

Available online at http://www.journalcra.com

International Journal of Current Research Vol. 7, Issue, 04, pp.14542-14551, April, 2015 INTERNATIONAL JOURNAL OF CURRENT RESEARCH

RESEARCH ARTICLE

EXPOSURE TO BISPHENOL A IN DENTISTRY - CURRENT VIEWS

^{*,1}Maya Grigorievna Lyapina, ²Maria Dencheva – Garova, ²Assya Krasteva – Panova, ²Mariana Tzekova – Yaneva, ³Mariela Yaneva – Deliverska and ²Angelina Kisselova-Yaneva

¹Department "Hygiene, Medical Ecology and Nutrition" Medical Faculty, Medical University, Sofia, Bulgaria ²Department "Oral and Image Diagnostic", Faculty of Dental Medicine, Medical University, Sofia, Bulgaria ³Military Medical Academy, Sofia, Bulgaria

ARTICLE INFO	ABSTRACT
<i>Article History:</i> Received 29 th January, 2015 Received in revised form 14 th February, 2015 Accepted 22 nd March, 2015 Published online 28 th April, 2015	Bisphenol a is xenoestrogen synthesized in large quantities worldwide for production of polymers (polycarbonates, epoxy resins) and thermal paper. This determines its universal presence - in everyday products (packaging, containers and bottles), food and drinking water. Food is considered to be the most important source of population exposure; however, in overall exposure assessment consumption of drinking water, inhalation of dust and dermal contact with thermal paper must be taken into account. Exposure to BPA and its derivatives from dental composites and sealants is possible. High levels of BPA in saliva (especially immediately or one hour after dental treatment), decreasing over time have been found. No BPA in the blood samples of dental patients have been
Key words:	
Bisphenol A, Adverse effects, Dental materials, Dentistry.	detected, as reported in the available studies. High urinary levels of BPA after treatment with dental composites and sealants have been reported. The degree of exposure to BPA from dental materials and the possible adverse health effects are insufficiently investigated. No data were found in the available literature concerning the urinary levels of BPA among occupationally exposed dental professionals, in comparison with those among dental patients after treatment with composites and sealants.

Copyright © 2015 Maya Grigorievna Lyapina et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

This appears to be interesting field for further investigations

Bisphenol A [2, 2 b-bis (4- hydroxyphenyl) propane; CAS 80-05-7], is synthesized by condensation of two phenol groups and one molecule acetone.



Bisphenol A (**BPA**) is a white, crystalline substance with a molecular weight of 228.29 g/cm³, melting point 156°C and boiling point 220°C, good solubility in fats and low solubility in water. Its solubility is higher at an alkaline pH. The presence of hydroxyl groups (Fig. 1) determines the good reactivity of Bisphenol A (Flint *et al.*, 2012 and Volkel *et al.*, 2002). Bisphenol A was first synthesized in 1891 by the Russian chemist A. Dianin and Thomas Zincke from the university in

Marburg, Germany, who published a note of its synthesis in 1905 (Zincke, 1905). In 1953, Hermann Schnell – Bayer, Germany and Dan Fox- General electric, USA developed the technology for synthesis of a new plastic material – polycarbonate, using BPA as row material. Since then, BPA has been used mainly as:

- monomer in the production of numerous polymers polycarbonate (PC) plastics, epoxy resins, polysulfones, and polyacrilates;
- antioxidant and inhibitor in the manufacture of polyvynil chloride (PVC) plastics;
- precursor for the synthesis of tetrabromobiphenol-A (Geens *et al.*, 2011).

Currently, polycarbonates are widely used for the manufacture of products intended for contact with food - plastic bottles, plates, cups, fireproof containers for microwave ovens, storage containers, etc., and epoxy resins used for internal coating of cans (EFSA, 2006). However, only 3% of totally produced polycarbonates and 10% of epoxy resins are used as materials for contact with food products (Plastics Europe, 2007). There are many other uses of polycarbonates, epoxy resins, polysulfones and polyacrylates, e.g. for manufacture of spectacle lenses, digital media (CD and DVD), mobile phones,

^{*}Corresponding author: Maya Grigorievna Lyapina,

Department "Hygiene, Medical Ecology and Nutrition" Medical Faculty, Medical University, Sofia, Bulgaria.

electronics, computers and other electrical equipment, household appliances, building glass, safety equipment in sport, cars, construction materials, medical equipment, dental materials, thermal print paper (Geens et al., 2011). Thermal paper is produced in large amounts as it is used for receipts, faxes, labels, and after recycling - for brochures, tickets, envelopes, newspapers, kitchen towels, toilet paper and cardboard packaging for food (Liao and Kannan, 2011 and Nam et al., 2010). In addition, BPA is widely used in the production of polyacrylates, polyesters and varnishes which, after degradation, could be an important source of this compound in environment and food (Vandenberg et al., 2007). Through condensation of a ketone or aldehyde with phenols or by alterations of carbonyl derivatives numerous analogues of bisphenol can be synthesized. Many of them are too expensive for wide industrial applications. Among them, widely used is bisphenol-F (BPF) because of the lower viscosity and better solvent resistance compared to BPA (Danzl et al., 2009). Bisphenol-S (BPS) is also used as a monomer in plastics industry. Typical for polycarbonate plastics is the unique combination of properties such as optical transparency, shockresistance and high heat resistance. These characteristics contribute for the diversity of applications of polycarbonate. It is calculated that the current annual production of BPA is about 3.8 million tons.

Routes of entry into environment and foods

The presence of BPA in the environment is associated with anthropogenic activities. It is difficult to define the most important sources for human exposure. It's considered that BPA enters the ecosystems and foods as a consequence of production, processing, and degradation (hydrolysis) of different polymers, e.g. epoxy resins and polycarbonates (Mercea, 2009).

Pharmacokinetics and metabolism

Pharmacokinetics of BPA have been studied in rodents, primates and humans (Doerge et al., 2010; Doerge et al., 2010; Völkel et al., 2005 and Vőlkel et al., 2008). After oral administration, BPA is subjected to rapid initial metabolism in the liver and intestines, and is completely absorbed in the gastrointestinal tract. In a subsequent phase, it binds to glucuronic acid to form BPA- glucuronide. Small amounts of BPA can react with sulfates to form bisphenol-sulfate. The glucoronide, a metabolite that has no known biological activity and, in particular, has been shown to be non-estrogenic, was cleared from the blood and excreted by the urine within the day of exposure (Matthews et al., 2001; Snyder et al., 2000; Völkel et al., 2005 and Vőlkel et al., 2008). The applied doses are excreted entirely; therefore, exposure to BPA may be assessed by its urinary levels (Vőlkel et al., 2008). Upon inhalation or skin exposure to BPA elimination is slower.

Environmental exposure

Air

BPA is presented in the atmosphere in various concentrations, mostly as a result of industrial activities. Berkner *et al.* (2004),

in a study performed in the region of Bavaria established low BPA concentration (5-15 pg/m³), while Matsumoto *et al.* (2005) established significant concentrations (10-1920 pg/m³) in Osaka. The concentrations of BPA were found to be low in marine areas, highest being over the east coast of Asia. Significantly higher concentrations (170-880 pg/m³) were measured over large urban agglomerations in Asia, New Zealand and USA. Highest concentrations (4.55 ng/m³) have been measured in the urban parts of India (Bombay), resulting from the intense burning of plastic products for household use. The polar region was also found to be polluted with BPA (in concentrations 1-17 pg/m³), most likely origination from the middle latitudes of Eurasia and North America. For the period 1991-2000, an increase in concentration in this region has been observed (Fu and Kawamura, 2010).

Water

Bisphenol A is usually presented in low concentrations in surface water. Rocha et al. (2013) established BPA in about half of the rivers in Portugal in concentrations 28.7-98.4 ng/dm³. Some studies indicate higher degree of pollution of surface water. In a study conducted in Germany, the levels of BPA in the Elbe were 4-92 μ g/ dm³ and its sediments -10-380 $\mu g/dm^3$ (Stachel *et al.*, 2003). It is considered that the main source of pollution is the industrial waste water (Lee and Peart, 2000). Cladiere et al. (2013) evaluated the impact of highly urbanized environment (Paris) for water pollution with BPA. The authors consider as main sources of BPA in urban waste water: atmospheric pollution (10-180 ng/dm³); sewer overflows (917-2098 ng/dm³); and effluents (287-1224 ng/dm³). Significantly higher concentrations of BPA have been found in groundwater near contaminated landfills or such with plastic materials. The average groundwater concentration of BPA near the landfill in Osaka was 740 µg/dm³ (Kawagoshi et al., 2003). High concentrations are found in industrial sewage as well. In a large-scale study of BPA contamination in Canada, Lee et al. (2000) measured in raw sewage sludge average BPA concentrations of 36.7 mg/kg (Lee and Peart, 2000).

Food and drinking water

The most important source of human exposure to BPA is by food. Its presence in food is in relation to the exposure of animal and vegetable raw materials, with accumulation in the environment and with the foods contact with polymers that contain it. The daily exposure to BPA with consumed food is evaluated to be 0.48-1.6 µg/kg body weight (Vőlkel et al., 2008). Several studies indicate that BPA can be released from polycarbonate and epoxy resins into food and drinking water. Bisphenol A is found in significant concentrations in meat products (0.49-56 µg/kg) (Shao et al., 2007), fish (7.1-102.7 µg/kg) (Munguia-Lopez et al., 2005), vegetables and fruits (11.0-95.3 µg/kg) and cereals (1.0-3.8 µg/kg) (Niu et al., 2012 and Yoshida et al., 2001). Studies demonstrate that BPA concentrations are significantly higher in cans than in fresh food. Yonekubo et al. (2008) found BPA in various canned products in concentrations of 0.1 -235.4 μ g / kg. Relatively high concentrations (3.7 - 265.6 µg/kg) were found in canned vegetables and fruits (Cunha and Fernandes, 2013). BPA was

also established in canned seafood in concentrations of 1.0 -99.9 µg/kg (Cunha et al., 2012) and in minor concentrations (0.032-4 µg/kg) in canned soft drinks (Cao et al., 2009). It has also been found in samples of milk. O'Mahony et al. (2013) analyzed 27 samples of commercially available dairy products, including stored in cartons milk, infant formula milk and condensed milk. In just four of the samples the concentrations were above the limit (1.32 μ g/kg), and one of the samples (tin of condensed milk) contained high concentration of BPA -176 µg/kg. An overall analysis of data available from 65 scientific publications, concerning the level of BPA in drinking water, indicates that its concentrations are low in comparison with other sources. Data indicate that in North America, Europe and Asia they were as follow: 0.099, 0.014, and 0.317 µg/dm³ (Arnold et al., 2013). Higher levels were found in bottled water. In a recent study, Colin et al. (2013) analyzed the exposure of the French population to BPA from drinking water. The concentrations ranged from 0.07 to 4.21 μ g/dm³ in PC bottles water, with levels being higher in newly produced ones.

Inhalational exposure

Other possible routes of exposure to BPA are through inhalation and the skin. Due to the migration of polymer products, BPA can be found in environmental dust. It is unlikely that inhalation exposure is significant, due to low vapour pressure and therefore low BPA concentrations in the air (Dekant and Völkel, 2008). Inhalation exposure to BPA from household dust is considered to be important for young children, frequently contacting their hands with mouth (Jones-Otazo et al., 2005). The use of BPA in consumer products such as epoxy based flooring, adhesives, paints, electronic equipment, etc. is widespread, so its evaporation and/or leaching from them could be a source of indoor dust pollution (Loganathan and Kannan, 2011). In a study, 56 dust samples from different parts of the United States were collected and analysed for BPA content. Significant levels, ranging from 0.5 to 10.2 mg/kg were found in 95% of them. The authors evaluated the exposure doses and concluded that they are close to those causing health effects in experimental animals (Loganathan and Kannan, 2011). Another study, conducted in Belgium showed that the concentrations of BPA in office dust were significantly higher if compared to that in house dust, probably due to the widespread use of electronic equipment in offices. It is generally considered that inhalation exposure of the population is significantly lower than exposure associated with food consumption (Geens et al., 2009).

Dermal exposure

Bisphenol A is used as a colour developer in thermal paper. One side of the paper is coated with a powder layer of BPA, which reacts with the ink when exposed to heat or pressure. Thermal paper is used mainly for devices such as cash registers and ATMs (Lassen *et al.*, 2011). The population is in everyday contact with BPA as a component of thermal paper, which this way contributes to the overall oral (through direct contact of unwashed hands with food or the mouth) or dermal exposure. Thermal paper is the main source of contamination of recycled paper with BPA (88,102). Braun *et al.* (2011) reported increased urinary levels of BPA in cashiers with frequent dermal contact with thermal paper (Braun *et al.*, 2011). More than 80% of paper products such as brochures, tickets, newspapers, and toilet paper contain BPA in concentrations up to 14.4 μ g/g, i.e. 3-4 times lower than those in thermal paper (53). The average dermal exposure for general population is estimated at 17.4 ng/day, while among the occupationally exposed it is 1303 ng/day. The contact with thermal paper contributes to more than 98 % of total dermal exposure.

Medical devices and BPA

Some polycarbonate and polysulfone BPA-based polymers are used for production of medical devices - transfusion and dialysis equipment, filters, pumps, bypass devices, spectacle lenses, surgical instruments, inhalers, oxygenators, incubators etc. (Geens *et al.*, 2011). Calafat *et al.* (2009) established that the urine BPA concentration among preterm infants is higher than the mean concentrations among the general population (children 6-11 years of age). BPA could be found in medicines (mostly liquid) in metal packing polycarbonate or with epoxy lining (FDA, 2009).

Dental materials and BPA

Dental composite resins are composed of different ingredients - bisphenol-A glycidyl methacrylate (Bis-GMA), bisphenol-A dimethacrylate (Bis-DMA), ethylene glycol dimethacrylate (EGDMA), urethane - dimethacrylate (UDMA) and triethylene glycol dimethacrylate (TEGDMA). Bisphenol A is a row material for production of Bis-DMA and Bis -GMA, and therefore it is possible that minor amounts of BPA are presenting in dental materials, containing such compounds (Fleisch et al., 2010; Fung et al., 2000; Nathanson et al., 1997 and Tarumi et al., 2000). These substances are main ingredients of dental restorative materials - composites, sealants, cements, crowns, orthodontic brackets etc. (Schmalz et al., 1999 and Tarumi et al., 2000). As a result of bis-DMA hydrolysis by the salivary esterases, after treatment with such materials BPA was detected in patient's saliva (Van Landuyt et al., 2012).

In summary, BPA may present in dental products in any of the following ways:

- As an ingredient, despite manufacturers' claims that BPA is used "rarely" in the production of dental materials;
- As by-product from the degradation of components of resin-based dental materials. These are a mixture of monomers, usually based on Bis-GMA. Some composite resins may contain other monomer, added in order to modify their properties. An example is bisphenol A dimethacrylate (Bis-DMA). Materials containing bis-DMA can release BPA after their degradation by salivary enzymes.
- In minimal quantities "traces". As mentioned, BPA is a row material for production of Bis-DMA and Bis-GMA, so trace amounts may present in the final dental material (http://www.ada.org/271.aspx).

Currently, no studies are available to determine directly and quantitatively "the contribution" of dental composite materials for the total BPA exposure (Fleisch et al., 2010). The main routes of entry of BPA from dental materials are as follows: through the gastrointestinal tract, through the pulp by the dentinal tubules and through inhalation of volatile compounds (Gerzina and Hume, 1996; Reichl et al., 2008; Rogalewicz et al., 2006 and Van Landuvt et al., 2012). It is assumed that nonpolymerized monomers are responsible for side effects manifestations among dental patients (Mohsen et al., 1998). The aaguatic environment in oral cavity enhances the chemical degradation thus changing the mechanical properties of dental composites (Cilli et al., 2012). It is assumed that, even under optimal conditions, 23-65 % of monomers are not polymerized and are free to enter the oral cavity (Polydorou et al., 2009). It is considered that higher amounts are released from polymerized than from non-polymerized composites (Olea et al., 1996 and Pulgar et al., 2000). Release of BPA from Bis-DMA-containing sealants was established (Fleisch et al., 2010 and Schmalz and Arenholt-Bindslev, 2009). Some of the analytical methods used for BPA evaluation are subjected to re-evaluation because of the conflicting results achieved (Imai and Komabayashi, 2000; Imai, 2000; Myers and Hutz, 2011; Olea et al., 1996 and Pulgar et al., 2000).

Due to its chemical structure, Bis-GMA is considered to be protected from the action of salivary esterases (Azarpazhooh and Main, 2008). The oxidation of polymer matrix, resulting in formaldehyde release, can also lead to BPA release (Bakopoulou et al., 2009). In vivo studies assessed salivary BPA levels after application of dental sealants. Salivary BPA levels decreased over time; highest exposures were established immediately after application. None of these studies detected BPA three hours later. A possible explanation is that analytical methods used in these studies are not sufficiently sensitive to detect the extremely low doses of BPA released from the resin.Therefore chronic exposure to low doses cannot be excluded (Fleisch et al., 2010). In 1996, Olea et al. establish high concentrations of BPA in saliva after sealant (Delton, Dentsply) application (Olea et al., 1996). BPA in saliva and urine of patients after application of fissure sealant was reported, with significant differences between the different brands (Joskow et al., 2006).

Sasaki et al. reported increased levels of BPA in saliva after treatment with composite dental materials, with levels turning to the baseline within 24 hours, when patients were rinsing the oral cavity with water (Sasaki et al., 2005). Van Landuyt investigated in vitro the levels of BPA released by dental materials (Van Landuyt et al., 2011). It was estimated that a molar crown can release 13 µg - 30 mg BPA after 24 hours. Release from dental materials for 24 hours may be of importance in patients with multiple or big restorations and dental resin-based materials can be a relevant source of BPA in such patients. The concentrations widely vary among manufacturers (Vandenberg et al., 2010). According to Von Goetz et al. (2010) in chronic exposure (after dental surgical procedures) the levels are 215 ng BPA/a day. This is probably the worst scenario of chronic exposure because the concentration in saliva decreases over time, and after 120 hours still detectable concentrations where found in only one

person. Kingman et al. (2012) used liquid chromatography/ mass spectrometry to determine the concentrations of BPA and five related compounds in saliva and urine in 171 persons before and 30 hours after treatment with composite restorations. Salivary concentrations of BPA and related compounds increased immediately (within one hour) after placing the composites and returned to baseline levels within eight hours. Except for the increase in BPA, the concentrations of tested compounds in urine reached baseline levels within 30 hours after insertion of the restorations. The changes in the levels of BPA in saliva and urine were similar when anterior and posterior restorations were placed. The authors concluded that treatment with composite restorations is associated with increased levels of BPA and other similar within one hour compounds after placement in saliva, and after 9-30 hours - in urine. In another study, BPA concentration in the of 19 children was examined – before, 1 hour, 24 hours, 7 and 14 days after treatment. The results demonstrated quick increase from the baseline levels (0.26 ng/ml) after 24 hours (1.18 ng/ml), with a peak concentrations detected at 7^{th} day (1.21) ng/ml). Concentrations have not reached the baseline yet 14 days after treatment (0.73 ng/ml) (Martin et al., 2005). In an in vitro study, simulating occupational environment during grinding composite materials, MCF- 7 cells were exposed to their extract for one month at 37°C. Rapid growth has been observed, suggesting estrogenic activity. The clinical significance of these findings is unclear, but according to the authors dental professionals can be occupationally exposed to such aerosols several times per day (Van Landuvt et al., 2012).

Preventive measures in dental practice

In dental practice, it is most likely that the exposure to potentially toxic substances will occur during processing of the surface (oxygen - inhibited) layer, where the amount of uncured monomers is higher. The removal of this layer reduces the risk of exposure for patients. Rueggeberg al. established that the use of safety "cup" with a cotton swab placed between it and the tooth and slow speed polishing are effective for removal of excess uncured monomers (Rueggeberg et al., 1999). Rinsing the mouth with water for 30 seconds after placement of restorations also reduces the levels of chemical agents near to the baseline. To reduce further the exposure, a rubber dam can be used (Fleisch et al., 2010). The implementation of good practices in dentistry adds to reduce exposure of the patients and the dentist. Below (Table 1) are presented data from studies on the levels of BPA in some dental materials and biological environment.

Biomonitoring

Since BPA is a chemically unstable compound, with several hours half-life, its concentrations in blood are lower than those in urine, and decrease rapidly after the end of exposure (Needham and Sexton, 2000). Therefore, BPA cannot be detected in most blood samples with current analytical methods (WHO, 2010). Furthermore, due to its ubiquitous presence in the environment is difficult to ignore a possible contamination of a sample with minimal amounts free BPA during its storage and analysis (Markham *et al.*, 2010; Yonekubo *et al.*, 2008 and Zhang *et al.*, 2011). In such cases,

the established concentrations do not necessarily mean real exposure. BPA is rapidly and almost completely eliminated as bisphenol-conjugates, urine examination is the best choice for the aims of biomonitoring. The concentration of total (free plus conjugated) BPA in urine is often used to evaluate the exposure from all possible sources (Vandenberg *et al.*, 2007).

Table 1. Level of BPA in dental materials and biological environments

Dental material	Data for the release of BPA
Composite restorations	Saliva
	0.43 ng/mL (before restoration);
	0.64 ng/mL (1hour after restoration);
	0.4 ng/mL (1-30 hours after restoration).
	Urine
	1.67 ng/mL (before restoration);
T. 1 (2.38 ng/mL (9-30 hours after restoration).
Fissure sealant	Urine (data from 495 examined children):
composites	2.6/ µg/g (children with 11 and more treated
	Saliya (Delton I.C. Dentanly)
	3.08 ng/mL (3 hours after 1 restoration with
	sealer):
	9.08 ng/mI (3 hours after 4 restoration with
	sealer). Reaching baseline levels (0.07-6 ng /
	mL) within 24 hours
	Saliva (Delton LC):
	5.8-105.6 ppb (1 hour, 3 hours after
	placement)
	0.3-2.8 ppm (immediately after placement)
	Exposure to BPA (14 contestants)
	110 µg BPA (Delton LC, BisDMA-based
	sealer);
	5.5 µg BPA (Helioseal F, Ivoclar Vivadent)
	Saliva
	90-931 µg BPA (1 hour after placement)
Orthodontic bonding	Saliva
NI 1 (1 1 (0.8-20.88 ng/mL
Polycarbonate brackets	
	$38-60 \ \mu\text{g} / \text{g}$ material (18 months)
Lingual fixator	Saliva
Lingual fixator	20.9 ng/mI (30 minutes after placement)
Composite restoration	Silux Plus (3M)
composite restoration	6.4 µg/g in non-polymerized resin
	releases 91.4 ng/g material in phosphate
	buffered physiological solution(24 hours)
Fissure	Concise (3M)
sealant/Composites	15.4 μ g/g in non-polymerized resin; releases
•	19.8 ng/g material in phosphate buffered
	physiological solution(24 hours)
	Teeth Mate A (Kuraray)
	20.2 µg /g in non-polymerized resin; releases
	освобождава 55.5 ng/g material in phosphate
	buffered physiological solution(24 hours)
Dental bonding	Clearfil Photo Bond (Kuraray)
Orthe dentie heredine	18.5 μg/g in non-polymerized resin
Orthodontic bonding	the detection limit 0.1 ppm (in
Polycarbonate bracers	$697 \mu g/g$ water (40 months in water)
1 Orycarbonate bracers	$37 \mu \text{g} / \text{g}$ material (40 months in water)
	$0.01-0.4 \mu g / g$ Material (34 month in water)
Polycarbonate	2.2 µg/g (34 months in water)
prosthetic plates	
Polycarbonate	2.8 μ g/g (34 months in water)
temporary crowns	
Bonding with a lingual	Transbond XT (3M ESPE)
fixator	2.9 µg/mL (1 month in water)

IIo: Chen L, Suh BI (2013) Bisphenol A in Dental Materials: A Review. JSM Dent 1: 1004.

Studies conducted in North America, Europe and Asia indicated worldwide exposure to BPA. Two recent large-scale

studies on exposure degree to BPA were conducted in the US and Canada, including respectively 2514 and 5476 participants. Ubiquitous exposure to BPA, in more than 90% of participants in both studies was established (Bushnik et al., 2010 and Calafat et al., 2008). In USA, highest concentrations in urine were detected in adolescents (12-19 years), followed by children (6-11 years) and adults (>19 years). In the Canadian study (Bushnik et al., 2010), higher were the levels of BPA in the youngest group (6-11 years) if compared with other groups defined by age. In a study performed in Germany, higher concentrations were established in children 3-5 years old if compared with other groups (6-8 years, 9-11 years and 12-14 years old children) (Becker et al., 2009). In a study conducted in seven Asian countries, BPA was detected in 94% of samples (Zhang et al., 2011). Practically, to perform biomonitoring studies single urine samples are used. Due to the short half-life of BPA, these samples reflect primarily the exposure over a relatively short period of time, prior to the analysis of the sample (Koch and Calafat, 2009). However, in large-scale studies this approach may be useful to evaluate the average exposure of population. Data on urinary levels indicate average range of exposure 0.01-0.05 µg/kg body weight per day for adults, and slightly higher (0.02-0.12 µg/kg body weight per day) for children. For the 95th percentile, exposure assessment is evaluated to be 0.27 µg/kg body weight per day for population, and to be higher for babies $(0.45-1.61\mu g/kg)$ body weight per day) and children 3-5-years old (0.78 $\mu g/kg$ body weight per day) (Zhang et al., 2011).

Adverse health effects -data from epidemiological studies

Most of the conducted epidemiological studies are crosssectional, with a single evaluation of BPA in urine. These types of studies are limited in the field of interpretation, especially for effects with long latency period (e.g. cardiovascular diseases, diabetes). Data on relationship between BPA exposure and manifestation of adverse health effects such as cancer, reproductive damage, cardiovascular disease, diabetes, and disorders of growth, pubertal and neuropsychological development are summarized in the report of the Joint FAO/WHO Expert Meeting (WHO, 2010). Bisphenol A belongs to the group of xenoestrogens - substances with estrogen-mimicking properties. A significant number of studies have confirmed the estrogenic potential of BPA (Chapin et al., 2008) and define it as endocrine disruptor chemical, because of its ability to bind to and activate estrogen receptors, although with a capacity of 1000-5000 times smaller than endogenous 17β-estradiol (Roy et al., 2009). In addition, BPA may react with alternative endocrine receptors such as those for thyroid hormone, and peroxisome proliferatoractivated receptor gamma (Diamanti-Kandarakis et al., 2009). BPA is classified in category 3 for reproductive toxicity, and is considered as main fertility risk factor in humans (INSERM, 2010).

Epidemiological studies demonstrated a correlation between increased urinary concentrations of BPA and poor sperm quality. There is no evidence of a correlation between urinary BPA levels and the age of puberty onset among girls. In a prospective study performed by Braun *et al.* (2009) was suggested that prenatal exposures, especially in the period of early pregnancy, may be associated with developmental and behavioral disorders - aggression and hyperactivity, especially among girls. Reproduction of this experiment is needed, with multiple examinations of urinary levels of BPA. Basing on data concerning urine its concentration, an association between exposure to BPA and development of cardiovascular diseases (Lang et al., 2008; Melzer et al., 2012 and Melzer et al., 2010), diabetes (Alonso-Magdalena et al., 2005; Alonso-Magdalena et al., 2006; Alonso-Magdalena et al., 2011 and Alonso-Magdalena et al., 2010), obesity (Bhandari et al., 2013), chronic kidney disease (Krieter et al., 2013), breast and uterus cancer (Hiroi et al., 2004 and Smith-Bindman, 2012), immune disorders (Clayton et al., 2011), chronic respiratory disease and asthma (Spanier et al., 2012) was found. Although the short, less than six hours half-life of BPA, recent data suggest that the substance can be accumulated in adipose tissue (Stahlhut et al., 2009).

Confirmation of observations above is needed, with prospective studies performing numerous measurements of BPA levels of in order to clarify the duration of exposure (years or even decades) before the onset of cardiovascular diseases, diabetes and reproductive disorders. Due to the short half-life of BPA, although ubiquitously presenting, it is unsecure if epidemiological studies in humans will make it possible to establish the correlation between BPA exposure and long-term effects manifestation (Völkel *et al.*, 2005 and Welshons *et al.*, 2006). Prenatal exposure to BPA may have harmful cumulative effects; the National Toxicology Program and the US Food and Drug Association Risk Assessment state that "BPA exposure may lead to alterations in nervous system development, in reproduction and metabolism throughout life" (Chapin *et al.*, 2008).

Conclusions

- 1. Bisphenol A is xenoestrogen synthesized in large quantities worldwide for production of polymers (polycarbonates, epoxy resins) and thermal paper. This determines its universal presence - in everyday products (packaging, containers and bottles), food and drinking water. Food is considered to be the most important source of population exposure; however, in overall exposure assessment consumption of drinking water, inhalation of dust and dermal contact with thermal paper must be taken into account.
- 2. BPA can cause numerous adverse health effects, disrupting endocrine system (e.g. affecting sex hormones, insulin, leptin, adiponectin and thyroxine), as well as immune and nervous systems. Hepatotoxic, mutagenic, and carcinogenic effects and increased risk of coronary diseases have been also discussed.
- 3. Exposure to BPA and its derivatives from dental composites and sealants is possible. High levels of BPA in saliva (especially immediately or one hour after dental treatment), decreasing over time have been found. No BPA in the blood samples of dental patients have been detected, as reported in the available studies. High urinary levels of BPA after treatment with dental composites and sealants have been reported.

4. The degree of exposure to BPA from dental materials and the possible adverse health effects are insufficiently investigated. No data were found in the available literature concerning the urinary levels of BPA among occupationally exposed dental professionals, in comparison with those among dental patients after treatment with composites and sealants.

REFERENCES

- Alonso-Magdalena, P., Laribi, O., Ropero, A.B., Fuentes, E., Ripoll, C., Soria, B., *et al.* 2005. Low doses of bisphenol A and diethylstilbestrol impair Ca2+ signals in pancreatic alpha-cells through a nonclassical membrane estrogen receptor within intact islets of Langerhans. *Environ Health Perspect*, 113:969–77
- Alonso-Magdalena, P., Morimoto. S., Ripoll, C., Fuentes, E., Nadal, A. 2006. The estrogenic effect of bisphenol A disrupts pancreatic beta-cell function in vivo and induces insulin resistance. *Environ Health Perspect*, 114:106–12.
- Alonso-Magdalena, P., Quesada, I., Nadal, A. 2011. Endocrine disruptors in the etiology of type 2 diabetes mellitus. *Nat Rev Endocrinol*, 7:346–53.
- Alonso-Magdalena, P., Ropero, A.B., Soriano, S., Quesada, I., Nadal, A. 2010. Bisphenol-A: a newdiabetogenic factor? Hormones (Athens), 9:118–26.
- American Dental Association Council on Scientific Affairs. ADA Professional Product Review. http://www.ada.org/271.aspx
- Arnold, S., Clark, K., Stapes, Ch., Klecka, G., Dimond, S., Caspers, N., Hentges, S. 2013. Relevance of drinking water as a source of human exposure to bisphenol A. J. Exp. Sci. Environ. Epidemiol, 23, 137–144.
- Azarpazhooh, A., Main, P.A. 2008. Is there a risk of harm or toxicity in the placement of pit and fissure sealant materials? A systematic review. J Can Dent Assoc, 74(2):179-183.
- Bakopoulou, A., Papadopoulos, T., Garefis, P. 2009. Molecular toxicology of substances released from resinbased dental restorative materials. *Int J Mol Sci*, 10(9):3861-3899.
- Becker, K., Goen, T., Seiwert, M., Conrad, A., Pick-Fub, H., Moller, J., Wittassek, M., Schulz, C., Kolossa-Gehring, M., 2009. GerES IV: phthalate metabolites and bisphenol A in urine of German children. *Int. J. Hyg. Environ. Health*, 212, 685–692.
- Berkner, S., Streck, G., Hermann, R. 2004. Development andvalidation of a method for determination of trace levels of alkylphenol and bisphenol A in atmospheric samples. *Chemosphere*, 54, 575–584.
- Bhandari, R., Xiao, J., Shankar, A. 2013. Urinary Bisphenol A and Obesity in US Children. *Am J Epidemiol*, 177:1263– 70.
- Braun, J.M., Kalkbrenner, A.E., Calafat, A.M., Bernert, J.T., Ye, X., Silva, M.J., Barr, D.B., Sathyanarayana, S., Lanphear, B.P. 2011. Variability and predictors of urinary bisphenol A concentrations during pregnancy. *Environ. Health Perspect*, 119, 131–137
- Braun, J.M., Yolton, K., Dietrich, K.N., Hornung, R., Ye, X.Y., Calafat, A.M., Lanphear, B.P. 2009. Prenatal

bisphenol A exposure and early childhood behaviour. Environ. *Health Perspect*, 117, 1945–1952.

- Bushnik, T., Haines, D., Levallois, P., Levesque, J., 2010. Lead and bisphenol A concentrations in the Canadian populations. *Stat. Can. Health Rep*, 21, 7–18.
- Calafat, A.M., Weuve, J., Ye, X., Jia, L.T., Hu, H., Ringer, S., Huttner, K., Hauser, R. 2009. Exposure to bisphenol A and other phenols in neonatal intensive care unit premature infants. *Environ. Health Perspect*, 117, 639–644.
- Calafat, A.M., Ye, X.Y., Wong, L.Y., Reidy, J.A., Needham, L.L. 2008. Exposure of the US population to bisphenol A and 4-tertiary-octylphenol: 2003–2004. *Environ. Health Perspect*, 116, 39–44.
- Cao, X., Corriveau, J., Popovic, S. 2009. Levels of bisphenol A incanned soft drink products in canadian markets. J. Agric. Food Chem, 57, 1307–1311.
- Chapin, R.E., Adams, J., Boekelheide, K., Gray, L.E. Jr., Hayward, S.W., Lees, P.S., McIntyre, B.S., Portier, K.M., Schnorr, T.M., Selevan, S.G., Vandenbergh, J.G., Woskie, S.R. 2008. NTP-CERHR expert panel report on the reproductive and developmental toxicity of bisphenol A. *Birth Defects Res B Dev Reprod Toxicol*, 83(3):157-395.
- Cilli, R., Pereira, J.C., Prakki, A. 2012. Properties of dental resins submitted to pH catalysed hydrolysis. *Journal of Dentistry*, 40:1144–50.
- Cladiere, M., Gasperi, J., Lorgeoux, C., Bonhomme, C., Rocher, V., Tassin, B. 2013. Alkylphenolic compounds and bisphenol A contamination within heavily urbanized area: case study of Paris. *Environ. Sci. Pollut. Res*, 20, 2973– 2983.
- Clayton, E.M., Todd, M., Dowd, J.B., Aiello, A.E. 2011. The impact of bisphenol A and triclosan on immune parameters in the U.S. population, NHANES 2003–2006. *Environ Health Perspect*, 119:390–6.
- Colin, A., Bach, C., Rosin, Ch., Munoz, JF., Dauchy, X. 2013. Is drinking water a major route of human exposure to alkylphenol and bisphenol contaminants in France? Arch. Environ. Contam. Toxicol, http://dx.doi.org/10.1007/ s00244-013-9942-0.
- Cunha, S., Cunha, C., Ferreira, A., Fernandes, J. 2012. Determination of bisphenol A and bisphenol B in canned seafood combining QuEChERS extraction with dispersive liquid-liquid microextraction followed by gas chromatography-mass spectrometry. *Anal. Bioanal. Chem*, 404, 2453–2463.
- Cunha, S., Fernandes, J. 2013. Assessement of bisphenol A andbisphenol B in canned vegetables by gaschromatography-mass spectrometry after QuEChERS and dispersive liquid-liquid microextraction. *Food Control*, 33,549–555.
- Danzl, E., Sei, K., Soda, S., Ike, M., Fujita, M. 2009. Biodegradation of bisphenol A, bisphenol F and bisphenol S in seawater. *Int. J. Environ. Res. Pub. Health*, 6, 1472– 1484.
- Dekant, W., Völkel, W. 2008. Human exposure to bisphenol A by biomonitoring: methods, results and assessment of environmental exposures. *Toxicol. Appl. Pharmacol*, 228, 114–134.
- Diamanti-Kandarakis, E., Bourguignon, J.P., Giudice, L.C., Hauser, R., Prins, G.S., Soto, A.M., et al. 2009. Endocrine-

disrupting chemicals: an endocrine society scientific statement. *Endocrine Reviews*, 30:293–342.

- Doerge, D.R., Twaddle, N.C., Vankandingham, M., Fisher, J.W. 2010. Pharmacokinetics of bisphenol-A in neonatal and adult Sprague-Dawley rats. *Toxicol. Appl. Pharmacol*, 247, 158–165.
- Doerge, D.R., Twaddle, N.C., Woodling, K.A., Fisher, J.W. 2010. Pharmacokinetics of bisphenol A in neonatal and adult rhesus monkeys. *Toxicol. Appl. Pharmacol*, 248, 1– 11.
- EFSA European Food Safety Authority. Opinion of the Scientific Panel on Food Additives, Flavourings, Processing Aids and Materials in Contact with Food on a request from the Commission related to 2,2-bis(4hydroxyphenyl) propane (Bisphenol A). Question number EFSA-Q-2005-100. 2006. *The EFSA Journal*, 428:1–75. Available from: http://www.efsa.europa.eu/fr/scdocs/scdoc/428.htm.
- FDA, Food and Drug Administration. 2009. Safety assessment of BPA in medical products. August 7, 2009. Available from: http://www.fda.gov/downloads/Advisory Committees/ CommitteesMeetingMaterials/ScienceBoardto theFoodand DrugAdministration/UCM176835.pdf.
- Fleisch, A.F., Sheffield, P.E., Chinn, C., Edelstein, B.L., Landrigan, P.J. 2010. Bisphenol A and related compounds in dental materials. *Pediatrics*, 126(4):760-8.
- Flint, S., Markle, T., Thomson, S., Wallace, E., 2012. Bisphenol Aexposure effects and policy; a wildlife perspective. J. Environ.Manag., 104, 19–34.
- Fu, P., Kawamura, K. 2010. Ubiquity of bisphenol A in theatmosphere. *Environ. Pollut*, 158, 3138–3143.
- Fung, E.Y.K., Ewoldsen, N.O., St Germain, H.A., Marx, D.B., Miaw, C.L., Siew, C., Chou, H.N., Gruninger, S.E., Meyer, D.M. 2000. Pharmacokinetics of bisphenol A released from a rental sealant. J. Am. Dent. Assoc, 131, 51–58.
- Geens, T., Goeyens, L., Covaci, A. 2011. Are potential sources for human exposure to bisphenol-A overlooked? *Int. J. Hyg. Environ. Health*, 214, 339–347.
- Geens, T., Roosens, L., Neels, H., Covaci, A. 2009. Assessment of human exposure to bisphenol-A, triclosan and tetrabromobisphenol-A through indoor dust intake in Belgium. *Chemosphere*, 76, 755–760.
- Gerzina, T.M., Hume, W.R. 1996. Diffusion of monomers from bonding resin-resin composite combinations through dentine in vitro. *J Dent*, 24(1-2):125-128.
- Hiroi, H., Tsutsumi, O., Takeuchi, T., Momoeda, M., Ikezuki, Y., Okamura, A., *et al.* 2004. Differences in serum bisphenol a concentrations in premenopausal normal women and women with endometrial hyperplasia. *Endocr* J, 51:595–600.
- Imai, Y., Komabayashi, T. 2000. Elution of Bisphenol A from Composite Resin: A Model Experiment. *Dental Materials Journal*, 19 (2):133-138.
- Imai, Y. 2000. Comments on "Determination of Bisphenol A and Related Aromatic Compounds Released from Bis-GMA-Based Composites and Sealants by High Performance Liquid Chromatography". *Environmental Health Perspectives*, 108 (12), 545.
- INSERM–Institute of National Health and Medical Research. 2010. Bisphenol A: Effects on reproduction. Preliminary report. Paris, INSERM.

- Jones-Otazo, H.A., Clarke, J.P., Diamond, M.L., Archbold, J.A., Ferguson, G., Harner, T., Richardson, G.M., Ryan, J.J., Wilford, B. 2005. Is house dust the missing exposure pathway for PBDEs? An analysis of the urban fate and human exposure to PBDEs. *Environ. Sci. Technol*, 39, 5121–5130.
- Joskow, R., Barr, D.B., Barr, J.R., Calafat, A.M., Needham, L.L., Rubin, C. 2006. Exposure to bisphenol A from bisglycidyl dimethacrylate-based dental sealants. JADA, 137(3):353–362.
- Kawagoshi., Y., Fujita, Y., Kishi, I., Fukunaga, I., 2003. Estrogenic chemicals and estrogenic activity in leachate from municipal waste landfill determined by yeast twohybrid assay. *J.Environ. Monit.*, 5, 269–274.
- Kingman, A., Hyman, J., Masten, S.A., Jayaram, B., Smith, C., Eichmiller, F., Arnold, M.C., Wong, P.A., Schaeffer, J.M., Solanki, S., Dunn, W.J. 2012. Bisphenol A and other compounds in human saliva and urine associated with the placement of composite restorations. JADA, 143(12):1292-1302.
- Koch, H.M., Calafat, A.M., 2009. Human body burdens of chemicals used in plastic manufacture. *Philos. Trans. R. Soc. B Biol. Sci*, 364, 2063–2078.
- Krieter, D.H., Canaud, B., Lemke, H.D., Rodriguez, A., Morgenroth, A., von Appen, K., *et al.* 2013. Bisphenol A in chronic kidney disease. Artif Organs, 37:283–90.
- Lang, I.A., Galloway, T.S., Scarlett, A., Henley, W.E., Depledge, M., Wallace, R.B., Melzer, D. 2008. Association of urinary bisphenol A concentration with medical disorders and laboratory abnormalities in adults. *JAMA*, 300, 1303–1310.
- Lassen, C., Mikkelsen, S.H., Brandt, U.K. 2011. Migration of bisphenol A from cash register receipts and baby dummies. In: Survey of Chemical Substances in Consumer Products. *Danish Ministry of the Environment*, No. 110.
- Lee, Ch-Ch., Jiang, L-Y., Kuo, Y-L., Hsieh Ch, Y., Chen, C., Tien Ch,J., 2013. The potential role of water quality parameters on occurrence of nonylphenol and bisphenol A and identification of their discharge sources in the river ecosystems. *Chemosphere*, 91, 904–911.
- Lee, H., Peart, T. 2000. Bisphenol A contamination in Canadian municipal and industrial wastewater and sludge samples. *Water Q. Res. J. Can.* 35, 283–298.
- Liao, C., Kannan, K. 2011. Widespread occurrence of bisphenol A in paper and paper products: implications for human exposure. *Environ. Sci. Technol*, 45, 9372–9379.
- Loganathan, S., Kannan, K. 2011. Occurrence of bisphenol A in indoor dust from two locations in the Eastern United States and implications for human exposures. *Arch. Environ.Contam. Toxicol*, 61, 68–73.
- Markham, D.A., Waechter Jr., J.M., Wimber, M., Rao, N., Connolly, P., Chuang, J.C., Hentges, S., Shiotsuka, R.N., Dimond, S., Chappelle, A.H., 2010. Development of a method for the determination of bisphenol-A at trace concentrations in human blood and urine and elucidation of factors influencing method accuracy and sensitivity. J. Anal. Toxicol, 34, 293–303.
- Martin, M.D., Bajet, D., Woods, J.S., Dills, R.L., Poulten, E.J. 2005. Detection of dental composite and sealant resin components in urine. Oral Surgery Oral Medicine Oral Pathology Oral Radiology and Endodontics, 99:429.

- Matsumoto, H., Adachi, S., Suzuki, Y. 2005. Bisphenol A inambient air particulates responsible for the proliferation of MCF-7 human breast cancer cells and its concentration changes over 6 months. *Arch. Environ. Contam. Toxicol*, 48,459–466.
- Matthews, J.B., Twomey, K., Zacharewski, T.R. 2001. In vitro and in vivo interactions of bisphenol A and its metabolite, bisphenol A glucuronide, with estrogen receptors alpha and beta. *Chem. Res. Toxicol*, 14, 149–157.
- Melzer, D., Osborne, N.J., Henley, W.E., Cipelli, R., Young, A., Money, C., McCormack, P., Luben, R., Khaw, K.T., Wareham, N.J., Galloway, T.S. 2012. Urinary bisphenol-A concentration and risk of future coronary artery disease in apparently healthy men and women. *Circulation*, 125,12:1482-90.
- Melzer, D., Rice, N.E., Lewis, C., Henley, W.E., Galloway, T.S. 2010. Association of urinary bisphenol A concentration with heart disease: evidence from NHANES 2003/06. PLoS ONE 5, e8673.
- Mercea, P. 2009. Physicochemical processes involved inmigration of bisphenol A from polycarbonate. J. Appl. Polym.Sci, 112, 579–593.
- Mohsen, N.M., Craig, R.G., Hanks, C.T. 1998. Cytotoxicity of urethane dimethacrylate composites before and after aging and leaching. *Journal of Biomedical Materials Research*, 39:252–60.
- Munguia-Lopez, E., Gerardo-Lugo, S., Peralta, E., Bolumen, S.,Soto-Valdez, H. 2005. Migration of bisphenol A (BPA) from cancoatings into a fatty-food simulant and tuna fish. *Food Addit.Contam.*, 22, 892–898.
- Myers, D.E., Hutz, R.J. 2011. Current status of potential bisphenol toxicity in dentistry. [Review] *General Dentistry*, 59(4):262-5.
- Nam, S.H., Seo, Y.M., Kim, M.G. 2010. Bisphenol A migration from polycarbonate baby bottle with repeated use. *Chemosphere*, 79, 949–952.
- Nathanson, D., Lertpitayakun, P., Lamkin, M.S., Edalatpour, M., Chou, L.L. 1997. In vitro elution of leachable components from dental sealants. J. Am. Dent. Assoc, 128, 1517–1523.
- Needham, L.L., Sexton, K., 2000. Assessing children's exposure to hazardous environmental chemicals: an overview of selected research challenges and complexities Introduction and overview. J. Exp. Anal. Environ. Epidemiol., 10, 611–629.
- Niu, Y., Zhang, J., Wu, Y., Shao, B. 2012. Analysis of bisphenol A and alkylphenols in cereals by automated online solid-phase extraction and liquid chromatography tandem massspectrometry. J. Agric. Food Chem., 60, 6116–6122.
- O'Mahony, J., Moloney, M., McCormack, M., Nicholls, I., Mizaikoff,B., Danaher, M. 2013. Design and implementation of an imprinted material for the extraction of the endocrine disruptor bisphenol A from milk. J. *Chromatogr*, 931,164–169.
- Olea, N., Pulgar, R., Pérez, P., et al. 1996. Estrogenicity of resin-based composites and sealants used in dentistry. *Environ Health Perspect*, 104(3):298–305.
- Plastics Europe. 2007. Applications of Bisphenol A. Available from: http://www.bisphenol-a-europe.org/uploads/BPA% applications.Pfd>.

- Polydorou, O., Konig, A., Hellwig, E., Kummerer, K. 2009. Long-term release of monomers from modern dentalcomposite materials. *Eur J Oral Sci.*, 117(1):68-75.
- Pulgar, R., Olea-Serrano, M. F., Novillo-Fertrell, A., Rivas, A., Pazos, P., Pedraza, V., Navajas, J. M., Olea, N. 2000. Determination of bisphenol A and related aromatic compounds released from BisDMA-based composites and sealants by high performance liquid chromatography. *Environ Health perspect*, 108, 21-27.
- Reichl, F.X., Seiss, M., Kleinsasser, N., Kehe, K., Kunzelmann, K.H., Thomas, P., Spahl, W., Hickel, R. 2008. Distribution and excretion of BisGMA in guinea pigs. *J Dent Res*, 87(4):378-380.
- Rocha, S., Domingues, V., Pinho, C., Fernandes, V., Delerue-Matos, C., Gameiro, P., Mansilha, C. 2013. Occurrence of bisphenol A, estrone, 17β-estradiol and 17αethinylestradiol in Portugalese Rivers. *Bull. Environ. Contam. Toxicol*, 90, 73–78.
- Rogalewicz, R., Voelkel, A., Kownacki, I. 2006. Application of HS-SPME in the determination of potentially toxic organic compounds emitted from resinbased dental materials. *J Environ Monit*, 8(3):377-383.
- Roy, J.R., Chakraborty, S., Chakraborty, T.R. 2009. Estrogenlike endocrine disrupting chemicals affecting puberty in humans – a review. *Medical Science Monitor*, 15:137–45.
- Rueggeberg, F.A., Dlugokinski, M., Ergle, J.W. 1999. Minimizing patients' exposure to uncured components in a dental sealant. J Am Dent Assoc, 130(12):1751-1757.
- Sasaki, N., Okuda, K., Kato, T., et al. 2005. Salivary bisphenol-A levels detected by ELISA after restoration with composite resin. J Mater Sci Mater Med,16(4):297– 300.
- Schmalz, G., Preiss, A., Arenholt-Bindslev, D. 1999. Bisphenol-A content of resin monomers and related degradation products. *Clin Oral Investig*, 3(3):114-9.
- Schmalz, G., Arenholt-Bindslev, D. 2009. Biocompatibility of dental materials. Springer, Berlin, Heidelberg.
- Shao, B., Han, H., Li, D., Ma, Y., Tu, X., Wu, Y. 2007. Analysis of alkylphenol and bisphenol A in meat by accelerated solvent extraction and liquid chromatography with tandem massspectrometry. *Food Chem*, 105, 1236– 1241.
- Smith-Bindman, R. 2012. Environmental causes of breast cancer and radiation from medical imaging: findings from the Institute of Medicine report. *Arch Intern Med*, 172:1023–7.
- Snyder, R.W., Maness, S.C., Gaido, K.W., Welsch, F., Summer, S.C.J., Fennell, T.R. 2000. Metabolism and disposition of bisphenol A in female rats. *Toxicol. Appl. Pharmacol*, 168, 225–234.
- Spanier, A.J., Kahn, R.S., Kunselman, A.R., Hornung, R., Xu, Y., Calafat, A.M., *et al.* 2012. Prenatal exposure to bisphenol A and child wheeze from birth to 3 years of age. *Environ Health Perspect*, 120:916–20.
- Stachel, B., Ehrhorn, U., Heemken, O., Lepom, P., Reinckle, H., Sawal, G., Theobald, N. 2003. Xenoestrogens in the River Elbeand its tributaries. *Environ. Pollut*, 124,497–504.
- Stahlhut, R.W., Welshons, W.V., Swan, S.H. 2009. Bisphenol A data in NHANES suggest longer than expected half-life, substantial nonfood exposure, or both. *Environ Health Perspect*, 117(5):784-789.

- Takahashi, Y., Shirai, A., Segawa, T., Takahashi, T., Sakakibara, K. 2002. Why does a color-developing phenomen occur on thermal paper comprising of a fluoran dye and a color developer molecule? *Bull. Chem. Soc. Jpn*, 75, 2225–2231.
- Tarumi, H., Imazato, S., Narimatsu, M., Matsuo, M., Ebisu, S. 2000. Estrogenicity of fissure sealants and adhesive resins determined by reporter gene assay. *J Dent Res*, 79(11):1838-43.
- Van Landuyt, K.L., Yoshihara, K., Geebelen, B., Peumans, M., Godderis, L., Hoet, P., Van Meerbeek, B. 2012. Should we be concerned about composite (nano-)dust? *Dental Materials*, 28, 1162-117015.
- Van Landuyt, K.L., Nawrot, T., Geebelen, B., De Munck, J., Snauwaert, J., Yoshihara, K., Scheers, H., Godderis, L., Hoet, P., Van Meerbeek, B., 2011. How much do resinbased dental materials release? A meta-analytical approach. Dent. Mater, 27, 723–747
- Vandenberg, L.N., Chahoud, I., Heindel, J.J., Padmanabhan, V., Paumgartten, F.J., Schoenfelder, G. 2010. Urinary, circulating, and tissue biomonitoring studies indicate widespread exposure to bisphenol A. *Environ Health Perspect*, 118(8):1055-1070.
- Vandenberg, L., Hauser, R., Marcus, M., Olea, N., Welshons, W. 2007. Human exposure to bisphenol A (BPA). Reprod. Toxicol, 24, 139–177.
- Volkel, W., Colnot, T., Csanady, G.A., Filser, J.G., Dekant, W. 2002. Metabolism and kinetics of bisphenol A in humans at low doses following oral administration. *Chem Res Toxicol.*, 15(10):1281-1287.
- Völkel, W., Bittner, N., Dekant, W. 2005. Quantitation of bisphenol A and bisphenol A glucuronide in biological samples by high performance liquid chromatography– tandem mass spectrometry. *Drug Metab. Dispos*, 33, 1748– 1757.
- Vőlkel, W., Kiranoglu, M., Fromme, H., 2008. Determination of free and total bisphenol A in human urine to assess daily uptake as a basis for a valid risk assessment. *Toxicol. Lett.* 179, 155–162.
- von Goetz, N., Wormuth, M., Scheringer, M., Hüngerbuhler, K. 2010. Bisphenol A: how the most relevant exposure sources contribute to total consumer exposure. Risk. Anal, 30, 473–487.
- Welshons, W.V., Nagel, S.C., vom Saal, F.S.V. 2006. Large effects from small exposures III – Endocrine mechanisms mediating effects of bisphenol A at levels of human exposure. *Endocrinology*, 147, S56–S59.
- WHO 2010 Joint FAO/WHO Expert Meeting to Review Toxicological and Health Aspects of Bisphenol A. Summary report. Available from: http://www.who.int/foodsafety/chem/chemicals/bisphenol release/en/index.html>.
- Yonekubo, J., Hayakawa, K., Sajiki, J. 2008. Concentrations ofbisphenol A, bisphenol A diglycidyl ether, and their derivatives in canned foods in Japanese markets. J. Agric. FoodChem., 56, 2041–2047.
- Yoshida, T., Horie, M., Hoshino, Y., Nakazawa, H. 2001. Determination of bisphenol A in canned vegetables and fruitby high performance liquid chromatography. *Food Addit. Contam.*, 18, 69–75.

- Zalko, D., Jacques, C., Duplan, H., Bruel, S., Perdu, E. 2011. Viable skin efficiently absorbs and metabolizes bisphenol A. Chemosphere, 82, 424–430.
- Zhang, Z., Alomirah, H., Cho, H.S., Li, Y.F., Liao, C., Minh, T.B., Mohd, M.A., Nakata, H., Ren, N., Kannan, K., 2011. Urinary bisphenol A concentrations and their implications for human exposure in several Asian countries. *Environ. Sci. Technol*, 45, 7044–7050.
- Zincke, T., 1905, "Mittheilungen AUS DEM chemischen LABORATORIUM Universitat Marburg," *Justus Leibigs Annals Chemie*, 343, 75-99.
