



RESEARCH ARTICLE

ASSESSING STERILIZATION AND INFECTION CONTROL PRACTICES: THE CASE OF PRIVATE DENTAL PRACTITIONERS IN EAST DELHI REGION

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ABSTRACT

**Background:** Biosafety is a concern in all health sector services. Confronted with a high biological risk both for patients and professionals in dental care and due to the constant development of new technologies, information, equipment, material and behavioral attitudes in this area.

**Aim:** The aim of this study was to assess sterilization and infection control practices that are followed by private dental practitioners in East Delhi region.

**Methodology:** A descriptive, cross-sectional survey was conducted to assess the attitude towards and sterilization practices among the private dental practitioners in East Delhi, India. A structured proforma was used for the collection of the information by the investigator.

**Results:** A large proportion (68.0%) of the private dental practitioners carried out the pre-cleaning of the instruments before sterilization. Autoclave (66.0%) was the principle method of sterilization of the instruments among the private dental practitioners followed by Chemiclave (18.0%). Most (46.0%) of the private dental practitioners utilized the sterilized instruments within 1 week of sterilization. Autoclave was used more by the dental practitioners with more than 10 years of experience (35.7%) for sterilization of burs. Most (40.0%) of the private dental practitioners were not sterilizing endodontic files in their office. Most (52.0% and 66.0% respectively) of the private dental practitioners sterilized paper points and guttapercha respectively in their office.

**Conclusion:** To achieve sterilization, it is necessary to adhere strictly to procedures for instrument cleaning and wrapping and to follow carefully the sterilizer manufacturer's operating and maintenance instructions.

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INTRODUCTION

Biosafety is a concern in all health sector services. Confronted with a high biological risk both for patients and professionals in dental care and due to the constant development of new technologies, information, equipment, material and behavioral attitudes in this area,<sup>15,33,21</sup> health Organizations such as the Center for Disease Control (CDC), the American Dental Association (ADA), the National Sanitary Department (ANVISA) and the Ministry of Health (MOH) among others, have developed guidelines to prevent, minimize or eliminate any threat to life or health during treatment. This includes all instruments and equipment used, regardless of the confirmed or presumed diagnosis, being infectious or not.<sup>9,11</sup> The dental

clinic is an environment where disease transmission occurs easily.<sup>41</sup> Prevention of cross infection in the dental clinic is therefore a crucial aspect of dental practice, and dental clinic workers must adopt certain basic routines while practicing. Dental health care professionals (DHCPs) are at risk of infections caused by various microorganisms such as Mycobacterium tuberculosis, hepatitis B and hepatitis C viruses, staphylococci, streptococci, herpes simplex virus types 1, human immunodeficiency virus (HIV), mumps, influenza, and rubella.<sup>16,38</sup> The environment in dentistry practices and clinics is far from ideal.<sup>23</sup> However, the adoption of infection control measures is an effective way to reduce occupational risk and the transmission of pathogens, mainly through saliva, blood, air or water. These measures essentially include (I) cleaning, disinfection and sterilization; (II) the use of personal equipment protection; (III) immunization; (IV) prevention and correct handling in occupational accidents which involve exposure to blood and bodily fluids; and (V) antisepsis.<sup>5,11,30</sup>

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Disinfection and sterilization are essential for ensuring that medical and surgical instruments do not transmit infectious pathogens to patients. Because sterilization of all patient-care items is not necessary, health-care policies must identify, primarily on the basis of the items' intended use, whether cleaning, disinfection, or sterilization is indicated.<sup>23,39,42,47</sup>

The American Dental Association recommends that all instruments, burs, mirrors, bands, and other devices used in intraoral treatments be sterilized by one of these four methods: Steam autoclaves, Dry heat ovens, Unsaturated chemical vapor sterilizers, and Ethylene oxide gas sterilizers. These four types of sterilization can be easily controlled and their effectiveness verified during use in the dental office.<sup>2,13,14</sup> However, the factors that affect the efficacy of both disinfection and sterilization include prior cleaning of the object; organic and inorganic load present; type and level of microbial contamination; concentration of and exposure time to the germicide; physical nature of the object (e.g., crevices, hinges, and lumens); presence of biofilms; temperature and pH of the disinfection process; and in some cases, relative humidity of the sterilization process (e.g., ethylene oxide). Since the occurrence of most sterilization problems is unpredictable, it is necessary to routinely monitor equipment performance. Physical observations and internal and external chemical monitors can provide immediate results. Unfortunately, such methods can measure only one parameter, such as temperature or the presence of a specific gas. They cannot assess all of the physical factors that must work in concert so that sterilization can be achieved. The main guarantee of sterilization is the successful killing of a bacterial endospore sample.<sup>1, 14,31</sup> Multiple studies in many countries have documented lack of compliance with established guidelines for disinfection and sterilization.<sup>23,39,42,47</sup> Failure to comply with scientifically-based guidelines has led to numerous outbreaks.<sup>12,20,25,26,39,44,45</sup> Therefore, the present study was conducted to assess the attitude and practices of the private dental practitioners in India towards the sterilization of the instruments and equipments and suggest suitable recommendations in the light of the current scenario.

## MATERIALS AND METHODS

A descriptive, cross-sectional survey was conducted to assess the attitude towards and sterilization practices among the private dental practitioners in East Delhi, India. A structured proforma was used for the collection of the information by the investigator.

**Ethical clearance:** The permission to carry out the study was obtained from the Institutional Committee of the Santosh Dental College, Ghaziabad. Written informed consent was obtained from all participants. The present research was conducted in full accordance with the World Medical Association Declaration of Helsinki.

**Study proforma:** The proforma included information such as demographic factors, including gender, age, and year of study. As, there was no standard proforma available, a proforma was framed with the help of experts in the field. The questions assessed the attitude and practices regarding the sterilization of the instruments and equipments among the private dental

practitioners. The proforma kept the study group in mind, and questions were linked to the sterilization practices. A self-administered proforma consisting of fourteen close-ended items was used for data collection.

**Study sample:** The study population consisted of private dental practitioners in East Delhi. A total of 300 dentists were delivered the survey questionnaire, of which 200 completed the whole questionnaire. The over-all response rate among these respondents was 70%.

**Statistical Analysis:** The data were tabulated and entered into Microsoft Excel 2010 and analysed using the SPSS software 16.0. The descriptive analysis of the responses was carried out and the results were expressed in the form of frequencies and percentages. The chi-square test was used for comparison of the categorical variables.

## RESULTS

All the private dental practitioners in the present study were concerned about the risk of cross-infection to themselves and their assistants. Majority (62.0%) of the dental practitioners did not sterilize their instruments themselves in their office and were sterilized by their assistants only. Majority (68.0%) of the dental assistants did not have any formal training in the sterilization and infection control practices and were trained in practice only. However, there was no difference in these practices among the dental practitioners with less than 10 years and more than 10 years of practice. (**Table 1**) A large proportion (68.0%) of the private dental practitioners carried out the pre-cleaning of the instruments before sterilization. Mostly, the pre-cleaning of the instruments was carried out either by washing with soap and water (29.4%) or washing with water only (23.5%) with 47.1% using an ultrasonic cleaner for the pre-cleaning of the instruments. There was not any significant difference in the pre-cleaning of the instruments as per the practice years. (**Table 2**) Surgical cloth (42.0%) was used most frequently by the private practitioners for the packaging of the instruments before sterilization. The private dental practitioners with experience more than 10 years were using both Plastic Pouches and Surgical cloth (50.0%) whereas the practitioners with less than 10 years of practice were using only surgical cloth (46.4%). There was a statistically significant ( $p < 0.05$ ) difference in the method of packaging of the instruments before keeping them for sterilization among the private dental practitioners. (**Table 3**) Autoclave (66.0%) was the principle method of sterilization of the instruments among the private dental practitioners followed by Chemiclave (18.0%).

There was a statistically significant ( $p < 0.05$ ) difference among the private dental practitioners in sterilization practices for the sterilization of the instruments. Autoclave (72.7%) and Dry heat (9.1%) followed significantly more frequently by the private dental practitioners with more than 10 years of practice. Boiling (14.3%) and Chemiclave (25.0%) were used significantly more by the private dental practitioners with less than 10 years of practice. (**Table 3**) Most (46.0%) of the private dental practitioners utilized the sterilized instruments within 1 week of sterilization.

**Table 1. Attitude towards the sterilization among the private dental practitioners**

	Response	Experience			p-value
		Less than 10 years	More than 10 years	Total	
Question 1: Are you concerned about the risk of cross-infection to yourselves and your dental assistants	Yes	56 100.0%	44 100.0%	100 100.0%	1.000 <sup>#</sup>
	Total	56 100.0%	44 100.0%	100 100.0%	
Question 2: Do you sterilize the instruments personally in your office	Yes	24 42.9%	14 31.8%	38 38.0%	0.259 <sup>#</sup>
	No	32 57.1%	30 68.2%	62 62.0%	
	Total	56 100.0%	44 100.0%	100 100.0%	
Question 3: Did your staff receive any formal training in the sterilization and infection control practices	Yes	18 32.1%	14 31.8%	32 32.0%	0.972 <sup>#</sup>
	No	38 67.9%	30 68.2%	68 68.0%	
	Total	56 100.0%	44 100.0%	100 100.0%	

Chi-square test

<sup>#</sup>Non-significant difference (p-value>0.05)**Table 2. Practice of Pre-cleaning of instruments among the private dental practitioners**

	Response	Experience			p-value
		Less than 10 years	More than 10 years	Total	
Question 4: Do you preclean your instruments before sterilization?	Yes	40 71.4%	28 63.6%	68 68.0%	0.407 <sup>#</sup>
	No	16 28.6%	16 36.4%	32 32.0%	
	Total	56 100.0%	44 100.0%	100 100.0%	
Question 5: How the pre-cleaning is carried out?	Ultrasonic	20 50.0%	12 42.9%	32 47.1%	0.703 <sup>#</sup>
	Washing with soap and water	12 30.0%	8 28.6%	20 29.4%	
	Washing with water only	8 20.0%	8 28.6%	16 23.5%	
	Total	40 100.0%	28 100.0%	68 100.0%	

Chi-square test

<sup>#</sup>Non-significant difference (p-value>0.05)**Table 3. Sterilization practices for the instruments among the private dental practitioners**

	Response	Experience			p-value
		Less than 10 years	More than 10 years	Total	
Question 6: How do you wrap/ pack your instruments before keeping them for sterilization	Plastic Pouches	16 28.6%	6 13.6%	22 22.0%	0.025*
	Surgical Cloth	26 46.4%	16 36.4%	42 42.0%	
	Plastic Pouches, Surgical cloth	14 25.0%	22 50.0%	36 36.0%	
	Total	56 100.0%	44 100.0%	100 100.0%	
Question 7: What is the preferred method for sterilization in your office?	Autoclave	34 60.7%	32 72.7%	66 66.0%	0.022*
	Boiling	8 14.3%	4 9.1%	12 12.0%	
	Chemoclave	14 25.0%	4 9.1%	18 18.0%	
	Dry heat	0 .0%	4 9.1%	4 4.0%	
	Total	56 100.0%	44 100.0%	100 100.0%	
Question 8: Preferred time of use of sterilized, wrapped or packed instruments	1 week	22 39.3%	24 54.5%	46 46.0%	0.033*
	4 weeks	12 21.4%	8 18.2%	20 20.0%	
	6 weeks	14 25.0%	4 9.1%	18 18.0%	
	12 weeks	0 .0%	4 9.1%	4 4.0%	
	More than 12 weeks	8 14.3%	4 9.1%	12 12.0%	
	Total	56 100.0%	44 100.0%	100 100.0%	

Chi-square test

\* Significant difference (p-value≤0.05)

Table 4. Sterilization practices of the burs and handpiece among the private dental practitioners

	Response	Experience		Total	p-value
		Less than 10 years	More than 10 years		
<b>Question 9: How do use sterilize burs in your clinic</b>	Autoclave	20	8	28	0.293 <sup>#</sup>
		35.7%	18.2%	28.0%	
	Chemical vapour	2	2	4	
		3.6%	4.5%	4.0%	
	Dip it in surgical spirit	6	10	16	
		10.7%	22.7%	16.0%	
	Dry heat	6	2	8	
		10.7%	4.5%	8.0%	
<b>Question 10: How do you sterilize handpieces between patients?</b>	Glass bead sterilizer	6	6	12	0.001***
		10.7%	13.6%	12.0%	
	No	16	16	18	
		28.6%	36.4%	18.0%	
	Total	56	44	100	
		100.0%	100.0%	100.0%	
	Autoclave	6	16	22	
		10.7%	36.4%	22.0%	
	Autoclave, Chemicalve	14	8	22	0.001***
		25.0%	18.2%	22.0%	
	Run it in a disinfectant solution	2	6	8	
		3.6%	13.6%	8.0%	
	Wipe it with surgical spirit	24	6	30	
		42.9%	13.6%	30.0%	
	No	10	8	18	
		17.9%	18.2%	18.0%	
Total	56	44	100		
	100.0%	100.0%	100.0%		

Chi-square test

\*\*\* Very Highly Significant difference (p-value≤0.05)

Table 5. Sterilization of endodontic files, paper points and guttapercha among study population

	Response	Experience		Total	p-value
		Less than 10 years	More than 10 years		
<b>Question 11: How do you sterilize endodontic files in your office?</b>	Autoclave	8	2	10	0.101 <sup>#</sup>
		14.3%	4.5%	10.0%	
	Chemiclave	6	4	10	
		10.7%	9.1%	10.0%	
	Dip in a disinfectant solution	12	10	22	
		21.4%	22.7%	22.0%	
	Glass bead sterilizer	8	10	18	
		14.3%	22.7%	18.0%	
<b>Question 12: Do you sterilize the paper points before use in your clinic</b>	Don't sterilize	22	18	40	0.246 <sup>#</sup>
		46.5%	40.9%	40.0%	
	Total	56	44	100	
		100.0%	100.0%	100.0%	
	No	32	20	52	
		57.1%	45.5%	52.0%	
	Yes	24	24	48	
		42.9%	54.5%	48.0%	
<b>Question 13: Do you sterilize Guttapercha points before use in your clinic</b>	Total	56	44	100	0.683 <sup>#</sup>
		100.0%	100.0%	100.0%	
	No	20	14	34	
		35.7%	31.8%	34.0%	
	Yes	36	30	66	
		64.3%	68.2%	66.0%	
	Total	56	44	100	
		100.0%	100.0%	100.0%	
<b>Question 14: How do you sterilize Guttapercha points before use in your clinic</b>	Ethyl alcohol	6	14	20	0.024*
		16.7%	46.7%	30.3%	
	Glass bead sterilizer	4	2	6	
		11.1%	6.7%	9.1%	
	Gluteraldehyde	12	2	14	
		33.3%	6.7%	21.2%	
	Gluteraldehyde, Ethyl alcohol	4	2	6	
		11.1%	6.7%	9.1%	
	Sodium hypochlorite	10	10	20	0.024*
		27.8%	33.3%	30.3%	
	Total	36	30	66	
		100.0%	100.0%	100.0%	

Chi-square test

<sup>#</sup>Non-Significant difference (p-value>0.05)

\* Significant difference (p-value≤0.05)

The private dental practitioners with experience more than 10 years used the sterilized instruments in 1 week only (54.5%) whereas dental practitioners with experience less than 10 years used the sterilized instruments in 6 weeks (25.0%) and more than 12 weeks (14.3%). There was a significant difference ( $p < 0.05$ ) in the time of use of sterilized instruments among dental practitioners with more and less than 10 years of experience. (Table 3) Autoclave was used more by the dental practitioners with more than 10 years of experience (35.7%) for sterilization of burs but as such there was no significant difference ( $p > 0.05$ ) in the sterilization of burs by the private dental practitioners with different levels of experience. Most of the dental practitioners with more than 10 years of experience were using Autoclave for sterilization of the handpiece whereas the dental practitioners with less than 10 years of experience were wiping it with surgical spirit before use. There was a significant ( $p < 0.05$ ) difference in the method of sterilization of the handpiece. (Table 4)

Most (40.0%) of the private dental practitioners were not sterilizing endodontic files in their office. Most (52.0% and 66.0% respectively) of the private dental practitioners sterilized paper points and guttaperchare spectively in their office. As such, there was no difference in the sterilization practices for the endodontic files, paper points and guttapercha among the dental practitioners with less and more than 10 years of experience. Glutraldehyde was used significantly ( $p < 0.05$ ) more frequently by the private dental practitioners with less than 10 years of experience (21.4%) whereas Ethyl alcohol significantly ( $p < 0.05$ ) more frequently by the private dental practitioners with more than 10 years of experience (31.8%). (Table 5)

## DISCUSSION

Due to the nature of their profession, dentists and dental assistants should not forget the risk of treating patients with probability of infectious diseases. Dentists, dental assistants and patients may be exposed to pathogenic microorganisms localized in oral cavity and respiratory tract including cytomegalovirus (CMV), HBV, HCV, herpes simplex virus (HSV) type 1 and 2, HIV, Mycobacterium tuberculosis, staphylococci, streptococci and other viruses and bacteria.<sup>6</sup> These microorganisms could be transmitted to the dental healthcare professionals by direct contact with a patient's saliva, blood, skin, and oral secretions, or by indirect contact through injuries caused by sharp contaminated instruments, or by droplet infection from aerosols or spatter.<sup>6,10,43</sup>

All the private dental practitioners in the present study were concerned about the risk of cross-infection to themselves and their assistants. This was much higher than the study by Yüzbaşıoğlu *et al.*,<sup>46</sup> in which, 74.10% of the dentists expressed concern about the risk of cross-infection to themselves and their dental assistants. The role of the dental assistant is vital to the process of infection control and sterilization; however, the adherence of this particular group to these guidelines is inadequate because they receive less formal training than provided for dentists.<sup>3</sup> Any infection control training the dental assistants receive is provided by the dentists who employ dental assistants.

Formal training and certification is equally necessary for the dental surgery assistants as for the other members of the dental team to ensure a sound knowledge understanding of all aspects of infection control in the dental setting. The training of the dental assistants is less rigorous in United Kingdom but however, it is more rigorous in unites states.<sup>16</sup> Prior disinfection is characterized by soaking the instruments contaminated by organic material in chemical disinfectant solutions, before cleaning, in order to reduce risks of pathogen exposures to the professional. However, research shows that this practice is not based on scientific evidence, as the organic material can interfere in the antimicrobial activity of disinfectants.<sup>17,34,35</sup> A large proportion (68.0%) of the private dental practitioners carried out the pre-cleaning of the instruments before sterilization mostly, either by washing with soap and water (29.4%) or washing with water only (23.5%) with 47.1% used an ultrasonic cleaner. The study by Bellissimo-Rodrigues *et al.*<sup>4</sup> reported that 75.0% of private and 93.5% of public dentists reported an adequate procedure ( $p = 0.047$ ), which means they were cleaned soon after being used or at the end of the day, after being immersed in non-antimicrobial liquid soap.<sup>7,11,40</sup> Alternatively, others allowed the dirty instruments to dry or submerged the devices in germicides, such as glutaraldehyde, bleach, hydrogen peroxide, ammonium quaternary compounds or formaldehyde. Use of germicides before cleaning is not recommended because disinfection is not achieved in the presence of organic material and may even promote adhesion of organic substances to the device, compromising subsequent cleaning. Additionally, it may pose hazards to health-care workers.<sup>7,11</sup> The private dental practitioners with experience more than 10 years were using both Plastic Pouches and Surgical cloth (50.0%) whereas the practitioners with less than 10 years of practice were using only surgical cloth (46.4%). Private dentists in our study were not found to be having adequate packing practice for packaging of the instruments. Packing devices prior to sterilisation is also relevant to achieve and maintain sterilization until the device is to be used.<sup>7,8,11</sup> The study by Bellissimo-Rodrigues *et al.*<sup>4</sup> reported that the adequate packing for dry heat sterilization was accomplished by 33.3% of private but none of the public dentists ( $p < 0.001$ ). For autoclave sterilization, adequate packing was accomplished by only 20.8% of private dentists and none of the public ones ( $p = 1.000$ ).

In the present study, Autoclave (66.0%) was the principle method of sterilization of the instruments among the private dental practitioners followed by Chemiclave (18.0%). Autoclave (72.7%) and Dry heat (9.1%) followed significantly ( $p < 0.05$ ) more frequently by the private dental practitioners with more than 10 years of practice. Boiling (14.3%) and Chemiclave (25.0%) were used significantly ( $p < 0.05$ ) more by the private dental practitioners with less than 10 years of practice. Asymptomatic patients, whether carriers of infectious diseases or not, are assisted by dental surgeons every day, making sterilization processes and validation of paramount importance. In the study by Matsuda *et al.*,<sup>22</sup> autoclave (moist-heat by steam), oven (dry-heat sterilizer), chemical solutions and alcohol were analyzed as methods of "sterilization" with autoclave being the preferred method of sterilization.

Autoclave is considered the preferred method due to its safety, quickness and its lethal effect of pressurized steam on all microorganisms.<sup>5,9,17</sup> Nonetheless, recent studies have shown that 12% to 33% of these devices present defects that are easily detectable with periodical monitoring of the cycles and the simultaneous use of different sterilization indicators.<sup>32</sup> Findings by Monarca *et al*<sup>28</sup> show 68.60% of autoclave users with 27.40% referring no use of indicators. Similar results were observed in the study by Matsuda *et al*,<sup>22</sup> where 69.38% of participants used autoclave and 33.80% of these did not use indicators. Daily use or at least weekly use of indicators, as well as the combined use of different indicators, were ignored by 81.75% and 83.21%, respectively. The free use of material and/or instruments with no certification of correct sterilization is a large concern in respect to biosafety. The literature describes the oven as a secure method for sterilization, but it is less appropriate than the autoclave, mainly as it permits the interruption of process, by the heterogeneity of penetration and heat distribution inside the chamber, by the absence of a precision thermostat to effectively control temperature and because it requires prolonged exposure to high temperatures.<sup>36</sup> Due to these issues, sterilization in a dry-heat sterilizer which seemed to be the practice of approximately 10% of the participants in our research is currently recommended just for metal blades, points, cutting or drill instruments sensitive to oxidation by steam.<sup>37</sup> Chemical solutions are also referred to as sterilizers provided their concentration and length of exposure.<sup>5</sup> They are also considered toxic and irritant, with limited action and effectiveness. Therefore they are restricted to thermo sensitive material and are used as a last resource for sterilization, on scientific grounds.<sup>17</sup> A large proportion (30.0%) of the private dental practitioners were sterilizing their handpieces by wiping them with the surgical spirit followed by Autoclave (22.0%) and a combination of Autoclave and Chemical (22.0%). In the study by Bellissimo-Rodrigues *et al*,<sup>4</sup> 80% of dentists preferred to clean hand pieces by wiping them with disinfectants, but only 17.8% of them stated that they preferred autoclave for sterilize handpieces. However, it is known live blood cells and bacterial and viral particles can survive inside handpieces even after thorough disinfection.<sup>19</sup> Many authors have emphasized the hazard of cross-infection by the use of dental instruments.<sup>27</sup> Some of these authors showed that 94% of dentists in Kuwait used autoclave to sterilize handpieces.<sup>29</sup> Kurdy and Fontaine<sup>18</sup> showed that 30% of dentists in Saudi Arabia had sterilized hand pieces with autoclave and 90% of them autoclaved their instruments at the end the day. Al-Rabeah and Mohamed<sup>3</sup> stated that 37.90% of dentists autoclaved handpieces. According to Miller,<sup>27</sup> the most common reason for not sterilizing hand pieces is the fear of damage to the equipment.

## Conclusion

The sterilization process is a combination of a variety of procedures, which include both human and material factors. It would have been interesting if this study had determined the critical importance of one or two practice or personal elements. However, the inability to establish such identifications only reinforces the known complexity of the sterilization process. To achieve sterilization, it is necessary to adhere strictly to procedures for instrument cleaning and wrapping and to follow

carefully the sterilizer manufacturer's operating and maintenance instructions.

## REFERENCES

1. AHA Committee on Infection within Hospitals. 1974. Infection control in the hospital. 3rd ed. Chicago: *Amedcan Hospital Association*, 97-8.
2. Al-Rabeah A & Mohamed AGI. 2002. Infection control in the private dental sector in Riyadh. *Ann Saudi Med.*, 22:1-2.
3. Amedcan Dental Association. 1983. Dentists' desk reference: materials, instruments and equipment. 2nd ed. Chicago: *Amedcan Dental Association*, 390-401.
4. Bellissimo-Rodrigues WT, Bellissimo-Rodrigues F, Machado AA. 2009. Infection control practices among a cohort of Brazilian dentists. *Int Dent J.*, 59(1):53-8.
5. Block SS. 2000. Disinfection, Sterilization, and Preservation. 5<sup>a</sup> ed. Philadelphia: Lippincott Williams & Wilkins. 1504 pp.
6. Bolyard EA, Tablan OC, Williams WW, Pearson ML, Shapiro CN, Deitchman SD. 1998. Guideline for infection control in health care personnel. Hospital Infection Control Practices Advisory Committee. *Am J Infect Control*, 26:289-354.
7. Brasil. Ministério da Saúde. Agência Nacional de Vigilância Sanitária. Serviços Odontológicos: Prevenção e Controle de Riscos. 2<sup>a</sup> ed. Brasília: ANVISA; (2006). 152 pp.
8. Brasil. Ministry of Health. National Agency of Sanitary Surveillance. Serviços odontológicos: prevenção e controle de riscos. Brasília 2006. 156 p. [Cited 2015 June 12] Available on line from: [https://www.anvisa.gov.br/servicosauade/manuais/manual\\_odonto.pdf](https://www.anvisa.gov.br/servicosauade/manuais/manual_odonto.pdf).
9. Brasil. Ministry of Health. Secretaria de Políticas de Saúde. Coordenação Nacional de DST e Aids. 2000. Controle de infecções e a prática odontológica em tempos de Aids: manual de condutas. Brasília, 118 p.
10. Centers for Disease Control and Prevention 2005. Pseudomonas aeruginosa infections associated with transrectal ultrasound-guided prostate biopsies—Georgia, (2006) MMWR CDC Surveill Summ 55:776-7.
11. Centers for Disease Control and Prevention. 1993. Recommended infection control practices for dentistry. MMWR Morbid Mortal Wkly Rep 42:1-12.
12. Centers for Disease Control and Prevention. *Guidelines for Infection Control in Dental Health-Care Settings* 2003. MMWR 2003;52(RR-17):1-66.
13. Council on Dental Materials and Devices, Council on Dental Therapeutics 1978. Infection control in the dental office. *J Am Dent Assoc* 97:671-3.
14. Council on Dental Materials, Instruments and Equipment. 1981. Current status of sterilization of instruments, devices and methods for the dental office. *J Am Dent Assoc* 102:683-9.
15. Garbin AJI, Garbin CAD, Arcieri RM, Crossato M, Ferreira NF 2005. Biosecurity in public and private office. *J Appl Oral Sci.*, 13(2):163-6.
16. Gordon BL, Burke FJ, Bagg J, Marlborough HS, McHugh ES. 2001. Systematic review of adherence to infection control guidelines in dentistry. *J Dent*, 29(8):509-16.

17. Graziano KU, Manrinque EI, Fernandes AT *et al.* 2000. Controle de infecção na prática odontológica. São Paulo: Associação Paulista de Estudos e Controle de Infecção Hospitalar (APECIH); 87 pp.
18. Kurdy S & Fontaine RE. 1997. Survey on infection control in MOH dental clinics. *Riyadh Saudi Epidemiol Bull*, (3,4):21-8.
19. Lewis DL, Arens M, Appleton SS, Nakashima K, Ryu J, Boe RK, *et al.* 1992. Cross-contamination potential with dental equipment. *Lancet*, 340:1252-4.
20. Lowry PW, Jarvis WR, Oberle AD *et al.* 1988. Mycobacterium chelonae causing otitis media in an ear-nose-and-throat practice. *N Engl J Med.*, 319:978-82.
21. Machado GL & Kather JM. 2002. Estudo do controle da infecção cruzada pelos cirurgiões-dentistas de Taubaté. *Rev Biociênc* 8(1):37-44.
22. Matsuda JK, Grinbaum RS, Davidowicz H. 2011. The assessment of infection control in dental practices in the municipality of São Paulo. *Braz J Infect Dis.*, 15(1):45-51.
23. McCarthy GM, Koval JJ, John MA, MacDonald JK. 1999. Infection control practices across Canada: do dentists follow the recommendations? *J Can Dent Assoc.*, 65:506-11.
24. Medeiros UV, Souza MIC, Bastos LF. 2003. Odontologia do trabalho: riscos ocupacionais do cirurgião-dentista. *Rev Bras Odontol.*, 60(4):277-80.
25. Mehta AC, Prakash UBS, Garland R *et al.* 2006. Prevention of flexible bronchoscopy-associated infection. *Chest*, 128: 1742-55.
26. Meyers H, Brown-Elliott BA, Moore D *et al.* 2002. An outbreak of Mycobacterium chelonae infection following liposuction. *Clin Infect Dis.*, 34:1500-7.
27. Miller CH. 1991. Sterilization: disciplined microbial control *Dent Clin North Am.*, 35:339-55.
28. Monarca S, Grottolo M, Renzi D *et al.* 2000. Evaluation of environmental bacterial contamination and procedures to control cross infection in a sample of Italian dental surgeries. *Occup Environ Med.*, 57(11):721-6.
29. Morris E, Hassan FS, Al Nafisi A, Sugathan TN. 1996. Infection control knowledge and practices in Kuwait: a survey on oral health care workers. *Saudi Dent J.*, 8:19-26.
30. Naressi SCM, Akama CM, Silva LMP, Siviero M. 2004. Análise das formas de esterilização e do meio de controle empregados pelos cirurgiões-dentistas de São José dos Campos - SP. *Rev Odontol UNESP* 33(4):169-74.
31. Palenik C J, Miller CH, Spandau DF. 1981. Monitoring the dental office sterilizer. *J Indiana Dent Assoc.*, 60:25-7.
32. Patiño-Marín N, Loyoa-Rodríguez JP, Tovar Reyes LF. 2001. Uso y verificación cónin dicadores biológico e seme sterilizadores de cirujanos dentistas de San Luis Potosí, México. *Salud Publica Mex*, 43(5):455-8.
33. Rosa MRD, Albuquerque SSL, Júnior AAV, Coelho AEL. 2001. Comportamento dos Cirurgiões-Dentistas em Relação à Utilização do Equipamento de Proteção Individual (EPI) no Controle de Infecções. *Rev Bras CiêncSaúde*, 5(2):125-30.
34. São Paulo (Estado). Secretaria de Estado da Saúde. Resolução SS- 27, de 28/02/2007. Aprova Norma Técnica que institui medidas de controle sobre o uso do Glutaraldeído nos Estabelecimentos Assistenciais de Saúde. Diário Oficial do Estado, São Paulo, nº 73 de 12/06/2015.
35. São Paulo (Estado). Secretaria de Estado da Saúde. Resolução SS-374, de 15/12/1995. Altera a Norma Técnica sobre reorganização do Centro de Material e Noções de Esterilização. Diário Oficial do Estado, São Paulo, 16/12/1995.
36. Silva ASF, Ribeiro MC, Risso M. 2009. Biossegurança em Odontologia e Ambientes de Saúde. 2º ed. São Paulo: Ícone; 262 pp.
37. Sindicato dos Odontologistas do Estado de São Paulo. 2009. *Manual do Cirurgião-Dentista*. São Paulo: SOESP; 22 pp.
38. Singh A, Purohit BM, Bhambal A, Saxena S, Singh A, Gupta A. 2011. Knowledge, Attitudes, and Practice Regarding Infection Control Measures Among Dental Students in Central India. *J Dent Educ.*, 75(3):421-7.
39. Spach DH, Silverstein FE, Stamm WE. 1993. Transmission of infection by gastrointestinal endoscopy and bronchoscopy. *Ann Intern Med.*, 118:117-28.
40. Spaulding EH. 1968. Chemical disinfection of medical and surgical materials. In Lawrence CA, Block, SS (ed). *Disinfection sterilization and preservation*. 4th.ed, pp 517-531. Philadelphia: Lea & Febiger
41. Taiwo JO & Aderinokun GA. 2002. Assessing cross infection prevention measures at the dental clinic, University College Hospital, Ibadan. *Afr J Med Sci.*, 31(3):213-7
42. Uttley AH & Simpson RA. 1994. Audit of bronchoscope disinfection: a survey of procedures in England and Wales and incidents of mycobacterial contamination. *J Hosp Infect* 26:301-8.
43. Verrusio AC, Neidle EA, Nash KD, Silverman S Jr, Horowitz AM, Wagner KS. 1989. The dentist and infectious diseases: a national survey of attitudes and behavior. *J Am Dent Assoc.*, 118:553-62.
44. Weber DJ & Rutala WA. 2001. Lessons from outbreaks associated with bronchoscopy. *Infect. Control Hosp Epidemiol.*, 22:403-8.
45. Weber DJ, Rutala WA, DiMarino AJ, Jr. 2002. The prevention of infection following gastrointestinal endoscopy: the importance of prophylaxis and reprocessing. In: DiMarino AJ, Jr, Benjamin SB, eds. *Gastrointestinal diseases: an endoscopic approach*. Thorofare, NJ: Slack Inc. 87-106.
46. Yüzbasioglu E, Saraç D, Canbaz S, Saraç YS, Cengiz S. 2009. A survey of cross-infection control procedures: knowledge and attitudes of Turkish dentists. *J Appl Oral Sci.*, 17(6):565-9.
47. Zaidi M, Angulo M, Sifuentes-Osornio J. 1995. Disinfection and sterilization practices in Mexico. *J Hosp Infect*, 31:25-32.

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