



ISSN: 0975-833X

RESEARCH ARTICLE

ANTIBACTERIAL ACTIVITY OF VARIOUS EXTRACTS OF LEAF AND WOOD OF *Anisomeles malabarica*

Suresh Subramanian and *Suriyavathana Muthukrishnan

Department of Biochemistry, Periyar University, Salem, Tamil Nadu, 636 011, India

ARTICLE INFO

Article History:

Received 27th December, 2012
Received in revised form
16th January, 2013
Accepted 19th February, 2013
Published online 19th March, 2013

Key words:

Antibiotic, *Anisomeles malabarica* (L),
E.Coli, Antiallergic.

ABSTRACT

Water is essential to life, but many people do not have access to clean and safe drinking water and many die of waterborne bacterial infections. Antibacterial properties of various extracts of *Anisomeles malabarica* (L) were investigated against both clinical and laboratory isolates of bacteria using the disc diffusion method. Aqueous extracts (10 mm zone diameter of inhibition, MIC 10µg/mL) demonstrated the highest activity, followed by Ethanol extract (8 mm zone diameter of inhibition, MIC 20µg/mL) methanolic extract demonstrated the least activity against the test bacteria (4 mm zone diameter of inhibition, MIC 40 µg/mL). Generally the antibacterial activities of *Anisomeles malabarica* are comparable to that of standard ofloxacin to a certain extent. Finally, these results suggested that *Anisomeles malabarica* (L) can be used to source antibiotic substances for possible treatment of bacterial infections and for the development of new drugs.

Copyright, IJCR, 2013, Academic Journals. All rights reserved.

INTRODUCTION

Natural products from microorganisms have been the primary source of antibiotics, but with the increasing acceptance of herbal medicine as an alternative form of healthcare, the screening of medicinal plants for active compounds has become very important because these may serve as promising sources of novel antibiotic prototypes Koduru *et al.*, 2006. Many efforts have been made to discover new antimicrobial compounds from various kinds of sources such as micro-organisms, animals, and plants. One of such resources is folk medicines. Systematic screening of them may result in the discovery of novel effective compounds Tomoko *et al.*, 2002. The side effects and the resistance that pathogenic microorganisms build against antibiotics, much recent attention has been paid to extracts and biologically active compounds isolated from plant species used in herbal medicine Essawi and Srour 2000. Further to that, natural extracts are normally perceived as "safer" and less harmful, if compared with artificial products; all these reasons caused an increase in their demand Bianco and Uccella 2000. Wood is composed of three main macromolecular components; cellulose, hemicellulose and lignin, which are present in all woods Fengel and Wegener 1984. The relative proportion of these components is variable within tree species and can range for cellulose between 28 and 50 %; hemicelluloses, 20 and 30 %; and for lignin, 18 and 30 % relative mass fraction Fengel and Wegener 1984. Significant variations in the concentration of these components can also be found within the same tree species Kenney, Sennerby-Forsseand Layton, 1990. The heartwood of trees has long been known to produce and accumulate biologically active compounds from a variety of chemical classes Rennerfelt and Nacht, 1955; Rudman, 1963; Scheffer and Cowling, 1966 that protect them from microbial decay. Alaska yellow-cedar (*Callitropsis nootkatensis*; previously *Chamaecyparis nootkatensis*; Little, 2007; Little and Schwarzbach, 2004 heartwood is an excellent example. In recent years multiple drug resistance in human pathogenic microorganisms has developed due to indiscriminate use of commercial antimicrobial drugs commonly used in the treatment of diseases. This situation forced the scientists

for searching new antimicrobial substances from various sources of novel antimicrobial chemotherapeutic agent. *Anisomeles malabarica* R.Br., (Lamiaceae) is distributed in major parts of India and especially in South India as a traditional medicinal plant commonly known as Peymarutti (Tamil), Gouzaban (Hindi), Chodhara (Marathi), Karithumbi (Kannada) and Malabar catmint (English). It is used for the traditional treatment of snakebite as Antidote Perumalsamy *et al.*, 2008. The herb is reported to the herb is reported to possess anticancer, allergenic, anthelmintic, antiallergic, antianaphylactic, antibacterial, anticarcinomic, antiedemic, antihistaminic, antiinflammatory, antileukemic, antinociceptive, antiplasmodial, antiseptic and antiperotic properties Jeyachandran *et al.*, 2007. The present work was set to investigate the antimicrobial activity of leaf and wood extracts of *Anisomeles malabarica* against some selected bacteria.

Experimental Section

Collection of sample

Anisomeles malabarica (L) were collected from kolli hills in Namakkal, District, Tamilnadu, India and were authenticated in Institute of Herbal Botany, Plant Anatomy Research Centre, Chennai. The leaf were separated from stem, both wood and leaf were washed in clean water, leaf were dried at room temperature Eloff, 1998. The dried plant leaf were milled to a fine powder, The wood was debarked and chipping was done with chipper. Both leaf powder and wood chip were stored in the dark at room temperature in closed containers until required.

Extraction

10g of powdered leaf and woodchips of *Anisomeles malabarica* extracted with 100ml of Ethanol, Aqueous and Methanol separately using soxhlet apparatus.

*Corresponding author: makkaisu@gmail.com

Culture media

The cultures were subcultured regularly (every 30 days) and stored at 40°C as well as 800°C by preparing suspensions in 10% glycerol.

Microorganisms

In this study *Escherichia coli*, *Salmonella typhi*, *Shigella dysenteriae*, *Campylobacter jejuni*, *Enterobacter aerogenes* were used.

Inoculum preparation

A loopful of isolated colonies was inoculated into 4 ml peptone water and incubated at 37°C for 4 h. The turbidity of actively growing bacterial suspension was adjusted to match the turbidity standard of 0.5 McFarland units prepared by mixing 0.5 ml of 1.75% (w/v) barium chloride dehydrate with 99.5 ml 1% (v/v) sulphuric acid. This turbidity was equivalent to approximately 1-2×10⁸ colony-forming units per milliliter (cfu/ml). This 2-h grown suspension was used for further testing.

RESULTS

The Results are represented in Table 1 2 3. Of the tested plant extract, aqueous extract of *Anisomeles malabarica* showed antibacterial activity against most of the bacterial species investigated including its wide spectrum of activity, while Ethanolic and Methanolic extract of *Anisomeles malabarica* showed less activity compared to that of Aqueous extract.

Manivann, 2010. The aqueous extract of wood shows lowest values (MIC 10 µg/ml), followed by ethanolic extract against *E. Coli* (MIC 20 µg/ml) and *Salmonella typhi* (MIC 30 µg/ml). Most of the MIC values were lower, it indicating that the extracts could be bactericidal in action and high efficacy. The active extracts are reported to play a role in the prevention of colonization by bacteria and fungi Chiang, 2003. Lower MIC values (Table 2) and higher zones of inhibition (Figure 1 and 2) for aqueous extracts connotes higher solubility of phytoconstituents in the aqueous compared to the other solvents used. Members of the bacterial genera *Actinomadura*, *Microbispora*, *Microtetraspora* and *Sphingobacterium* were also found in similar environments to wood chip piles Novinscak *et al.*, 2009; Adams and Frostick 2009 and were able to degrade hemicellulose compounds, eg. xylan, under aerobic conditions Holtz, 1991; Berens, 1996; Pankratov, 2007. Table 3 shows the effect of combination of extracts and antibiotic agents on the test organisms. Results revealed an increased activity of ofloxacin (30 µg/mL) in the presence of both leaf and wood extract (30 µg/mL). At 30 µg/mL ofloxacin extracts had no effect on *Enterobacter*, but when combined, there was a remarkable activity (7 mm zone diameter of inhibition). At 30 µg/mL the activity of the extract and ofloxacin against *Shigella dysenteriae* were 10 and 14 mm (zone diameter of inhibition), respectively but this increased to 19 mm when the extract and the antibiotics were combined. There has been much interest in recent years on compounds derived from plants and herbs for their medicinal properties or biological activities Al-dabbas *et al.*, 2006. Throughout human history, infectious diseases are known to have been treated with herbal remedies Ozturk and Ercisli, 2006.

Table 1. Antimicrobial screening of *Anisomeles malabarica* extract by pour plate method

| Species | Control | Leaf | Wood |
|-------------------------------|---------|------|------|
| <i>E. Coli</i> | + | - | - |
| <i>Salmonella typhi</i> | + | - | - |
| <i>Shigella dysenteriae</i> | + | - | - |
| <i>Campylobacter jejuni</i> | + | - | - |
| <i>Enterobacter aerogenes</i> | + | - | - |

Table 2. Minimum inhibitory concentration (MIC) of extracts of *Anisomeles malabarica* (L)

| Organisms | MIC (µg/ml) | | | | | | | |
|-------------------------------|-------------|----|----|-----|------|----|----|-----|
| | Leaf | | | | Wood | | | |
| | EE | AE | ME | Ofx | EE | AE | ME | Ofx |
| <i>E. Coli</i> | 20 | 30 | 50 | 30 | 20 | 30 | 60 | 10 |
| <i>Salmonella typhi</i> | 20 | 10 | 60 | 20 | 30 | 20 | 50 | 30 |
| <i>Shigella dysenteriae</i> | 30 | 10 | 40 | 40 | 20 | 10 | 50 | 50 |
| <i>Campylobacter jejuni</i> | 50 | 50 | 70 | 40 | 40 | 30 | 60 | 30 |
| <i>Enterobacter aerogenes</i> | 40 | 40 | 60 | 60 | 50 | 40 | 90 | 40 |

EE-Ethanol Extract; AE-Aqueous Extract; ME-Methanol Extract; Ofx- Ofloxacin.

Table 3. Synergistic activity of extracts of *Anisomeles malabarica* (L) (30µg/ml) with antibiotics (30µg/ml)

| | Zone of Inhibition (mm) | | | | | |
|-------------------------------|-------------------------|----|----|------|---|----|
| | Leaf | | | Wood | | |
| | E | O | EO | E | O | EO |
| <i>E. coli</i> | 7 | 10 | 13 | 4 | 7 | 10 |
| <i>Salmonella typhi</i> | 9 | 11 | 17 | 6 | 9 | 12 |
| <i>Shigella dysenteriae</i> | 10 | 14 | 19 | 5 | 8 | 11 |
| <i>Campylobacter jejuni</i> | - | 7 | 9 | - | 5 | 9 |
| <i>Enterobacter aerogenes</i> | - | - | 7 | - | 4 | 7 |

E-Extract; O-Ofloxacin; EO-Extract/Ofloxacin.

DISCUSSION

The MIC of the extracts ranged from 10-100 µg/mL, with the aqueous extract of leaf demonstrating the lowest values (MIC 10 µg/ml) against *Salmonella typhi* and *Shigella dysenteriae*, followed by the ethanolic extract against *E. coli* (MIC 20 µg/mL) and *Shigella dysenteriae* (MIC 30 µg/ml) (Table 2). The methanolic extract of *Anisomeles malabarica* also showed inhibitory activity for Gram-positive and negative bacteria Boobalan Raja and Jeganathan

A similar trend was observed with wood extract-ofloxacin combination against the bacterial species. At 30 µg/mL, both ofloxacin and the wood extracts had very less effect on *Enterobacter* and *Enteromonas*, but when combined, there was a remarkable activity (7 mm and 6 mm zone diameter of inhibition respectively). At 30 µg/mL the activity of the extracts and ofloxacin against *Salmonella typhi* were 6 and 9mm (zone diameter of inhibition), respectively but this increased to 12 mm when the extracts and the antibiotics were combined. Many reports are available on the antiviral, antibacterial,

antifungal, anthelmintic, antimolluscal and anti-inflammatory properties of plants Samy and Ignacimuthu, 2000; Palombo and Semple, 2001; Kumaraswamy, 2002; Stepanovic, 2003; Bylka, 2004; Behera and Misra, 2005; Govindarajan, 2006.

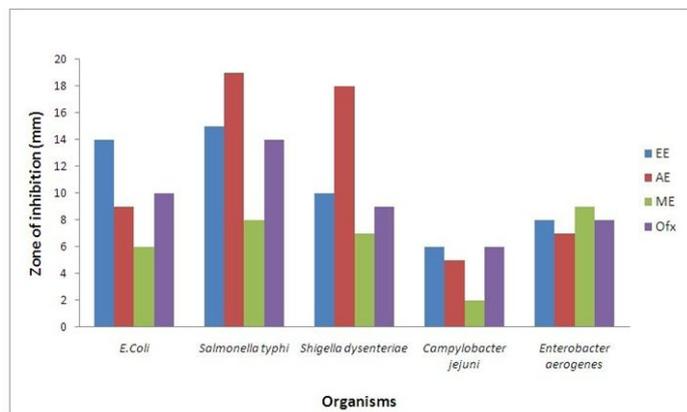


Figure 1. represents antibacterial activity of *Anisomeles malabarica* leaf extract

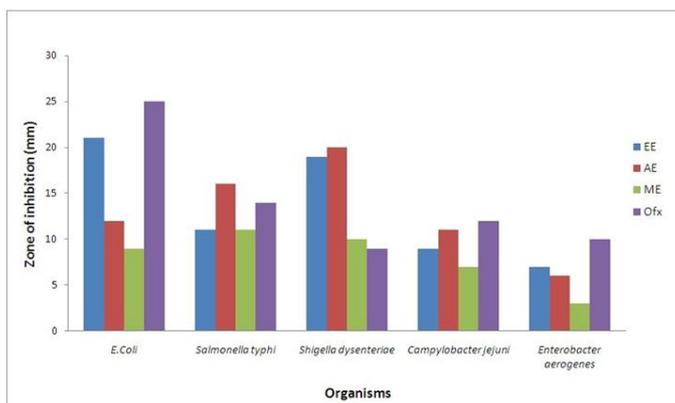


Figure 2. represents Antibacterial activity of *Anisomeles malabarica* wood extract

Conclusions

In recent times, there have been increases in antibiotic resistant strains of clinically important pathogens, which have led to the emergency of new bacterial strains that are multi-resistant. Plants are important source of potentially useful structures for the development of new chemotherapeutic agents. The use of natural products as antibacterials seems to be an interesting way to control the presence of pathogenic bacteria and to extend the shelf life of processed food. Extract showed varying of inhibitory effects. The plant-based, traditional medicine system continues to play an essential role in health care, with about 80% of the world's inhabitants relying mainly on traditional medicines for their primary health care. Thus, the study ascertains the value of *Anisomeles malabarica* plant used in ayurveda, which could be of considerable interest to the development of new drugs.

Acknowledgments

We are grateful to the Department of Biochemistry, Periyar University, Salem-11, Tamil Nadu, India, for their financial support.

REFERENCES

Adams JD, Frostick LE. Analysis of bacterial activity, biomass and diversity during windrow composting. *Waste Manag* 2009;29:598–605

Al-dabbas MM, Saganuma T, Kitahara K, Hou DX, Fujii M. Cytotoxic, antioxidant and antibacterial activities of *Varthemia*

iphionoides Boiss. extracts. *Journal of Ethnopharmacology* 2006;108:287–293.

Behera SK, Misra MK. Indigenous phytotherapy for genito-urinary diseases used by the Kandha tribe of Orissa, India. *J. Ethnopharmacol* 2005;102:319-325.

Berens S, Kaspari H, Klemme JH. Purification and characterization of two different xylanases from the thermophilic actinomycete *Microtetraspora flexuosa* SIIX. *Antonie Van Leeuwenhoek* 1996;69:235–241.

Bianco A, Uccella N. Biphenolic components of olives. *Food Res. Int* 2000; 33 (6):475–485.

Boobalan Raja, Jeganathan Manivann. *Journal of Pharmacy Research* 2010;3(6):1188-1191.

Bylka W, Szafer-Hajdrych M, Matalawskan I, Goslinka O. Antimicrobial activity of isocytoside and extracts of *Aquilegia vulgaris* L. *Lett. Appl. Microbiol* 2004;39:93-97.

Chiang LC, Chiang W, Liu MC, Lin CC. In vitro antiviral activities of *Caesalpinia pulcherrima* and its related flavonoids. *The Journal of Antimicrobial Chemotherapy* 2003;52:194–198.

Eloff JN. Which extractant should be used for the screening and isolation of antimicrobial components from plants? *J. Ethnopharmacol* 1998; 60:1–8.

Essawi T, Srour M. Screening of some Palestinian medicinal plants for antibacterial activity. *Journal of Ethnopharmacology* 2000; 70:343-349.

Fengel D, Wegener G. *Wood chemistry, ultrastructure, reactions.* de Gruyter, Berlin, 1984.

Govindarajan R, Vijayakumar M, Singh M, Rao CHV, Shirwaikar A, Rawat AKS, Pushpaganan P. Antiulcer and antimicrobial activity of *Anogeissus latifolia*. *J. Ethnopharmacol* 2006;106:57-61.

Holtz C, Kaspari H, Klemme JH. Production and properties of xylanases from thermophilic actinomycetes. *Antonie Van Leeuwenhoek* 1991;59:1–7.

Jeyachandran R, Mahesh, A, Cindrella L. DEN-induced cancer and its alleviation by *Anisomeles malabarica* (L.) R.Br. ethanol leaf extract in male albino mice. *Int J Cancer Res* 2007;3:174–9.

Kenney WA, Sennerby-Forsse L, Layton P. A review of biomass quality research relevant to the use of poplar and willow for energy conversion. *Biomass* 1990;21:163–188

Koduru S, Grierson DS, Afolayan, A.J. Antimicrobial activity of *Solanum aculeastrum*. *Pharm Biol* 2006; 44(4):283-286.

Kumaraswamy Y, Cox PJ, Jaspars M, Nahar L, Sarker SD. Screening seeds of Scottish plants for antibacterial activity. *J.Ethnopharmacol* 2002;83:73-77.

Little DP, Schwarzbach AE, Adams RP, Hsieh CF. The circumscription and phylogenetic relationships of *Callitropsis* and the newly described genus *Xanthocyparis* (Cupressaceae). *Amer. J. Bot* 2004;91:1872–1881.

Little DP. Evolution and circumscription of the true cypresses (Cupressaceae: Cupressus). *Syst. Bot* 2007;31:461–480.

Novinscak A, Decoste NJ, Surette C, Filion M. Characterization of bacterial and fungal communities in composted biosolids over a 2 year period using denaturing gradient gel electrophoresis. *Can J Microbiol* 2009;55:375–387.

Ozturk S, Ercisli S. The chemical composition of essential oil and in vitro antibacterial activities of essential oil and methanol. *Journal of Ethnopharmacology* 2006;106:372–376.

Palombo EA, Semple SJ. Antibacterial activity of traditional medicinal plants. *J. Ethnopharmacol* 2001;77:151-157.

Pankratov TA, Tindall BJ, Liesack W, Dedysh SN. *Mucilagini- bacter paludis* gen. nov., sp. nov. and *Mucilagini- bacter gracilissp.* nov., pectin-, xylan and laminarin-degrading members of the family Sphingobacteriaceae from acidic Sphagnum peat bog. *Int J Syst Evol Microbiol* 2007;57:2349–2354.

Perumalsamy R, Maung Thwin M, Gopalakrishnakone P, Ignacimuthu S. Ethnobotanical survey of folk plants for the treatment of snakebites in southern part of Tamilnadu, India. *Journal of Ethnopharmacology* 2008;115 (2): 302-312.

- Rennerfelt E, Nacht G. The fungicidal activity of some constituents from heartwood of conifers. *Sv. Bot. Tidskr* 1955;49:419–432.
- Rudman P. The causes of natural durability in timber. Part XI. Some tests on the fungi toxicity of wood extractives and related compounds. *Holzforschung* 1963;17:54–57.
- Samy RP, Ignacimuthu S. Antibacterial activity of some folklore medicinal plants used by tribals in Western Ghats in India. *J. Ethnopharmacol* 2000;69:63-71.
- Scheffer T, Cowling EB. Natural resistance of wood to microbial deterioration. *Annu. Rev. Phytopathol* 1966;4:147–170.
- Stepanovic S, Antic N, Dakic I, Svabic-vlahovic M. In vitro antimicrobial activity of propolis and antimicrobial drugs. *Microbiol. Res* 2003;158:353-357.
- Tomoko N, Takashi A, Hiromu T, Tetsuro I, Hiroko M, Munekazu I, Totshiyuki T, Tetsuro I, Fujio A, Iriya I, Tsutomu N, Kazuhito W. Antibacterial activity of extracts prepared from tropical and subtropical plants on methicillin-resistant staphylo-coccus. *J Health Sci* 2002; 48:273-276.
