor which is transmembrane protein involved in Alzheimer disease (Shearman *et al.*, 2000). FASN has strong positive correlation in T-cell ALL and negative correlation in endometrium that showed its differential behavior towards L-685458. Nilotinib is inhibitor of Bcr-Abl mediated proliferation, PDGFR, c-KIT and GISTs in CML therapies (Manley *et al.*, 2010).

FASN has strong positive correlation in breast and glioma which showed drug resistance. Nutlin-3 inhibits MDM2 and activates TP53 apoptotic inducer to prevent the process of tumorigenesis. FASN has positive correlation in melanomas to decrease the effects of drug of apoptotic induction. Paclitaxel is anti-tumor drug that promote tubulin polymerization leading cell cycle arrest to cell death (Ganguly *et al.*, 2010). FASN has negative correlation in liver and soft tissue cancer specify its progressive role in drug efficiency. Panobinostat is the inhibitor of Histone deacetylase enzymes to induce apoptosis (Gaur *et al.*, 2015). FASN has positive correlation in AML and multiple myeloma which gives resistance to drug competence. It also has negative correlation in liver and lymphomas with drug effects. Palbociclib or PD-0332991 is the inhibitor of cyclin-dependent kinases in ER-positive and HER2-positive breast malignancies (Finn *et al.*, 2009). FASN has negative correlations in colorectal, lung and ovary cancer that promotes the effects of drug (Fig 1).

CCLE



CCLE



CCLE

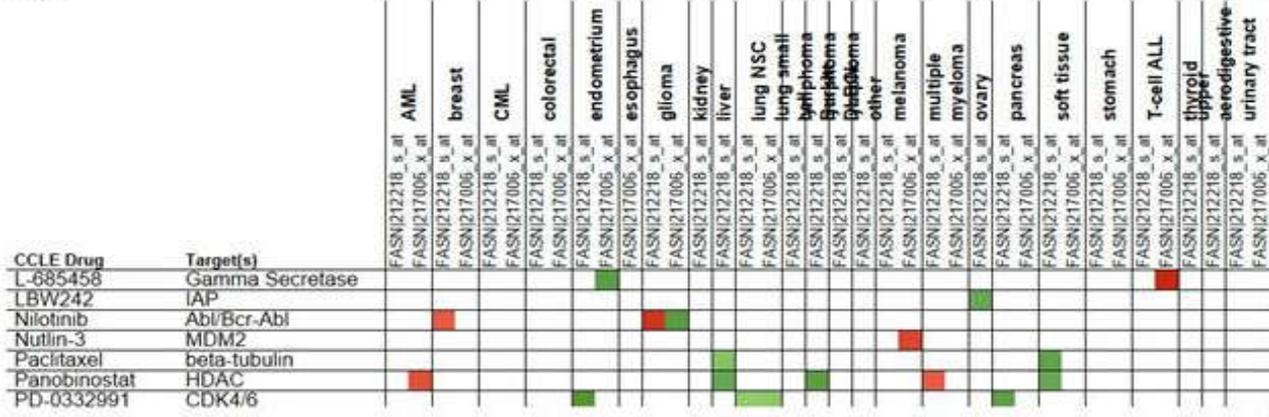


Fig. 1. FASN therapeutic response in 24 cancers with 21 anti-cancers drugs

DISCUSSION

FASN has two types i.e. cytosolic and mitochondrial both are capable to synthesize lipid contents but mitochondrial FASN induced fatty acid synthesis played significant role in mitochondrial physiology (Lee *et al.*, 2013; Menendez, 2004). The cytosolic FASN is 270 KDa multi-dimensional polypeptide protein that has 6 catalytic domains including KS, MAT, DH, ER, KR, TE and ACP that gives the X shape to FASN protein (Zeng *et al.*, 2011). The cancer cell synthesizes de novo fatty acids that develop into phospholipids which initiate signaling cascade (Kuhajda, 2006). Various studies reported the FASN overexpression resistance correlation with chemotherapeutics such as its elevated expression increased the resistance in doxorubicin-targeted breast cancer cell lines (Milgraum *et al.*, 1997). In pancreatic cancer cells FASN up-regulation acquired gemcitabine resistance that shared its post-translational regulation (Buchholz *et al.*, 2005).

The FASN knockout studies in cancer cell lines determine that cancer cells become sensitized to anti-cancer drugs such as Trastuzumab, 5-FU,  $\gamma$ -irradiation, TRAIL and DNA-damaging drugs (Ventura *et al.*, 2015; Alò *et al.*, 1999; Puig, 2011). In this study we determine the FASN overall therapeutic response to anti-cancer drugs and evaluate the resistance to various drugs in multiple tissues in a single approach. The FASN overexpression provides strong resistance to Erlotinib, TKI258, Lapatinib and Vandetanib anti-cancer drugs which mainly target the EGFR that modulate the signaling pathways of growth, survival, migration, adhesion and differentiation of tumor cells (Yewale *et al.*, 2013). The EGFR activation stimulated by ligand-induced SH2 and PTB domains receptor dimerization that further trigger STAT, AKT, PI3K and MAPAK pathway that leads to cell adhesion, survival, migration and proliferation (Cargnello *et al.*, 2011; Lim, 2006). The anti-EGFR drugs normally inhibit intracellular

kinase domain that harbor ATP binding pocket which prevent the autophosphorylation of downstream signaling pathways (Cohen, 2003). These drugs induce apoptosis, cell cycle arrest and activation of anti-growth signals (Herbst, 2002). The FASN resistance behaviors signify its functional association with EGFR mediated proliferative pathways. In EGFR therapy the FASN showed the role of secondary drug target for upcoming drugs. The PF-2341066 and PHA-665752 drugs inhibit c-MET receptor tyrosine kinase protein which promotes oncogenesis by increasing the cell motility, invasion and metastasis (Bachleitner-Hofmann *et al.*, 2008). The anti-c-MET therapy decreases the PI3K/AKT/mTOR growth and survival pathways. It also inhibits the PDGF- $\beta$ , JAK2 and ABL receptor mediated proliferation (Andrae *et al.*, 2008). The FASN overexpression showed negative correlation with these drugs that indicates their best performance to prevent the oncogenesis in the presence of FASN up-regulation. The AZD0530 and Nilotinib inhibit ABL/BCR-ABL pathway that drives various pathways of proliferation and survival (Quintás-Cardama, 2009). The BCR/ABL autophosphorylation trigger the development of GRB2/GAB2/SOS/ RAS/PI3K/ AKT/ MAPK/FOXO complex pathway of cell survival, migration and proliferation (Zhang *et al.*, 1998; Smith *et al.*, 1999; Pendergast *et al.*, 1993; Sattler, 2002; Skorski *et al.*, 1995). The FASN overexpression becomes a barrier in anti-BCR/ABL therapy due to positive resistance correlation with these drugs. The upcoming anti-cancer drugs for BCR/ABL accounted the expression of FASN as a drug target. The AZD6244 and PD-0325901 inhibit the MEK protein which is the component of MAPK pathway that is very significant in both normal/malignant states. The FASN showed resistance to anti-MEK therapy which is clear sign of its acquired oncogenic property to promote cell survival and growth. The Topotecin inhibit topoisomerase 1 which are responsible for DNA replication leads to cell division. The FASN showed strong resistance to Topotecin which indicates their gene regulatory essential role in oncogenesis. The 17-AAG inhibits HSP90 which is involved in tumor growth related protein stability (Pratt *et al.*, 2003). The FASN has resistance to HSP90 inhibitor to enhance the process of tumorigenesis. The AEW541 inhibits IGF-1R which is growth promoter and anti-apoptotic mechanism supporter. Its overexpression reported in several malignancies including lung, prostate and breast cancers (Tognon *et al.*, 2012; Chen, 2013). The FASN provides strong obstacles to anti-IGFR-1R therapy.

The determination of FASN-IGFR-1R crosstalk finds out novel targets in drug resistance. The RAF265 target RAF-B and KDR that initiate the signals to drive cell growth on large scale cellular level (Wu *et al.*, 2000). The FASN showed resistance to drug therapy and promote the onco-proteomic signaling. The Nutlin-3 inhibits anti-apoptotic MDM2 which involved in negative regulation of TP53 (Tovar *et al.*, 2006). The FASN has drug resistance to escaping of TP53 activation that makes the FASN as a component of anti-apoptotic cascade. The Panobinostat inhibit the HDAC family that involved in gene expression/regulation, cell cycle and notch signaling pathway (Rajendran *et al.*, 2013). The FASN has opposition to anti-HDAC drug to support the progression of oncogene mediated carcinogenesis. The L-685458 inhibit  $\gamma$ -secretases which is transmembrane protein played crucial role in regulation of cell cycle regulatory membrane proteins such as CD44, Notch, ErbB4 and E-Cadherins (Allenspach *et al.*, 2002; Zöller, 2011). The FASN resistance to L-685458 indicates its role in diseases other than cancer. The FASN drug

resistance mapping in 24 cancers with 21 drugs displayed its key role in cancer progression to metastasis. This approach explores the FASN functional link with cell growth, survival, differentiation, proliferation and anti-apoptotic pathways. Our work opens the channel to examine the cross-talk among FASN and these drug targets which provides synergistic associations. The FASN overexpression behaves as a predictive marker in drug resistance by large screening in scale cancer cell lines. In future the concept of combinatorial drug dosage therapy eliminates from prescriptions due to FASN type drug resistance secondary targets. This study invites the system-level molecular medicine approach to design rational drugs to inhibit co-target pathways for efficient therapy.

## REFERENCES

- Allenspach, E.J., Maillard, I., Aster, J.C. and Pear, W.S., 2002. Notch signaling in cancer. *Cancer biology and therapy*, 1(5), pp.466-476.
- Alò PL, Visca P, Trombetta G, Mangoni A, Lenti L, Monaco S, Botti C, Serpieri DE, Di Tondo U. Fatty acid synthase (FAS) predictive strength in poorly differentiated early breast carcinomas. *Tumori*. 1999;85:35–40.
- Ammer AG., Kelley LC., Hayes KE., Evans JV., Lopez-Skinner LA., Martin KH. *et al.*, 2009. Saracatinib impairs head and neck squamous cell carcinoma invasion by disrupting invadopodia function. *J Cancer Sci Ther.*;1:52–61.
- Andrae J, Gallini R, Betsholtz C. Role of platelet-derived growth factors in physiology and medicine. *Genes Dev*. 2008;11:1276–1312.
- Bachleitner-Hofmann T., Sun M.Y., Chen C.T., Tang L., Song L., Zeng Z., *et al.* (2008) HER kinase activation confers resistance to MET tyrosine kinase inhibition in MET oncogene-addicted gastric cancer cells. *Mol Cancer Ther* 7: 3499–3508
- Benjamin, D. I., Li, D. S., Lowe, W., Heuer, T., Kemble, G., and Nomura, D. K. (2015). Diacylglycerol metabolism and signaling is a driving force underlying FASN inhibitor sensitivity in cancer cells. *ACS Chemical Biology* 10(7), 1616–1623
- Bhatt, A. P., Jacobs, S. R., Freermerman, A. J., Makowski, L., Rathmell, J. C., Dittmer, D. P., and Damania, B. 2012. Dysregulation of fatty acid synthesis and glycolysis in non-Hodgkin lymphoma. *PNAS* 109(29), 11818–11823.
- Buchholz TA, Tu X, Ang KK, Esteva FJ, Kuerer HM, Pusztai L, Cristofanilli M, Singletary SE, Hortobagyi GN, Sahin AA. Epidermal growth factor receptor expression correlates with poor survival in patients who have breast carcinoma treated with doxorubicin-based neoadjuvant chemotherapy. *Cancer*. 2005;104:676–81
- Cai, Y., Wang, J., Zhang, L., Wu, D., Yu, D., Tian, X., Huang, P. 2014. Expressions of fatty acid synthase and HER2 are correlated with poor prognosis of ovarian cancer. *Medical Oncology* 32(1), 391.
- Cargnello, M. and Roux, P.P., 2011. Activation and function of the MAPKs and their substrates, the MAPK-activated protein kinases. *Microbiology and molecular biology reviews*, 75(1), pp.50-83.
- Chen, H.X. and Sharon, E., 2013. IGF-1R as an anti-cancer target—trials and tribulations. *Chinese journal of cancer*, 32(5), p.242.
- Chirala, S. S., Chang, H., Matzuk, M., Abu-Elheiga, L., Mao, J., Mahon, K., Wakil, S. J. 2003. Fatty acid synthesis is essential in embryonic development: Fatty acid synthase null mutants and most of the heterozygotes die in utero. *Proceedings of the National Academy of Sciences of the United States of America* 100(11), 6358–6363.

- Chougnat CN., Borget I., Lebouilleux S., de la Fouchardiere C., Bonichon F., Criniere L. *et al.*, 2015. Vandetanib for the treatment of advanced medullary thyroid cancer outside a clinical trial: results from a French cohort. *Thyroid* 25:386–91.10.1089
- Christensen JG., Schreck R., Burrows J., Kuruganti P., Chan E., Le P., Chen J., Wang X., Ruslim L., Blake R., Lipson KE., Ramphal J., Do S., Cui JJ., Cherrington JM., Mendel DB. 2003. A selective small molecule inhibitor of c-Met kinase inhibits c-Met-dependent phenotypes in vitro and exhibits cytoreductive antitumor activity in vivo. *Cancer Res.*, 63:7345–55
- Cohen RB 2003 Epidermal growth factor receptor as a therapeutic target in colorectal cancer. *Clinical Colorectal Cancer* 2 246–251.
- Conacci-Sorrell, M., Zhurinsky, J. and Ben-Ze'ev, A., 2002. The cadherin-catenin adhesion system in signaling and cancer. *The Journal of clinical investigation*, 109(8), pp.987-991
- Cui JJ., Tran-Dube M., Shen H., Nambu M., Kung PP., Pairish M. *et al.* 2011. Structure based drug design of crizotinib (PF-02341066), a Potent and selective dual inhibitor of Mesenchymal-Epithelial Transition Factor (c-MET) Kinase and Anaplastic Lymphoma Kinase (ALK) *J Med Chem.*, 54:6342–63.
- Dimopoulos, M.A., Mitsiades, C.S., Anderson, K.C. and Richardson, P.G., 2011. Tanespimycin as antitumor therapy. *Clinical Lymphoma Myeloma and Leukemia*, 11(1), pp.17-22.
- Dong M., Rice L., Lepler S., Pampo C., Siemann DW. 2010. Impact of the Src inhibitor saracatinib on the metastatic phenotype of a fibrosarcoma (KHT) tumor model. *Anticancer Res.*, 30:4405–13.
- Engebraaten O., Bjerkvig R., Pedersen PH., *et al.*, 19993. Effects of EGF, bFGF, NGF and PDGF(bb) on cell proliferative, migratory and invasive capacities of human brain-tumor biopsies in vitro. *Int J Cancer.*, 53:209–14.
- Fallahi P., Di Bari F., Ferrari SM., Spisni R., Materazzi G., Miccoli P. *et al.*, 2015. Selective use of vandetanib in the treatment of thyroid cancer. *Drug Des Devel Ther.*, 9:3459–70.10.2147
- Finn, R.S., Dering, J., Conklin, D., Kalous, O., Cohen, D.J., Desai, A.J., Ginther, C., Atefi, M., Chen, I., Fowst, C. and Los, G., 2009. PD 0332991, a selective cyclin D kinase 4/6 inhibitor, preferentially inhibits proliferation of luminal estrogen receptor-positive human breast cancer cell lines in vitro. *Breast Cancer Research*, 11(5), p.R77.
- Fiorentino, M., Zadra, G., Palescandolo, E., Fedele, G., Bailey, D., Fiore, C., Loda, M. 2008. Overexpression of fatty acid synthase is associated with palmitoylation of Wnt1 and cytoplasmic stabilization of  $\beta$ -catenin in prostate cancer. *Laboratory Investigation* 88(12), 1340–1348.
- Ganguly, A., Yang, H. and Cabral, F., 2010. Paclitaxel-dependent cell lines reveal a novel drug activity. *Molecular cancer therapeutics*, pp.1535-7163.
- García-Echeverría, C., Pearson, M.A., Marti, A., Meyer, T., Mestan, J., Zimmermann, J., Gao, J., Brueggen, J., Capraro, H.G., Cozens, R. and Evans, D.B., 2004. In vivo antitumor activity of NVP-AEW541—a novel, potent, and selective inhibitor of the IGF-IR kinase. *Cancer cell*, 5(3), pp.231-239.
- Gaur, R., K Yadav, D., Kumar, S., P Darokar, M., Khan, F. and Singh Bhakuni, R., 2015. Molecular modeling based synthesis and evaluation of in vitro anticancer activity of indolylchalcones. *Current topics in medicinal chemistry*, 15(11), pp.1003-1012.
- Gonzalez-Guerrico, A. M., Espinoza, I., Schroeder, B., Park, C. H., Kvp, C. M., Khurana, A., Lupu, R. 2016. Suppression of endogenous lipogenesis induces reversion of the malignant phenotype and normalized differentiation in breast cancer. *Oncotarget* 7(44), 71151–71168.
- Graner, E., Tang, D., Rossi, S., Baron, A., Migita, T., Weinstein, L. J., Loda, M. 2004. The isopeptidase USP2a regulates the stability of fatty acid synthase in prostate cancer. *Cancer Cell* 5(3), 253–261
- Hansmannel, F., Mordier, S., and Iynedjian, P. B. 2006. Insulin induction of glucokinase and fatty acid synthase in hepatocytes: Analysis of the roles of sterol-regulatory-elementbinding protein-1c and liver X receptor. *Biochemical Journal* 399(2), 275–283.
- Herbst RS and Hong WK 2002 IMC-C225, an anti-epidermal growth factor receptor monoclonal antibody for treatment of head and neck cancer. *Seminars in Oncology* 29 18–30.
- Herbst RS., Johnson DH., Mininberg E., *et al.*, 2005b. Phase I/II trial evaluating the anti-vascular endothelial growth factor monoclonal antibody bevacizumab in combination with the HER-1/epidermal growth factor receptor tyrosine kinase inhibitor erlotinib for patients with recurrent non-small cell lung cancer. *J ClinOncol.*, 23:2544–55
- Heuer, T. S. 2016. De novo palmitate synthesis supports oncogenic signalling and tumor growth through diverse mechanisms: Implications for FASN-targeted therapeutics. *Journal of Cell Signaling* 1(3), 1–4.
- Ishii, S., Iizuka, K., Miller, B. C., and Uyeda, K. 2004. Carbohydrate response element binding protein directly promotes lipogenic enzyme gene transcription. *Proceedings of the National Academy of Sciences of the United States of America* 101(44), 15597–15602
- Jensen-Urstad, A. P. L., Song, H., Lodhi, I. J., Funai, K., Yin, L., Coleman, T., and Semenkovich, C. F. 2013. Nutrient-dependent phosphorylation channels lipid synthesis to regulate PPAR. *The Journal of Lipid Research* 54(7), 1848–1859
- Knobloch, M., Braun, S. M. G., Zurkirchen, L., von Schoultz, C., Zamboni, N., Araúzo-Bravo, M. J., Jessberger, S. 2012. Metabolic control of adult neural stem cell activity by Fasn-dependent lipogenesis. *Nature* 493(7431), 226–230.
- Kuhajda FP. Fatty acid synthase and cancer: new application of an old pathway. *Cancer Res.* 2006;66:5977–80
- Kumar R., Crouthamel M. C., Rominger D. H., Gontarek R. R., Tummino P. J., Levin R. A., King A. G. Myelosuppression and kinase selectivity of multikinase angiogenesis inhibitors. *Br. J. Cancer* 2009, 101, 1717–172310
- Kwak EL., Bang YJ., Camidge DR., Shaw AT., Solomon B., Maki RG. *et al.* 2010. Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N Engl J Med.*, 363:1693–703.
- Lee JS, Sul JY, Park JB, Lee MS, Cha EY, Song IS, Kim JR, Chang ES. (2013). Fatty acid synthase inhibition by amentoflavone suppresses HER2/neu (erbB2) oncogene in SKBR3 human breast cancer cells. *Phytother Res.* 27:713–20
- Léger, F., Loos, W.J., Bugat, R., Mathijssen, R.H., Goffinet, M., Verweij, J., Sparreboom, A. and Chatelut, E., 2004. Mechanism-based models for topotecan-induced neutropenia. *Clinical Pharmacology and Therapeutics*, 76(6), pp.567-578.
- Li WQ., Kawakami K., Ruszkiewicz A., Bennett G., Moore J., *et al.*, 2006 BRAF mutations are associated with distinctive clinical, pathological and molecular features of colorectal cancer independently of microsatellite instability status. *Mol Cancer*. 5. 2 p. 1476-4598-5-2
- Li, J., and Cheng, J. -X. 2014. Direct visualization of de novo lipogenesis in single living cells. *Scientific Reports* 4, 6807
- Lim, C.P. and Cao, X., 2006. Structure, function, and regulation of STAT proteins. *Molecular biosystems*, 2(11), pp.536-550.
- Manley, P.W., Stiefl, N., Cowan-Jacob, S.W., Kaufman, S., Mestan, J., Wartmann, M., Wiesmann, M., Woodman, R. and Gallagher, N., 2010. Structural resemblances and comparisons of the relative pharmacological properties of imatinib and nilotinib. *Bioorganic and medicinal chemistry*, 18(19), pp.6977-6986.

- Menendez JA, Lupu R. (2004). Fatty acid synthase-catalyzed de novo fatty acid biosynthesis: from anabolic-energy-storage pathway in normal tissues to jack-of-all-trades in cancer cells. *ArchImmunolTher Exp.* 52:414–26
- Migita, T., Ruiz, S., Fornari, A., Fiorentino, M., Priolo, C., Zadra, G., Loda, M. 2009. Fatty acid synthase: A metabolic enzyme and candidate oncogene in prostate cancer. *Journal of the National Cancer Institute* 101(7), 519–532.
- Milgraum LZ, Witters LA, Pasternack GR, Milgraum Z, Pasternack R, Kuhajda F. Enzymes of the fatty acid synthesis pathway are highly expressed in in situ breast carcinoma. *Clin Cancer Res.* 1997;3:2115–20.
- Moy B., Goss PE. 2007. Lapatinib-associated toxicity and practical management recommendations. *Oncologist* 2007; 12(7): 756–65
- Nguyen, P. L., Ma, J., Chavarro, J. E., Freedman, M. L., Lis, R., Fedele, G., Loda, M. 2010. Fatty acid synthase polymorphisms, tumor expression, body mass index, prostate cancer risk, and survival. *Journal of Clinical Oncology* 28(25), 3958–3964
- Pendergast AM, Quilliam LA, Cripe LD, et al. BCR-ABL-induced oncogenesis is mediated by direct interaction with the SH2 domain of the GRB-2 adaptor protein. *Cell.* 1993;75:175–185.
- Pratt, W.B. and Toft, D.O., 2003. Regulation of signaling protein function and trafficking by the hsp90/hsp70-based chaperone machinery. *Experimental biology and medicine*, 228(2), pp.111-133.
- Puig T, Aguilar H, Cufí S, Oliveras G, Turrado C, Ortega-Gutiérrez S, Benhamú B, López-Rodríguez ML, Urruticochea A, Colomer R. A novel inhibitor of fatty acid synthase shows activity against HER2+ breast cancer xenografts and is active in anti-HER2 drug-resistant cell lines. *Breast Cancer Res.* 2011;13:R131.
- Quintás-Cardama, A. and Cortes, J., 2009. Molecular biology of bcr-abl1-positive chronic myeloid leukemia. *Blood*, 113(8), pp.1619-1630.
- Rajendran, P., Kidane, A.I., Yu, T.W., Dashwood, W.M., Bisson, W.H., Löhr, C.V., Ho, E., Williams, D.E. and Dashwood, R.H., 2013. HDAC turnover, CtIP acetylation and dysregulated DNA damage signaling in colon cancer cells treated with sulforaphane and related dietary isothiocyanates. *Epigenetics*, 8(6), pp.612-623.
- Rikova K., Guo A., Zeng Q., Possemato A., Yu J., Haack H., et al. 2007. Global survey of phosphotyrosine signaling identifies oncogenic kinases in lung cancer. *Cell.*, 131:1190–203.
- Röhrig, F., and Schulze, A. 2016. The multifaceted roles of fatty acid synthesis in cancer. *Nature Reviews Cancer* 16(11), 732–749
- Rysman, E., Brusselmans, K., Scheys, K., Timmermans, L., Derua, R., Munck, S., Swinnen, J. V. 2010. De novo lipogenesis protects cancer cells from free radicals and chemotherapeutics by promoting membrane lipid saturation. *Cancer Research* 70(20), 8117–8126
- Rysman, E., Brusselmans, K., Scheys, K., Timmermans, L., Derua, R., Munck, S., Swinnen, J. V. 2010. De novo lipogenesis protects cancer cells from free radicals and chemotherapeutics by promoting membrane lipid saturation. *Cancer Research* 70(20), 8117–8126
- Sattler M, Mohi MG, Pride YB, et al. Critical role for Gab2 in transformation by BCR/ABL. *Cancer Cell.* 2002;1:479–492
- Shearman, M.S., Beher, D., Clarke, E.E., Lewis, H.D., Harrison, T., Hunt, P., Nadin, A., Smith, A.L., Stevenson, G. and Castro, J.L., 2000. L-685,458, an aspartyl protease transition state mimic, is a potent inhibitor of amyloid  $\beta$ -protein precursor  $\gamma$ -secretase activity. *Biochemistry*, 39(30), pp.8698-8704.
- Skorski T, Kanakaraj P, Nieborowska-Skorska M, et al. Phosphatidylinositol-3 kinase activity is regulated by BCR/ABL and is required for the growth of Philadelphia chromosome-positive cells. *Blood.* 1995;86:726–736
- Smith JM, Katz S, Mayer BJ. Activation of the Abl tyrosine kinase in vivo by Src homology 3 domains from the Src homology 2/Src homology 3 adaptor Nck. *J Biol Chem.* 1999;274:27956–27962.
- Tognon, C.E. and Sorensen, P.H., 2012. Targeting the insulin-like growth factor 1 receptor (IGF1R) signaling pathway for cancer therapy. *Expert opinion on therapeutic targets*, 16(1), pp.33-48.
- Tovar, C., Rosinski, J., Filipovic, Z., Higgins, B., Kolinsky, K., Hilton, H., Zhao, X., Vu, B.T., Qing, W., Packman, K. and Myklebost, O., 2006. Small-molecule MDM2 antagonists reveal aberrant p53 signaling in cancer: implications for therapy. *Proceedings of the National Academy of Sciences*, 103(6), pp.1888-1893.
- Vazquez-Martin, A., Colomer, R., Brunet, J., Lupu, R., and Menendez, J. A. 2008. Overexpression of fatty acid synthase gene activates HER1/HER2 tyrosine kinase receptors in human breast epithelial cells. *Cell Proliferation* 41(1), 59–85
- Ventura R, Mordec K, Waszczuk J, Wang Z, Lai J, Fridlib M, Buckley D, Kemble G, Heuer TS. Inhibition of de novo palmitate synthesis by fatty acid synthase induces apoptosis in tumor cells by remodeling cell membranes, inhibiting signaling pathways, and reprogramming gene expression. *EBioMedicine.* 2015;2:806–22.
- Wang, J., Zhang, X., Shi, J., Cao, P., Wan, M., Zhang, Q., Sui, G. 2016. Fatty acid synthase is a primary target of MiR-15a and MiR-16-1 in breast cancer. *Oncotarget* 5(0).
- Wong DW., Leung EL., So KK., Tam IY., Sihoe AD., Cheng LC., et al. 2009. The EML4-ALK fusion gene is involved in various histologic types of lung cancers from nonsmokers with wild-type EGFR and KRAS. *Cancer.* 115:1723–33
- Wu, L.W., Mayo, L.D., Dunbar, J.D., Kessler, K.M., Ozes, O.N., Warren, R.S. and Donner, D.B., 2000. VRAP is an adaptor protein that binds KDR, a receptor for vascular endothelial cell growth factor. *Journal of Biological Chemistry*, 275(9), pp.6059-6062.
- Yeh TC, Marsh V, Bernat BA, et al. Biological characterization of ARRY-142886 (AZD6244), a potent, highly selective mitogen-activated protein kinase kinase 1/2 inhibitor. *Clin Cancer Res.* 2007;13(5):1576–1583.
- Yewale, C., Baradia, D., Vhora, I., Patil, S. and Misra, A., 2013. Epidermal growth factor receptor targeting in cancer: a review of trends and strategies. *Biomaterials*, 34(34), pp.8690-8707.
- Zaytseva, Y. Y., Elliott, V. A., Rychahou, P., Mustain, W. C., Kim, J. T., Valentino, J., Evers, B.M. 2014. Cancer cell-associated fatty acid synthase activates endothelial cells and promotes angiogenesis in colorectal cancer. *Carcinogenesis* 35(6), 1341–1351
- Zeng XF, Li WW, Fan HJ, Wang XY, Ji P, Wang ZR, Ma S, Li LL, Ma XF, Yang SY. (2011). Discovery of novel fatty acid synthase (FAS) inhibitors based on the structure of ketoacyl synthase (KS) domain. *Bioorg Med Chem Lett.* 21:4742–4744.
- Zhang X, Ren R. Bcr-Abl efficiently induces a myeloproliferative disease and production of excess interleukin-3 and granulocyte-macrophage colony-stimulating factor in mice: a novel model for chronic myelogenous leukemia. *Blood.* 1998;92:3829–3840.
- Zöller, M., 2011. CD44: can a cancer-initiating cell profit from an abundantly expressed molecule?. *Nature Reviews Cancer*, 11(4), p.254.